APPENDIX G

Aquatic Ecology and Water Quality Impact Assessment (frc environmental)



Six Mile Creek Dam Safety Upgrade Project:

Aquatic Ecology and Water Quality Impact Assessment

Prepared for:

SMEC on behalf of Seqwater

frc environmental

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Summary

frc environmental was commissioned by SMEC on behalf of Seqwater to undertake an assessment of potential risks to the aquatic ecological values of Six Mile Creek, and to identify impact mitigation strategies for the Six Mile Creek Dam Safety Upgrade Project (the Project). The scope of the study involved:

- · describing the aquatic Environmental Values of Six Mile Creek and Lake Macdonald
- · reviewing the legislative and policy framework for protecting aquatic ecological values in Queensland
- · identifying potential sources of impact to the aquatic ecological values associated with the Project and assessing the level of risk to aquatic ecological values
- · identifying likely impact mitigation strategies and assessing the effectiveness of each strategy, and
- providing recommendations regarding preferred impact mitigation strategies and other management controls to protect the aquatic ecological values of Six Mile Creek.

Six Mile Creek is a large tributary of the Mary River, originating inland from Noosa Heads and flowing for approximately 60 km north-west to join the Mary River approximately 4.5 km south of Gympie. Six Mile Creek Dam is in the upper reaches of Six Mile Creek at ATMD 55 km. The existing dam has a capacity of 8,018 megalitres at a Full Supply Level of 95.32 m, which will not change after the completion of the Project. The existing dam does not incorporate a fishway.

The aquatic ecological values of Six Mile Creek and Six Mile Creek Dam were assessed using a literature and database review, synthesis of existing data for aquatic habitat, water quality, aquatic plants, fish and turtles (including from past field surveys), field survey, and consultation with key experts. Overall, aquatic ecology and water quality data was assessed at 13 sites on Six Mile Creek and two sites on Lake Macdonald. Water quality data collected by Seqwater at an additional four sites was also assessed. Platypus and stygofauna were also assessed using a desktop approach.

Two threatened fish and two freshwater turtle species listed under the *Environment Protection and Biodiversity Act 1994* (i.e. aquatic Matters of National Environmental Significance) are known from the Mary River and Six Mile Creek:

- white-throated snapping turtle (*Elseya albagula*), critically endangered
- Mary River cod (*Maccullochella mariensis*), endangered

- Mary River turtle (*Elusor macrurus*), endangered, and
- Australian lungfish (*Neoceratodus forsteri*), vulnerable.

Mary River cod and Australian lungfish are known from Six Mile Creek downstream of Lake Macdonald, and it is possible that Mary River turtle and white-throated snapping turtle sometimes occur in the lower reaches of Six Mile Creek. Mary River cod are known to breed in Six Mile Creek downstream of Lake Macdonald. Within Lake Macdonald, and upstream of Lake Macdonald, platypus are known to occur, Mary River cod and Australian lungfish may occur but are unlikely to be breeding, and Mary River turtle or white-throated snapping turtle are likely to be rare or absent. Matters of State Environmental Significance downstream and upstream of Lake Macdonald (other than fish passage) include several categories of regulated vegetation and High Ecological Value (watercourse) waters.

The desktop assessment indicated that the alluvium of the Project area is *unlikely to be suitable* for stygofauna due to high clay content, low hydraulic conductivity and high total dissolved solids, and the Kin Kin sandstone is *suitable* for stygofauna due to the higher hydraulic conductivity.

The following sources of potential impact from the Project to the aquatic Environmental Values of Six Mile Creek and Lake Macdonald were identified:

- · Impacts to water quality in Lake Macdonald and downstream of the lake
- · Impacts to aquatic habitat in Lake Macdonald and downstream
- · Impacts to aquatic fauna (injury, mortality or stranding) in Lake Macdonald
- · Impacts to aquatic flora in Lake Macdonald and downstream of the lake
- · Spread of biosecurity matters downstream of Lake Macdonald
- · Impacts to stygofauna communities in shallow groundwater systems, and
- · Barriers to fish passage at the dam wall.

The risks of, and mitigations for, each of the identified potential sources of adverse impact of the Project on the Environmental Values of Six Mile Creek were assessed using a riskbased approach.

Potential direct and indirect impacts of the Project are likely to affect both Lake Macdonald and Six Mile Creek downstream. As the Project is replacing an existing dam wall the longterm impact will be no change from current condition. Most sources of impact during the Project were assessed as having a low risk of impact when appropriate mitigations are applied. Most potential impacts will be temporary (i.e. for the duration of the drawdown and construction periods; approximately 2 to 3 years in total), and if appropriate mitigations are applied there will be no ongoing impacts to the aquatic environmental values of Lake Macdonald or Six Mile Creek.

However, the temporary loss of aquatic habitat in Lake Macdonald due to the drawdown of the lake for safety during construction still resulted in a moderate residual impact after mitigation measures were applied. There will be a temporary loss of approximately 97.2% of aquatic habitat (by water volume) in Lake Macdonald, which requires additional mitigation in the form of a comprehensive aquatic fauna salvage operation. This is an unavoidable risk given the safety requirements of the Project.

Matters of National Environmental Significance were also assessed against the Significant Impact Criteria for critically endangered, endangered and vulnerable aquatic species. The assessment indicates that, while there may be temporary impacts, a significant impact from the Project on aquatic species that are Matters of National Environmental Significance is unlikely.

Fish passage is currently not provided at the Six Mile Creek dam. While fishway options have been considered for the Six Mile Creek dam upgrade (e.g. frc environmental, 2016; Seqwater 2018), preliminary advice from the Department of Agriculture and Fisheries indicates that the risk of aiding upstream dispersal by the noxious fish, tilapia, outweighs the benefit of providing fish passage for native species. Off-site mitigation measures for fish passage are therefore proposed; specifically the provision of fish passage at Gympie Weir by installation of a suitable fishway.

1 Introduction

1.1 Scope and Objectives of Study

frc environmental was commissioned by SMEC on behalf of Seqwater to undertake an assessment of potential risks to the aquatic ecological values of Six Mile Creek, and to identify impact mitigation strategies for the Six Mile Creek Dam Safety Upgrade project (the Project). The scope of the study involved:

- · describing the aquatic Environmental Values of Six Mile Creek and Lake Macdonald
- · reviewing the legislative and policy framework for protecting aquatic ecological values in Queensland
- identifying potential sources of impact to the aquatic ecological values associated with the construction and operational phases of the project, and assessing the level of risk to aquatic ecological values associated with each source of impact
- identifying likely impact mitigation strategies to reduce the level of risk associated with each identified source of impact, and assessing the likely effectiveness of each strategy, and
- providing recommendations regarding preferred impact mitigation strategies and other management controls to protect the aquatic ecological values of Six Mile Creek.

2 Overview of the Mary River and Six Mile Creek and Lake Macdonald

2.1 Mary River

From its headwaters in the Sunshine Coast hinterland near the township of Conondale, the Mary River flows north for approximately 290 km past the towns of Kenilworth, Gympie, Tiaro and Maryborough, before flowing into the Great Sandy Strait near Hervey Bay (Map 2.1). The Mary River downstream of the confluence with Six Mile Creek is stream order 8.

The predominant land use in the Mary River Basin is grazing on cleared land; however, there are also forestry reserves, national parks, and rural and urban areas throughout the basin (Johnson 1997). There are numerous weirs and dams along the Mary River and its tributaries, including Gympie Weir, Borumba Dam, Lake Baroon, Tallegalla Weir, Teddington Weir and the Mary River Barrage (Map 2.1).

The Mary River has perennial flow, with flows greater than 10 ML/day occurring approximately 95% of time (Figure 2.1 and Figure 2.2). The 50th percentile flow is approximately 500 ML/day in the upper Mary River and approximately 900 ML/day in the lower Mary River.

Applicable Environmental Values (EVs) pursuant to the Environmental Protection (Water) Policy (DERM 2010) for the upland and lowland freshwaters of the Mary River are shown in Table 2.1. The Water Plan (Mary Basin) 2006 presents ecological outcomes for the Mary River:

- · minimise changes to the low flow regime of the river, and
- minimise changes to the hydraulic habitat requirements of species such as the Mary River cod, Mary River turtle and lungfish.



Figure 2.1 Flow duration curve since 1963 for the upper Mary River at Moy Pocket (gauging station 138111A).

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Figure 2.2 Flow duration curve since 1968 for the lower Mary River at Fisherman's Pocket (gauging station 138007A).

Environmental Value	Upper Mary River	Lower Mary River	Six Mile Creek
Aquatic ecosystems	\checkmark	\checkmark	\checkmark
Irrigation	\checkmark	\checkmark	\checkmark
Farm supply	\checkmark	\checkmark	_
Stock water	\checkmark	\checkmark	\checkmark
Aquaculture	\checkmark	\checkmark	\checkmark
Human consumers of fisheries	\checkmark	\checkmark	\checkmark
Primary recreation	\checkmark	\checkmark	\checkmark
Secondary recreation	\checkmark	\checkmark	\checkmark
Visual recreation	\checkmark	\checkmark	\checkmark
Drinking water	\checkmark	\checkmark	\checkmark
Industrial use	\checkmark	\checkmark	_
Cultural and spiritual values	\checkmark	\checkmark	\checkmark

Table 2.1Environmental Values pursuant to the EPP(Water) for the Mary River and Six
Mile Creek.

✓ EV applies

EV does not apply



2.2 Six Mile Creek

Six Mile Creek is a large (i.e. stream order 5) tributary of the Mary River, originating inland from Noosa Heads and flowing for approximately 60 km north-west to join the Mary River approximately 4.5 km south of Gympie at 36 km from the Mary River estuary (Map 2.1). Land uses in the catchment of Six Mile Creek include forestry, grazing, horticulture, rural residential and urban areas.

Six Mile Creek also has reasonably perennial, albeit variable, flow, with flows greater than 10 ML/day occurring approximately 80% of the time at the Cooran gauging station (AMTD 32.4 km) (Figure 2.3). Flow in Six Mile Creek is an order of magnitude lower than for the upper Mary River (i.e. the 50th percentile flow in Six Mile Creek is approximately 50 ML/day). Monthly median flows are highest in March and lowest through the late winter and spring months (Hydrobiology 2008), although there is high variability in the magnitude of daily and inter-annual flows (Figure 2.4 and Figure 2.5).

Applicable Environmental Values (EVs) pursuant to the Environmental Protection (Water) Policy (EPP) for Six Mile Creek are shown in Table 2.1. The aquatic ecosystems EV is described in detail in Section 3. The Water Plan (Mary Basin) 2006 presents ecological outcomes for the Six Mile Creek, including minimising changes to the low flow regime of the creek, and minimising changes to the hydraulic habitat requirements of species such as the Mary River cod and lungfish



Figure 2.3 Flow duration curve since 1981 for Six Mile Creek at Cooran (gauging station 138107B).



Figure 2.4 Daily flow from 2007 to 2017 in Six Mile Creek at Cooran (gauging station 138107B).



Figure 2.5 Total annual flow from 1982 to 2017 in Six Mile Creek at Cooran (gauging station 138107B)

2.3 Six Mile Creek Dam

Six Mile Creek Dam is located in the Noosa Shire local government area on the upper reaches of Six Mile Creek at ATMD 55 km, and is 95 km from the Mary River estuary. It was built in 1965 for the purpose of town water supply, and upgraded (i.e. 3.6 m was added to the original height of the dam to increase storage capacity) in 1980. The catchment area for the dam is approximately 49 km², with surrounding land uses including agriculture, forestry, rural residential, and a variety of uses such as conservation, tourism and recreational activities. Seqwater leases areas of land adjacent to Lake Macdonald to the Gerry Cook Fish Hatchery, which is managed by the Mary River Catchment Coordinating Committee (MRCCC). Seqwater also provides a range of recreation facilities at Lake Macdonald including parklands, picnic facilities, boat ramps, a viewing platform and a designated canoe trail, and allow unpowered boating (electric motors only) and fishing on the lake:

The existing dam is an ungated zoned earth and rock fill dam (Table 2.2). The spillway consists of anchored concreted slabs on compacted earth fill, with an uncontrolled ogee crest. The capacity of the dam is 8,018 megalitres (ML) at a Full Supply Level (FSL) of 95.32 m Australian Height Datum (AHD), creating an impoundment area of approximately 260 hectares (ha) at this level with a maximum depth of 10.5 m. The existing dam does not incorporate a fishway.

Total annual outflows (i.e. combined spillways flows and environmental releases via the outlet works) occur 75% of the time on average, with seasonal outflows highest in autumn (94% of the time on average) and lowest in spring (51% of the time on average). The Resource Operations Plan licences the SEQ Water Grid Manager and Queensland Bulk Water Supply to take a combined annual total of 3500 ML to be extracted from Six Mile Creek Dam when it is above the minimum operating volume of 22 ML / 87.7 m AHD, at prescribed rates for each water licence holder. Daily environmental releases are made from Six Mile Creek Dam through the outlet works pursuant to the Resource Operational Plan using the following in-flow / out-flow rules:

- \cdot 0.25 ML out-flow when in-flows are 1.0 10.0 ML,
- \cdot 2.0 ML out-flow when in-flows are 10.0 30 ML, and
- 5.0 ML out-flow when inflows are >30 ML.

Design Parameter	Existing Dam
Spillway type	Uncontrolled fixed ogee crest
Spillway Description	Concrete slab broad crest weir
Spillway crest elevation (low level)	Notch/initial: RL 95.32 m AHD
	Full width: RL 95.35 m AHD
Spillway crest elevation (high level)	Not applicable
Stilling basin floor elevation	RL 83.5 m
Energy dissipation method	Plunge pool/stilling basin
Full supply level	8,018 ML
Dead storage	RL 87.7 m
Historical No Failure Yield	7,118 ML/y
Maximum depth	10.5 m
Area inundated at FSL	260 ha

Table 2.2 Ke	y parameters of the	existing dam
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3 Review of Applicable Legislation

3.1 Commonwealth Environment Protection and Biodiversity Conservation Act 1999

The Commonwealth *Environment Protection and Biodiversity Conservation Act* 1999 (EPBC Act) provides the legal framework for the protection and management of nationally and internationally threatened flora and fauna (including migratory species), ecological communities, internationally important wetlands, heritage places, the Great Barrier Reef, and Commonwealth marine areas, which are collectively defined as Matters of National Environmental Significance (MNES). Water resources in relation to coal seam gas and large mining projects, and nuclear actions, are also regulated under the EPBC Act.

The EPBC provides protection for threatened flora, fauna and ecological communities by:

- · identifying and listing of species and ecological communities as threatened
- developing conservation advice and recovery plans for listed species and ecological communities
- · developing a register of critical habitat
- · recognising key threatening processes
- where appropriate, reducing the impacts of these processes through threat abatement plans and non-statutory threat abatement advices, and by
- requiring approval for certain actions or activities that will, or are likely to, have a significant impact on an MNES or other protected matter.

Thirty-six species of freshwater fish and seven species of freshwater turtles¹ are listed as threatened under the EPBC Act, with four of these species known from the Mary River and Six Mile Creek²:

- white-throated snapping turtle (*Elseya albagula*), critically endangered
- Mary River cod (*Maccullochella mariensis*), endangered
- Mary River turtle (*Elusor macrurus*), endangered, and
- Australian lungfish (*Neoceratodus forsteri*), vulnerable.

¹ EPBC Act list of threatened fauna (<u>http://www.environment.gov.au/cgi-bin/sprat/public/publicthreatenedlist.pl#fishes extinct in the wild</u>); viewed 29 March 2018.

² The PMST (Appendix A) indicated that black rock cod (*Epinephelus daemelii*) may occur within 10 km of the Project area. However, this species is a marine fish associated with rocky reefs, and it is not known from freshwater habitats, and therefore would not occur in any habitat that is potentially influenced by the Project. Consequently, this species was not considered in this study.

The EPBC provides guidance on whether an action (e.g. a proposed development) is likely to have a significant impact on a MNES. Significant Impact Guidelines 1.1 (DEE 2013) provide guidance, in the form of assessment criteria, in relation to significant impacts on threatened species under the EPBC Act.

An action is likely to have a significant impact on a threatened species (i.e. critically endangered, endangered and vulnerable species) if there is a real chance or possibility that it will:

- lead to a long-term decrease in the size of a population (important population for vulnerable species)
- reduce the area of occupancy of the species (important population of a vulnerable species)
- fragment an existing population (important population of a vulnerable species) into two or more populations
- · adversely affect habitat critical to the survival of a species
- disrupt the breeding cycle of a population (important population of a vulnerable species)
- modify, destroy, remove, isolate or decrease the availability or quality of habitat to the extent that the species is likely to decline
- result in invasive species that are harmful to a critically endangered, endangered or vulnerable species becoming established in the endangered, critically endangered or vulnerable species' habitat
- · introduce disease that may cause the species to decline, and / or
- · interfere with the recovery of the species.

Where assessment identifies that an action will have a significant impact on a threatened species, then the action will be determined as a 'controlled' action and require appropriate environmental assessment within the approval application process.

3.2 Queensland Environmental Protection Act 1994

The Queensland *Environmental Protection Act 1994* (EPA 1994) provides the legislative framework for ecologically sustainable development in Queensland, requiring people, companies and government to take all reasonable and practical steps to protect Environmental Values (i.e. avoid harm to the environment). The Act provides a range of mechanisms to achieve the objective of the Act, including establishing Environmental Protection Policies that present the strategies for protecting Environmental Values.

3.3 The Environmental Protection (Water) Policy 2009

The Environmental Protection (Water) Policy 2009 (EPP Water) supports the Act through:

- · identifying High Ecological Value (HEV) waters
- · identifying Environmental Values (EV) and management goals for water
- providing water quality guidelines and Water Quality Objectives (WQO) to enhance or protect the identified EV
- · providing a framework for decision making about Queensland waters, and
- · requiring the monitoring of, and reporting on, the condition of Queensland waters.

EVs for Queensland waters includes the protection of aquatic ecosystems. The components of aquatic ecosystems to be protected are generally specified under the EPP (Water) for a given waterway if WQOs have been listed under Schedule 1 of the EPP (Water). For the Mary River, the components of aquatic ecosystems to protected are:

- existing water quality (20th, 50th and 80th percentiles), habitat, biota (fish and macroinvertebrates), flow and riparian areas in HEV waters, and
- water quality and riparian vegetation for all other waters, with:
 - the median concentration of several independent samples to achieve the scheduled WQO for the appropriate water type for physico-chemical water quality parameters
 - the 95th percentile concentration of several independent samples to achieve the National WQO for the appropriate water type for toxicant water quality parameters, and
 - riparian vegetation to achieve the applicable vegetation code under the Vegetation Management Act 1999.

A defensible assessment of the Environmental Values for water within the framework established by the EPP (Water) would also include the assessment of protected matters relevant to aquatic ecology, including matters protected under the EPBC Act and Queensland's *Nature Conservation Act 1992*, *Fisheries Act 1994* and *Vegetation Management Act 1999*³.

³ These and other protected matters also comprise Matters of State Environmental Significance (MSES), which are components of Queensland's biodiversity of state interest defined under the State Planning Policy and the *Environmental Offsets Regulation 2014*. MSES comprise certain Environmental Values protected under Queensland Legislation, including the *Environmental Protection Act 1994*, *Nature Conservation Act 1992*, *Fisheries Act 1994* and *Vegetation Management Act 1999*.

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3.4 Queensland Nature Conservation Act 1992

The *Nature Conservation Act 1992* provides for the conservation of Queensland's nature by declaring and managing a protected area network, protecting threatened species (wildlife) and their habitats, regulating the taking of wildlife and co-ordinating nature conservation with Traditional Owners and other land owners. Several freshwater species are protected wildlife under the *Nature Conservation Act 1992*.

Protected wildlife listed under the *Nature Conservation Act 1992* must be protected from threatening processes, and critical habitat for protected wildlife is required to be protected to the greatest extent possible.

3.5 Queensland Fisheries Act 1994

The *Fisheries Act 1994* provides for the management and protection of fisheries resources, including regulating development that might impact declared fish habitat areas and fish passage. Several fish species of special interest are listed as 'no take' species under the Act, including Australian lungfish.

Fisheries resources, including declared fish habitat areas, waterways providing for fish passage and marine plants which are MSES, contribute to the Environmental Values of waterways and wetlands.

Fish passage is applicable to the current study (assessable development), with development potentially impacting fish passage being either:

- accepted development, where the design of infrastructure strictly conforms to the Department of Agriculture and Fisheries' (DAF's) Accepted Development Requirements for Operational Work that is Constructing or Raising a Waterway Barrier Works (2017), or
- assessable development, where the proposed development requires assessment by DAF (via SARA) and the design of the development is required to demonstrate compliance with the State Development Assessment Provisions (SDAP State code 18).

3.6 Queensland Vegetation Management Act 1999

The Vegetation Management Act 1999 (VMA 1999), as updated by the Vegetation Management and Other Legislation Amendment Act 2018, regulates the clearing of

vegetation to conserve threatened regional ecosystems, protect biodiversity and maintain ecological processes, amongst other purposes.

The VMA 1999 provides for the chief executive to certify various classes of regulated vegetation maps, with regulated vegetation a MSES. Classes of vegetation under the VMA include: vegetation that is remnant and / or threatened (category B), high value regrowth vegetation (category C) or regrowth vegetation in a wetland, watercourse or drainage feature area within a Great Barrier Reef catchment (category R). Vegetation in wetland areas and vegetation intersecting a watercourse is also regulated vegetation under the VMA. Vegetation clearing and development is regulated for Category R vegetation areas, and Riverine Protection Permits are required to clear vegetation in watercourses.

3.7 Queensland Water Act 2000

The Queensland *Water Act 2000* provides for the sustainable management of water resources in Queensland, including requiring permits for works within watercourses and providing for the sustainable allocation of water for environmental purposes (i.e. environmental flows to protect ecological functions in rivers). Many of these functions are documented in Resource Operations Plans (ROP) or Water Plans for a catchment, which include ecological outcomes and obligations for water licence holders or water scheme operators. For example, in relation to the Project, Seqwater (as a water licence holder for Six Mile Creek Dam) is obliged to make specified environmental flow releases from the dam to meet ecological outcomes in Six Mile Creek.

3.8 Queensland Biosecurity Act 2014

The aim of the *Biosecurity Act 2014* is to manage risks associated with exotic pests and diseases that impact plant and animal industries including aquaculture and wild capture fisheries, tourism, infrastructure including water supply, shipping, biodiversity, and the natural environment. The Act seeks to provide a framework to minimise biosecurity risks and to support a 'risk-based' approach to biosecurity management.

The Act defines biosecurity matters (i.e. *prohibited* matters, which are not yet present in Queensland; and *restricted* matters, which are currently present in Queensland); establishes a General Biosecurity Obligation (GBO); and establishes specific obligations in relation to prohibited and restricted matter.

The GBO requires those who deal with a biosecurity matter or carries out an activity, and knows or ought reasonably to know, that the biosecurity matter or activity poses, or is likely

to pose, a biosecurity risk are obliged to take all reasonable measures to minimise the likelihood of causing a biosecurity risk, and / or do whatever is reasonably required to minimise the adverse effects of dealing with a biosecurity matter. Specifically, they may not keep or possess, whether intentionally or otherwise, the biosecurity matter, or propagate, raise, distribute or transport the biosecurity matter.

Prohibited matter are listed in Schedule 1 of the Act, and restricted matter are listed in Schedule 2. Aquatic pests that are restricted biosecurity matters listed in Schedule 2 of the Act include⁴:

- · various pathogens
- fish, including but not limited to eastern Gambusia (*Gambusia holbrooki*), carp (*Cyprinus carpio*) and tilapia (*Oreochromis mossambicus*)
- aquatic plants, including but not limited to salvinia (*Salvinia molesta*), water hyacinth (*Eichhornia crassipes*), and cabomba (*Cabomba caroliniana*), and
- other plants that are common weeds of riparian areas.

3.9 Local Government Plans

Six Mile Creek Dam, and Six Mile Creek, are located in the Noosa Shire Council (NSC) Local Government Area (LGA).

Corporate Plan 2017-2037

The NSC Corporate Plan 2017-2037 presents a vision for the Noosa Shire, including for the shire's natural environment. The 20 year environment goal under the Corporate Plan is 'Our environment is protected, enhanced and values by the community'.

Over the next five years adopting an over-arching Environment Strategy (incorporating a Biodiversity Plan, Waterways and Wetlands Management Plan, amongst other sub-plans) and revitalising the Noosa River Plan are key focus areas identified in the Corporate Plan.

In 20 years, aspirational targets include (amongst others):

· The Noosa River is clean, has abundant fish life and the river maintains it's a rating

⁴ Reprint current from 3 July 2017, viewed 23 April 2018.

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- Integrated catchment management practices have improved our waterways and wetlands
- · Biodiversity in the Noosa Shire has been preserved, and
- The Noosa natural environment is protected and enhanced.

While the NSC Corporate Plan focuses on the Noosa River, protecting Mary River cod and lungfish (and white-throated snapping turtle and Mary River turtle if they occur) in Six Mile Creek will be a key contribution to achieving preservation of biodiversity in the LGA.

Operational Plan 2018 – 2019

The NSC Operational Plan underpins the Corporate Plan, with the Operational Plan updated annually to present specific projects to be undertaken to progress the achievement of the Corporate Plan's vision. The Operational Plan is closely aligned with Councils annual budget priorities.

Priority projects under the current (i.e. 2018 – 2019) Operational Plan that support the Corporate Plan's environmental vision 'Our environment is protected, enhanced and valued by the community' include (amongst others):

- adopt an initiate implementation of Environmental Strategy, and
- adopt and implementation of Noosa River Management Plan.

Protection of water quality, habitat and aquatic biodiversity of Six Mile Creek and Six Mile Creek dam would contribute towards the implementation of NSC's Environmental Strategy (noting the Environmental Strategy is still being drafted).

4 Aquatic Ecological Values of Six Mile Creek and Lake Macdonald

The aquatic ecological values of Six Mile Creek and Lake Macdonald have been assessed using:

- · literature and database review, including:
 - DNRM (2004) Mary Basin Draft Water Resource Plan: Environmental Conditions Report including Mary River, Burrum River and Beelbi Creek Catchments
 - The MNES Protected Matters Search Tool and the Species Profile and Threats (SPRAT) database
 - The MSES search tool, for relevant MSES including declared fish habitat areas, wetland protection areas, HEV waters, regulated vegetation in riparian areas)
 - Department of Environment and Science's (DES's) Wildlife Online database
 - Atlas of Living Australia
- **synthesis of existing data** for aquatic habitat, water quality, aquatic plants, fish and turtles, collected:
 - within the scope of the Northern Pipeline Interconnector Stage 2 (NPI2) Aquatic Habitat Monitoring Program
 - within the scope of baseline studies, including field surveys, completed for the Six Mile Creek Dam Upgrade Project (frc environmental 2016)
 - by Seqwater during routine water quality monitoring programs
- **field survey**, for macroinvertebrates because there was no recent existing data
- **consultation with key experts**, including academics, agency staff and other relevant stakeholders (frc environmental 2016).

Overall, aquatic ecology and water quality data was assessed at 13 sites on Six Mile Creek, and two sites on Lake Macdonald, for the Project (Table 4.1 and Map 4.1). Water quality data collected by Seqwater at an additional four sites was also assessed. Consultation with key experts was completed in 2015 / 2016 and was presented with the detailed baseline aquatic ecology study (frc environmental 2016). Data for observations of platypus was provided by SMEC.

Table 4.1. Survey sites.

Site	Description	Easting ^a	Northing ^a	Assessed Parameters ^b	Survey Dates	
Six Mile Creek – Downstream of Lake Macdonald						
SMC4	Six Mile Creek; approximately 19 km DS of the Lake Macdonald spillway	480965	7087785	H, WQ	Oct-13, Nov-14, Nov-15, Oct-16, Nov-17	
SMC5	Six Mile Creek; approximately 28.5 km DS of the Lake Macdonald spillway	473906	7095982	H, WQ	Oct-13, Nov-14, Nov-15, Oct-16, Nov-17	
SMCDS05	Six Mile Creek; approximately 45 km DS of the Lake Macdonald spillway	469799	7098501	H, WQ, F, T	Aug-15	
SMCDS04	Six Mile Creek; approximately 7 km DS of the Lake Macdonald spillway	489171	7086480	H, WQ, F, T	Aug-15, Oct-15, Feb-18	
SMCDS03	Six Mile Creek; approximately 4.5 km DS of the Lake Macdonald spillway	491312	7085143	H, WQ, P, F, T, M	Aug-15, Oct-15, Feb-18	
SMCDS02	Six Mile Creek; approximately 1.5 km DS of the Lake Macdonald spillway	492715	7083047	H, WQ, P, F, T, M	Aug-15, Oct-15, Feb-18	
SMCDS01	Six Mile Creek; directly DS of the Lake Macdonald spillway within the spillway stilling basin	493026	7082149	H, WQ, P, F, T, M	Aug-15, Oct-15, Feb-18	
Lake Macdon	ald					
DS Lake	Lake Macdonald; approximately 1 km US of the spillway	493864	7081893	H, WQ, F, T, M	Aug-15, Oct-15, Feb-18	
US Lake	Lake Macdonald; approximately 2.5 km US of the spillway	494340	7080694	H, WQ, F, T	Aug-15, Oct-15	

Six Mile Cree	Six Mile Creek – Upstream of Lake Macdonald						
CU02	Cooroy Creek; approximately 4 km US of the Lake Macdonald spillway	493582	7078933	H, WQ, F, T	Aug-15, Oct-15		
CU01	Cooroy Creek; approximately 4.5 km US of the Lake Macdonald spillway	493179	7078657	H, WQ, F, T	Aug-15, Oct-15		
CU03	Cooroy Creek; approximately 5 km US of the Lake Macdonald spillway	493019	7078615	H, WQ, F, T	Aug-15, Oct-15		
SMCUS02	Six Mile Creek; approximately 5.5 km US of the Lake Macdonald spillway	495930	7079951	H, WQ, F, T	Aug-15, Oct-15		
SMCUS01	Six Mile Creek; approximately 6 km US of the Lake Macdonald spillway	496073	7079594	H, WQ, F, T, M	Aug-15, Oct-15, Feb-18		
SMCUS03	Six Mile Creek; approximately 11 km US of the Lake Macdonald spillway	494453	7077331	H, WQ, F, T	Aug-15, Oct-15		

^a WGS84 (Zone 56J)

^b H = habitat; WQ = water quality; P = aquatic plants; M = macroinvertebrates; F = fish; T = turtles



4.1 Aquatic Habitat

Assessment Methods

In addition to literature and databases described above, the aquatic habitat of Six Mile Creek and Lake Macdonald was assessed through the review of:

- Aquatic Conservation Values, as assessed by the Queensland Environmental Protection Agency using the Aquatic Biodiversity Assessment and Mapping Method (AquaBAMM)
- DAF's Waterway barrier risk layer
- DES's Wetland Maps layer, and
- · Queensland's floodplain assessment and groundwater-dependent ecosystem (GDE) layers.

The survey data that were synthesised were collected using the methods used for the NPI2 Aquatic Habitat Monitoring Program (for Seqwater), which were modified from State of the River methodology (Anderson 1993a, Anderson 1993b). The habitat assessment method was developed to specifically assess suitability of aquatic habitat for Mary River cod, Australian lungfish, Mary River turtle and white-throated snapping turtle at each site, which are the aquatic MNES potentially occurring in the Project area (Appendix A⁵ and B). The habitat assessment included:

- habitat condition, and type and cover of key habitat features (e.g. pools, riffles, undercut banks, large woody debris)
- · depth and width of streams
- substrate composition
- · habitat sensitivities to various impacts, and
- · identification of existing disturbances, including fish passage barriers.

⁵ The PMST (Appendix A) indicated that black rock cod (*Epinephelus daemelii*) may occur within 10 km of the Project area. However, this species is a marine fish associated with rocky reefs, and it is not known from freshwater habitats, and therefore would not occur in any habitat that is potentially influenced by the Project. Consequently, this species was not considered in this study

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Results

Six Mile Creek is a low-gradient, low energy stream, with notophyll vine forest the dominant native riparian vegetation (DNRM 2004). Extensive deposits of large woody debris are an important natural feature of low energy streams, and are a common habitat element in Six Mile Creek (DNRM 2004). Medium length pools (i.e. between 6 and 12 channel widths in length) that are less than 2 m deep are common in Six Mile Creek, with riffles and shallow glides over sand also present (DNRM 2004).

With the exception of the Six Mile Creek Dam, disturbances along Six Mile Creek are relatively minor (DNRM 2004). Downstream of Lake Macdonald, flood flows have been reduced, low flows have been substantially reduced, and dry spells increased (DNRM 2004). As a result of the lower water levels in Six Mile Creek downstream of Lake Macdonald, large woody debris is more exposed (i.e. no longer providing as much submerged habitat for fish or turtles), riffles have declined, and consequently there is less suitable habitat for fish and turtle passage along this section of Six Mile Creek (DNRM 2004), although hydrological analysis by Hydrobiology (2008) suggest that the hydrological regime is not significantly different between current and pre-development scenarios. Sediment transport processes in the lower reaches of Six Mile Creek are likely to have been altered by Lake Macdonald, which captures over 90% of the incoming sediment load (DNRM 2004).

Six Mile Creek is a stream order 5 watercourse along its downstream reaches, with mapped floodplain and GDEs along or near Six Mile Creek (Map 4.2). The following Matters of State Environmental Significance, and aquatic conservation areas, are present (Map 4.3, 4.4, and 4.5):

- Matters of State Environmental Significance:
 - High Ecological Value (HEV) waters (waterways)
 - regulated vegetation: category R vegetation; vegetation intersecting a watercourse; and vegetation within 100m of a wetland
 - Waterways providing for fish passage (Six Mile Creek is identified as having major risk of impact to fisheries resources from waterway barrier works
- Aquatic Conservation Assessment:
 - riverine and lacustrine (i.e. Lake Macdonald) wetlands, and

 very high and high conservation significance riverine areas in Six Mile Creek downstream of Lake Macdonald, and medium conservation significance riverine areas upstream of Lake Macdonald.

The detailed results of habitat surveys for each site from 2013 to 2018 are presented in Appendix C. The key habitat features of:

- Six Mile Creek *downstream* of Lake Macdonald include:
 - well-defined channel with high steep banks, and undercut banks present at most sites
 - substrate dominated by clay and silt, with gravel riffles present at most sites and bedrock present at some sites
 - high variation in flow, with riffles, runs and shallow and deep pools present at most sites
 - low abundance of submerged aquatic flora, except at site SMCDS01 immediately downstream of Six Mile Creek Dam where there was some Cabomba caroliniana⁶ and Nymphoides indica
 - abundant large woody debris and leaf packs at each site
 - riparian vegetation is in good condition, providing shade and a supply of fine and large woody material
 - presence of suitable breeding locations for Mary River cod, but very limited breeding habitat for Australian lungfish
 - absence of suitable nesting habitat for Mary River and white-throated snapping turtle, and few places for turtle basking
 - an existing waterway barrier (i.e. rubble from the old road bridge) approximately
 50 m downstream of the Six Mile Creek Dam
- within Lake Macdonald include:
 - a single flow habitat (deep pool)

⁶ Cabomba caroliniana is a restricted biosecurity matter under the Biosecurity Act 2014

- high abundance of submerged aquatic flora, predominantly Cabomba caroliniana and Nymphoides indica, and beds of emergent Persicaria spp. near lake margins
- substrate dominated by silt, with some sand near banks
- absence of suitable breeding locations for Mary River cod and Australian lungfish
- absence of suitable nesting habitat for Mary River turtle and white-throated snapping turtle, and few places for turtle basking
- limited large woody debris
- Six Mile Creek *upstream* of Lake Macdonald include:
 - well-defined channel with high steep banks, and undercut banks present at most sites
 - the flow habitats of some sites were influenced by impounded water; with site SMCUS03 having moderate diversity of flow habitats, with riffles, runs and deep pools all present
 - SMCUS01, SMCUS02 and Cooroy Creek sites were choked by Cabomba caroliniana
 - presence of potentially suitable breeding locations for Mary River cod, but very limited breeding habitat for Australian lungfish
 - limited suitable nesting habitat for Mary River and white-throated snapping turtle, and
 - turtle basking places present at most sites.














4.2 Water Quality

Assessment Methods

In addition to the data and information sources described above, water quality data from the Department of Natural Resources, Mines and Energy's (DNRME's) gauging station on Six Mile Creek at Cooran (station number 138107B) was also assessed.

Water quality data that was collected within the scope of the NPI2 AHMP and the baseline studies for the Six Mile Creek project was collected in accordance with the DES' *Monitoring and Sampling Manual* (DES 2018). Water temperature (°C), pH, dissolved oxygen (percent saturation and mg/L) and electrical conductivity (μ S/cm) were measured in situ within 0.3 m of the water surface using a Hydrolab Quanta multi-parameter water quality meter. The meter was calibrated daily according to the manufacturer's instructions. Turbidity (NTU) was measured in situ within 0.3 m of the water surface using a HACH 2100Q turbidity meter, which was calibrated at the commencement of each field survey.

Water quality results were compared to the Water Quality Objectives (WQOs) for lowland streams as presented in the *Environmental Protection (Water) Policy 2009* Mary River Environmental Values and Water Quality Objectives Basin No. 138, including all tributaries of the Mary River (DERM 2010). These WQOs relate to protection of the aquatic ecosystem Environmental Value.

Parameter	Unit	WQO
Turbidity	NTU	<50
Dissolved oxygen	percent saturation	85 – 110
рН	unit	6.5 - 8.0
Electrical conductivity ^a	μS/cm	<626
Suspended solids	mg/L	<6
Chlorophyll a	µg/L	<5
Total nitrogen	µg/L	<500
Oxidised nitrogen	μg/L	<60
Ammonia	µg/L	<20
Organic nitrogen	μg/L	<420
Total phosphorus	μg/L	<50
Filterable reactive phosphorus	μg/L	<20

Table 4.2Published WQOs for protection of aquatic ecosystems for lowland freshwater
in the Mary River for selected water quality parameters.

^a based on the 75th percentile of the Sandy Coastal salinity zone in Appendix G of the Queensland Water Quality Guidelines.

Results

Water quality in Six Mile Creek is generally good and typically achieves the WQO for the protection of aquatic ecosystems EV, although dissolved oxygen levels are often low due to decomposition of organic matter from the adjacent rainforest canopy (DNRM 2004). DNRM (2004) also report that water quality typically achieves other EVs identified for Six Mile Creek, including Primary Industry, Recreation and Drinking Water (assuming disinfection) (DNRM 2004).

Water quality data for water temperature, electrical conductivity and pH were available on the DNRME Water Monitoring Portal, with records commencing in December 1998. Results showed that both electrical conductivity and pH complied with the applicable WQO for aquatic ecosystems (there is no WQO for temperature) (Table 4.3).

Seqwater's water quality monitoring data for Lake Macdonald (Appendix D) shows that:

- total nitrogen, organic nitrogen, chlorophyll-a and dissolved aluminium were commonly higher than the applicable WQO in Lake Macdonald
- total aluminium, total zinc and total cobalt were sometimes higher than the applicable WQO in Lake Macdonald
- pH, dissolved oxygen, total suspended solids, total nitrogen, organic nitrogen, ammonia, total aluminium, total chromium, total copper, total mercury, total zinc and dissolved aluminium were non-compliant with the applicable WQO at the Lake Macdonald tailwater, and
- all other parameters (where data was available) complied with the applicable WQO in Lake Macdonald and at the Lake Macdonald tailwater.

Biogeochemical cycling of key elements, including nutrients and metals, in benthic sediments has a significant influence on water quality in reservoirs, with elevated concentrations of these parameters commonly observed in reservoirs (Grinham et al., 2018). Therefore, the above described results are likely typical of water quality conditions for reservoirs.

The *Water Monitoring Data Collection Standards* (DNRW 2007) define a reservoir as stratified if the temperature difference between surface and basement layers exceeds 5°C. Depth profile measurements of water temperature through the depth profile in Lake Macdonald (mid-lake) were summarised on a monthly basis between November 2011 and November 2017 (i.e. 70 months), with measurements for 69 of these months indicating no stratification. Stratification was detected in only one month (January 2015). Overall, these results indicate that Lake Macdonald rarely stratifies, and when it does it is only weakly stratified.

Survey data for water quality measured in situ since 2013 was pooled based on reach (i.e. downstream of Six Mile Creek Dam, in the impounded section of Six Mile Creek Dam, or upstream of Six Mile Creek Dam). Results showed that (Table 4.4, Table 4.5, Table 4.6; Appendix E):

- electrical conductivity, pH and turbidity complied with the WQO in all reaches of Six Mile Creek, and
- the percent saturation of dissolved oxygen was below the WQO in Six Mile Creek upstream and downstream of Lake Macdonald, but complied with the WQO in Lake Macdonald.

Summary statistic	Temperature (ºC) ª	Electrical conductivity (µS/cm)	pH (unit)
count of mean monthly data points	225	221	93
minimum	10.0	91	6.4
20 th percentile	15.0	145	6.6
median	19.6	171	6.8
80 th percentile	23.2	203	7.1
maximum	25.3	275	7.8

Table 4.3Summary of mean monthly water quality since December 1989 measured at
gauging station 138107B on Six Mile Creek at Cooran.

Grey shading indicates if a parameter exceeds the applicable WQO (note median was compared to the WQO).

^a There is no published WQO for water temperature.

Table 4.4Summary of water quality since from October 2013 to February 2018
measured at Six Mile Creek downstream of Lake Macdonald (i.e. sites SMC4,
SMC5, SMCDS05, SMCDS04, SMCDS03, SMCDS02 and SMCDS01).

Summary statistic	Temperature (°C) ª	Electrical conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	24	24	24	24	24	24
minimum	17.7	76	5.55	2.0	24.0	2.5
20 th percentile	19.0	114	6.21	3.0	33.5	5.2
median	20.9	161	6.78	4.5	49.1	7.9
80 th percentile	24.5	180	7.18	5.7	65.0	11.0
maximum	26.6	244	7.43	7.9	93.3	15.6

Grey shading indicates where the median value of a parameter does not comply with the WQO

Table 4.5 Summary of water quality since from October 2013 to February 2018 measured at Six Mile Creek in Lake Macdonald (i.e. sites DS Lake and US Lake).

Summary statistic	Temperature (°C) ª	Electrical conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	6	6	6	6	6	6
minimum	22.9	42	6.69	4.2	54.8	3.1
20 th percentile	23.2	53	6.77	7.7	99.2	3.5
median	24.5	87	6.99	8.8	105.6	4.5
80 th percentile	28.0	92	7.31	9.9	117.1	5.6
maximum	28.8	103	7.46	10.0	119.8	5.8

Table 4.6Summary of water quality since from October 2013 to February 2018
measured at Six Mile Creek upstream of Lake Macdonald (i.e. sites CU02,
CU01, CU03, SMCUS02, SMCUS01, SMCUS03).

Summary statistic	Temperature (°C) ^a	Electrical conductivity	pH (unit)	Dissolved Oxygen	Dissolved Oxygen (%	Turbidity (NTU)
	. ,	(µ5/cm)		(mg/L)	saturation)	. ,
count	13	13	13	13	13	13
minimum	17.4	59	5.80	2.1	24.7	4.5
20 th	18.9	89	6.25	3.1	34.1	5.8
percentile						
median	21.2	107	6.67	4.9	51.1	7.4
80 th	24.0	142	7.09	7.3	91.6	13.6
percentile						
maximum	188.0	189	7.61	8.9	110.1	15.4

Grey shading indicates where the median value of a parameter does not comply with the WQO

4.3 Aquatic Plants

Assessment Methods

Aquatic plants of Six Mile Creek were assessed through the review of:

- published literature, such as DNRM 2004 and the Mary River Aquatic Weeds Strategy 2010-2014 (Mary River Pest Management Group 2010), and
- publicly available databases, including
 - MNES and MSES search tools, to identify threatened plants species potentially occurring in Six Mile Creek
 - the Wildlife online and Atlas of Living Australia's databases.

The field survey was based on a timed meander methodology (approximately 15 minutes per site), as described in Flora Survey Guidelines for Protected Plants (EHP 2016), with both in-stream and bank habitats surveyed.

Results

Aquatic plants are absent or rare in Six Mile Creek downstream of Lake Macdonald, although they are common in Lake Macdonald, with the noxious weed *Cabomba caroliniana* growing prolifically in the dam (DNRM 2004) and the noxious *Hygrophila costata* occurring in high cover along the lake margins. Various attempts to control Cabomba in Lake Macdonald have been made, particularly by Noosa Shire Council in the past through the use of mechanical harvesting. Eradication of Cabomba is generally considered to be unviable with current control methods and attempts to reduce cover being labour intensive. Hygrophila infestations around Lake Macdonald have been effectively controlled by local stakeholder groups and its occurrence is now sparse. A plan to eradicate remaining Hygrophila from Lake Macdonald and the upper catchment is currently being considered.

The diversity and growth forms of aquatic plants in the wider Mary River are greater than in Six Mile Creek, with the diversity of aquatic weeds also higher in the Mary River compared to Six Mile Creek (Mary River Pest Management Group 2010).

The field surveys found (Table 4.7):

- the overall diversity of aquatic plant species and growth forms was low, with Lake Macdonald having the highest diversity of aquatic plant species both in the water and on the banks, and the highest diversity of growth forms in the water
- the cover of aquatic plants was low, mostly restricted to isolated occurrences, with the exception of:
 - Cabomba and water snowflake in Lake Macdonald
 - Lomandra on banks of the downstream and upstream reaches of Six Mile Creek
- no listed threatened species were recorded, reflecting the absence of records of threatened aquatic plants in the Project area
- introduced species included *Cabomba* and cape lily, with *Cabomba* a restricted biosecurity matter.

Species	Common name	Growth form	Six Mile Creek DS	Lake Macdonald	Six Mile Creek US	
Plants in water (in-stream)						
Cabomba caroliniana	Cabomba ^c	submerged	a	D	S	
Nymphoides indica	water snowflake	floating- attached	l a	S	I	
Ludwigia peploides	water primrose	emergent		b	I	
Philydrum lanuginosum	frog's mouth	emergent		b		
Eleocharis sp.	spike rush	emergent		b		
Nymphae caerulea	cape waterlily $^{\rm c}$	floating- attached		I		
<i>Azolla</i> sp.	Azolla	floating			I	
<i>Typha</i> sp.	bull rush	emergent		l p		
Potamogeton javanicus	Javan pondweed	submerged		I		
Lemnoideae	duck weed	floating			Ι	
Plants not in water (ba	nks)					
Lomandra spp.	mat rush	_	D	I	S	
Carex spp.	Sedge	_	I			
<i>Cyperus</i> spp.	flat sedge	-	I	I	I	
Persicaria spp.	knot weeds	-	I	I	I	
Ludwigia octovalvis	willow primrose	-		I		
Schoenoplectus mucronatus	bog bulrush	-	I			

Table 4.7 Results of aquatic plant surveys.

I = isolated; S = scattered; D = dense cover

^a Cabomba was found in Six Mile Creek downstream of Lake Macdonald, but only within several hundred meters of the dam. Most of Six Mile Creek downstream was without instream aquatic plants, including Cabomba.

^b these emergent species only occurred in shallow water along the edge of Lake Macdonald

^c denotes pest species

4.4 Macroinvertebrates

Assessment Methods

The field survey was implemented in February 2018, following nearly two months of baseflow conditions and several brief elevated flows in October, November and early December 2017, but before a high flow event in late February 2018. Thus, conditions were typical wet season conditions at the time of sampling. Sampling used the AUSRIVAS sampling protocol (DNRM 2001), with samples were collected over a 10 m length of stream using a standard triangular-framed dip net (250 µm mesh size) and the kick-sweep method to dislodge macroinvertebrates from the substrate. At each site, one sample was collected from bed habitat and another one from edge habitat. Samples were picked in frc environmental's biological laboratory, and identified to the lowest practical taxonomic level (in most instances, family) and counted, to comply with AUSRIVAS protocols.

Standard macroinvertebrate indices were calculated (i.e. taxonomic richness, PET richness and SIGNAL-2 Scores), and were compared to the guidelines for biological indicators for South East Queensland, as reported in the Queensland Water Quality Guidelines (EHP 2013) for lowland freshwaters, as there are no biological guidelines presented in the EPP(Water) for the Mary River:

- · taxonomic richness: \geq 22
- · PET richness: \geq 4, and
- · SIGNAL-2 score: \geq 4.

Results

Macroinvertebrate communities are highly variable between sites in Six Mile Creek, with taxonomic diversity low at some sites due to limited microhabitat diversity (DNRM 2004).

Survey results found low taxonomic diversity of macroinvertebrates in bed and edge habitat (Table 4.8 and Table 4.9) in Six Mile Creek, ranging from only eight taxa in bed habitat at site SMCDS04 to 25 taxa in edge habitat at site SMCDS04. The diversity of sensitive taxa, indicated by both PET richness, and the overall proportion of sensitive taxa, indicated by SIGNAL-2 Scores, was similar for bed and edge habitat, and often lower than the WQO, suggesting that sensitive taxa are not a dominant feature of macroinvertebrate communities in Six Mile Creek. Whilst there is no WQO for abundance, the results indicate that macroinvertebrates are abundant in Six Mile Creek. The number of samples collected upstream of, and within, Lake Macdonald were less than the number collected downstream, but the macroinvertebrate indices collected upstream of, and within Lake Macdonald were

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typically in the range recorded in Six Mile Creek downstream of Lake Macdonald, except SIGNAL-2 score in edge habitat (upstream of, and within, Lake Macdonald) and PET richness in edge habitat (upstream of Lake Macdonald).

	Macroinvertebrat	e Indices		
Site	Abundance Taxonomic PET Richne		PET Richness	SIGNAL 2
Six Mile Creek downstream of Lake Macdonald				
SMCDS04	167	8	1	3.40
SMCDS03	169	15	4	4.11
SMCDS02	153	19	5	4.12
SMCDS01	190	14	3	3.48
Lake Macdonald				
DS Lake	131	10	2	3.83
Six Mile Creek upstream of Lake Macdonald				
SMCUS01	204	13	1	3.59

Table 4.8	Mean macroinvertebrate indices in bed habitat at each site in February 2018

Grey shading indicates where a macroinvertebrate index does not comply with the WQO

Table 4.9 Mean macroinvertebrate indices in edge habitat at each site in February 2018.

	Macroinvertebrate Indices				
Site	Abundance	Taxonomic Richness	PET Richness	SIGNAL 2	
Six Mile Creek downstream of Lake Macdonald					
SMCDS04	177	25	4	4.06	
SMCDS03	125	14	4	3.82	
SMCDS02	123	16	5	3.77	
SMCDS01	232	19	3	3.60	
Lake Macdonald					
DS Lake	163	15	4	3.31	
Six Mile Creek upstream of Lake Macdonald					
SMCUS01	126	17	1	3.45	

Grey shading indicates where a macroinvertebrate index does not comply with the WQO

4.5 Fish

Assessment Methods

In addition to the data and information sources described above, fish stocking data and reports for Lake Macdonald were reviewed.

The methods used to collect the fish data that was synthesised for this are presented in Appendix F, and were designed to record species diversity, abundance and life history stage (total length). The habitat preferences, reproductive ecology and migration strategies of fish species from Six Mile Creek were reviewed.

Results

The fish fauna in Six Mile Creek downstream of Lake Macdonald is generally in good condition and the creek provides significant breeding habitat for Mary River cod (DNRM 2004). Within Lake Macdonald, the impounded water is likely to favour some species native to Six Mile Creek (e.g. bony bream) as well as the recreational species (not native to the creek) that have been stocked in the dam (e.g. Australian bass, yellow belly).

Overall, a total of 26 native species, and five pest species, are known or likely to occur in Six Mile Creek, with a number of these species having been caught in the field surveys (Table 4.10). Mary River cod and Australian lungfish are both Matters of National and State Environmental Significance (Appendix A and B), and eastern Gambusia and tilapia are restricted biosecurity matters under the Queensland *Biosecurity Act 2014*. Tilapia were only recently recorded for the first time in Six Mile Creek downstream of Lake Macdonald. It is currently unknown if tilapia occur in Lake Macdonald, or if the dam wall has prevented this species extending further upstream.

Detailed results of the fish surveys for each site are presented in Appendix G, with the key results being:

- several species, including Pacific blue eye, were only present in Six Mile Creek downstream of Lake Macdonald, suggesting that the dam prevents these species from migrating further upstream
- several diadromous species expected to be present in Six Mile Creek were absent (e.g. sea mullet, pink eye mullet, estuary glassfish, striped gudgeon, empire gudgeon), which may reflect the cumulative impacts of barriers to fish passage in the lower Mary River between the estuary and Six Mile Creek

- Mary River cod and Australian lungfish were caught in Six Mile Creek downstream of Lake Macdonald
- common gudgeons were the most abundant species, with this species also having very high abundance in Lake Macdonald
- a seasonal pattern in abundance was recorded, with abundance of most species higher in spring compared to summer
- the abundance of pest fish (eastern Gambusia and swordtails) was highest in Six
 Mile Creek upstream of Lake Macdonald
- most species had healthy populations comprised of juveniles, sub-adults and adults
- bony bream attained a larger size in Lake Macdonald than in Six Mile Creek, but there were no other notable size differences between waterway and reservoir sections of Six Mile Creek for any other species, and
- during periods of low flow fish may become isolated in the tailwater pool downstream of Lake Macdonald.

Review of literature describing the preferred habitat, reproductive ecology and migration pattern of fish species in Six Mile Creek (Table 4.11; Fishbase (2010), Pusey et al. (2004), DAF (2015), DoE (2015)) showed that:

- · larger bodied species, such as Mary River cod and Australian lungfish, have a preference for deep pool habitats with large woody debris
- estuarine tolerant species (e.g. striped gudgeon, empire gudgeon, mullet and estuary glassfish), are predominantly distributed within lowland sections of the Mary River, including lowland tributaries
- most species prefer fine substrates (sand/gravel) with low to moderate flow
- Mary River cod spawn annually in spring, soon after the water temperature rises to 20°C. Spawning activity may be increased on a full moon.
- Australian lungfish spawn in slow-flowing shallow pools among aquatic plants between August and December, where they deposit eggs on plants or submerged matts of fine roots
- bullrout, sea mullet, Australian bass, Australian smelt, common gudgeon, and striped gudgeon spawn in winter and early spring, prior to increased water temperature and flows
- most other fish species spawn in summer with increased water temperature and water flow

- The fish species in Six Mile Creek have a range of migration and dispersal patterns, including species that must migrate to estuaries or the ocean to complete their life cycle (diadromous – including species with catadromous and amphidromous forms of diadromy), and species that migrate within freshwater reaches (potamodromous).
- Australian lungfish are generally inactive during the day, where they occupy shaded, deep-water complex habitats, although juveniles typically occupy dense submerge aquatic plant meadows (Kind 2002). Movement occurs mostly at night, with daily ranges greater in summer than in winter, with total linear ranges varying between 0.3 km and 74.9 km and mean linear home range being approximately 1.5 km (Kind 2002). Movement distances are larger in impounded reaches than in flowing reaches, as suitable spawning locations are less common in impounded waters, thus movement distances linked with the annual breeding cycle are greater
- Mary River cod have a tendency to move upstream and / or from the Mary River to tributaries during spring and summer when stream flow increases, and downstream during winter, with the distances travelled often spanning many kilometres (e.g. up to approximately 30 km; Simpson and Jackson 1996a)
- the southern shortfin eel, longfin eel, Australian bass, empire gudgeon, flathead gudgeon and mullet species are diadromous, with spawning occurring in estuarine or marine waters
- the flyspecked hardyhead, crimson-spotted rainbowfish, Agassiz's glassfish, carp gudgeon, dwarf flathead gudgeon, bony bream and Australian smelt are potamodromous, These species use various cues for migration, including increased water temperature and rise in river discharge, and
- mouth almighty, silverstreak hardyhead and eel-tailed catfish have limited migration.

The total length of each fish caught⁷ for each assessment zone for each survey is shown by species in Appendix G. Results show:

- a gradual increase in fish lengths for most species in each assessment zone, indicating a healthy population with juveniles, sub-adults and adults present
- a steep increase in the length of bony bream caught in the reservoir zone, suggesting that this species can grow to a larger size in reservoir habitat and individuals of intermediate size are uncommon, and
- a trend with a strong inflection point (i.e. gradual incline that changes to a steep incline) for some species, suggesting the presence of adults of different size (i.e. some relatively very large individuals were present). For some species (e.g. bony

⁷ Length data was graphed only for species that were caught in sufficient numbers to assess trends.

bream), this may be related to the age of the fish or the habitat within which it was caught (see previous point), but for other species (e.g. eastern Gambusia and carp gudgeons) it may be linked to sexual dimorphism of adult fish (i.e. where adult males are larger than adult females for carp gudgeons, and adult females are larger than adult males for eastern Gambusia; (Allen et al. 2002).

Spawning Mary River cod have been recorded from Six Mile Creek downstream of Lake Macdonald (Dunlop 2016). Consequently, this reach of Six Mile Creek supports a key population of Mary River cod, although inter-annual recruitment success is likely to be variable. Reproduction by Mary River cod in and upstream of Lake Macdonald has not been assessed, although at least 112,730 Mary River cod fingerlings were released to Lake Macdonald between 1983 and 2015 (MRCCA 2016). Reproduction by Australian lungfish in Six Mile Creek is likely to be very limited as preferred breeding habitats are absent, and it is expected that there would be no reproduction in Lake Macdonald (see Kind 2002) or in Six Mile Creek upstream of Lake Macdonald.

Species	Common Name	Caught during field surveys
Native Species		
Ambassidae		
Ambassis agassizii	Agassiz's glassfish	\checkmark
Ambassis marianus	estuary glassfish	_
Anguillidae		
Anguilla australis	southern shortfin eel	\checkmark
Anguilla reinhardtii	longfin eel	\checkmark
Apogonidae		
Glossamia aprion	mouth almighty	\checkmark
Atherinidae		
Craterocephalus marjoriae	silverstreak hardyhead	_
Craterocephalus stercusmuscarum	flyspecked hardyhead	\checkmark
Eleotridae		
Gobiomorphus australis	striped gudgeon	_
Hypseleotris spp.	common gudgeons	\checkmark
Hypseleotris compressa	empire gudgeon	_

Table 4.10Fish species known from, or likely to occur in Six Mile Creek.

Species	Common Name	Caught during field surveys
Mogurnda adspersa	purple spotted gudgeon	\checkmark
Philypnodon macrostomus	dwarf flathead gudgeon	\checkmark
Philypnodon grandiceps	flathead gudgeon	\checkmark
Melanotaeniidae		
Melanotaenia duboulayi	crimson-spotted rainbowfish	\checkmark
Percichthyidae		
Maccullochella mariensis	Mary River cod	\checkmark
Percalates novemaculeata	Australian bass	\checkmark
Macquaria ambigua	yellowbelly	\checkmark
Plotosidae		
Tandanus tandanus	eel-tailed catfish	\checkmark
Neosilurus hyrtlii	Hyrtl's tandan	_
Pseudomugilidae		
Pseudomugil signifer	Pacific blue-eye	\checkmark
Retropinnidae		
Retropinna semoni	Australian smelt	\checkmark
Clupeidae		
Nematolosa erebi	bony bream	\checkmark
Ceratodontidae		
Neoceratodus forsteri	Australian lungfish	\checkmark
Mugilidae		
Trachystoma petardi	pinkeye mullet	_
Mugil cephalus	sea mullet	_
Tetrarogidae		
Notesthes robusta	bullrout	_
Terapontidae		
Leiopotherapon unicolor	spangled perch	\checkmark
Osteoglossidae		
Scleropages leichardti	southern saratoga	√

Species	Common Name	Caught during field surveys
Pest Species		
Gambusia holbrooki	eastern Gambusia	\checkmark
Xiphophorus maculatus	platy	_
Xiphophorus hellerii	swordtail	\checkmark
Poecilia reticulata	guppy	_
Oreochromis mossambicus	tilapia	✓ a

^a observed only.

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
Ambassis agassizii	Agassiz's glassfish	mid to upper catchment	submerged aquatic plants, fine substrates (sand/gravel) and open water	low gradient, slow moving, moderate depth pools	spring to autumn, triggered by rising temperatures	potamodromous
Ambassis marianus	estuary glassfish	lower catchment; brackish estuaries, tidal creeks and lower reaches	mangrove roots, woody debris and aquatic vegetation	_	_	_
Anguilla australis	southern shortfin eel	lower to mid catchment	fine substrates (sand/gravel), leaf litter, woody debris, undercut banks and submerged roots	shallow, still water pools	outward migration during summer and autumn, spawning between June and September	catadromous
Anguilla reinhardtii	longfin eel	entire catchment, commonly lower to mid	no discernible habitat preference	prefers flowing water including riffles and runs	outward migration during summer and autumn, unknown spawning time	catadromous

Table 4.11 Summary of literature review of habitat requirements of native fish species known or likely to occur in Six Mile Creek ^a.

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
Glossamia aprion	mouth almighty	lower to mid catchment	large rivers, limited riparian cover, fine substrates (sand/gravel), submerged macrophytes	deep slow-flowing pools	spring and early summer	limited migration
Craterocephalus marjoriae	silverstreak hardyhead	entire catchment, commonly mid to upper	intermediate substrates (gravel/cobble), submerged macrophytes, filamentous algae, leaf litter, undercut banks and submerged roots	moderate depth and velocity, but also occurs in shallow riffles with high velocity	late winter to summer, with increased water temperature	limited migration
Craterocephalus stercusmuscarum	flyspecked hardyhead	entire catchment	macrophyte beds and in-stream cover	no discernible flow regime, prefers moderate current velocities, seen to congregate where streams flow into still water	late winter to summer with increased water temperate	potamodromous
Gobiomorphus australis	striped gudgeon	lower to mid catchment including estuaries and dune lake systems	fine substrates (sand/gravel), leaf- litter, undercut banks and submerged roots	pools and runs with low velocity and moderate depth	autumn and winter	amphidromous

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
<i>Hypseleotris</i> spp.	common carp gudgeons	entire catchment, commonly lower to mid	fine to intermediate substrates (sand/gravel), submerged aquatic plants, undercut banks, submerged roots	narrows streams with low flow and moderate depth (0.43 m)	late winter to early autumn, prior to high flows	potamodromous
Hypseleotris compressa	empire gudgeon	lower catchment in coastal rivers and streams	fine substrates (mud, sand, gravel), leaf litter, undercut banks and submerged roots	moderate streams (5 m wide, with low flow and moderate depth (0.35 m)	January to May with increased water temperature	amphidromous
Mogurnda adspersa	purple spotted gudgeon	entire catchment, commonly coastal uplands	fine to intermediate substrates (sand, gravel, cobble), aquatic plants, filamentous algae, leaf litter, submerged roots and undercut banks	pools with low velocity and low to moderate depth	spring to late summer	limited migration
Philypnodon macrostomus	dwarf flathead gudgeon	entire catchment, commonly lower to mid-upper catchment	intermediate substrates (gravel, cobble), submerged plants, leaf litter, undercut banks, submerged roots	low flow, moderate depth (0.31 m) but can occur in shallow, high velocity riffle habitats	spring to autumn, with increasing water temperature	amphidromous

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
Philypnodon grandiceps	flathead gudgeon	entire catchment, commonly lower to mid catchment	intermediate to coarse sediment (gravel, cobbles, bedrock), aquatic plants, filamentous algae, leaf litter, submerged roots, undercut banks	low flow, moderate depth (0.4 m), but has been classified as a riffle-dwelling species	spring to autumn, with increasing water temperature	amphidromous
Melanotaenia duboulayi	crimson-spotted rainbowfish	entire catchment, commonly mid catchment	fine to intermediate substrate (sand, gravel), submerged plants, filamentous algae, leaf litter, undercut banks, submerged roots	low-moderate flow, moderate depth (0.43 m), but occasionally in shallow riffles with high water velocity	late winter to summer with increasing water temperature	potamodromous
Percalates novemaculeata	Australian bass	entire catchment, males predominantly estuarine and females in lagoons and upstream	submerged woody debris, undercut banks and overhanging vegetation	slow, deep (>2 m) pools	June to August with elevated discharge	catadromous
Maccullochella mariensis	Mary River cod	entire catchment, but now thought to be restricted to three isolated mid catchment regions	mud/clay substrate, woody debris and log jams, extensive overhanging vegetation, undercut banks, rock ledges	slow, deep pools	annually around spring when temperatures rise above 20°C	potamodromous

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
Tandanus tandanus	eel-tailed catfish	entire catchment, mostly mid to upper catchment	variety of habitats but prefers intact low to moderate riparian zone, diverse substrates (sand, gravel, cobble), leaf litter, undercut banks	low flow streams with moderate depths (0.4m)	spring and summer with increasing temperature	limited migration
Neosilurus hyrtlii	Hyrtl's tandan	entire catchment	uses virtually every aquatic habitat except estuarine reaches, commonly from areas with muddy or sandy substrates, leaf litter, submerged plants and blue-green algae	slow, deep (>2m) pools	summer wet season, with increasing water levels	potamodromous
Pseudomugil signifer	Pacific blue-eye	entire catchment, commonly in the lower catchment	intermediate substrates (gravel, cobble), leaf litter, submerged roots, undercut banks, submerged plants and filamentous algae	common in streams of intermediate width (6-10m) and low to moderate flow.	late winter to late summer, with increasing water temperature	no migration pattern is known

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
Retropinna semoni	Australian smelt	entire catchment, commonly lower to mid catchment	intermediate to coarse substrate (gravel, cobble), aquatic plants, filamentous algae	shallow (0.23m), high velocity riffles and runs, but is found in deeper slow flowing pools during periods of low flow	winter to summer, with low water temperatures	potamodromous
Nematolosa erebi	bony bream	lower to mid catchment, although known to survive in upper catchment conditions when translocated	no discernible habitat known, instead thought to refuge with fellow fish	most common in lowland channel lagoons, with juvenile fish common in open shallow areas (30- 150cm) and adults in deeper waters	all year except June to August (the coldest months)	potamodromous
Neoceratodus forsteri	Australian lungfish	lower to mid catchment, but restricted to freshwater	mud, sand and gravel substrates, overhanging vegetation, submerged woody debris and dense macrophyte beds. Tends to avoid open water	slow-flowing rivers and still water reservoirs and deep pools	August to December with peak activity within three months of the winter solstice.	potamodromous
Trachystoma petardi	pinkeye mullet	lower catchment, predominantly in estuarine waters	-	deep pools or gently flowing sections of rivers	summer	catadromous

Species	Common Name	Catchment Position	Physical habitat	flow-habitat	Reproductive Ecology	Migration Pattern
Mugil cephalus	sea mullet	lower catchment, predominantly in estuarine waters	varies on life history stage, larva are initially pelagic in estuarine water, juveniles prefer shallow water with sandy substrate and adults have no discernable preference	predominantly tidally influence slow-flowing rivers, but can migrate upstream into fast- flowing tributaries	autumn to winter, stimulated by decreased water temperature	catadromous
Notesthes robusta	bullrout	lower catchment, predominantly within 50km of the river mouth	intermediate substrate (gravel/cobble) and in-stream cover	riffle/run habitats with a depth of 0.5m and flow less than 0.3 m/s	possibly winter to spring	catadromous
Leiopotherapon unicolor	spangled perch	lower to mid-upper catchment,	fine substrate (sand and gravel), predominantly near undercut banks and submerged woody debris	little to no flow at depths between 0.3 to 0.6 m	summer during the wet season with rising water temperature	potamodromous

^a sources of information:, Fishbase (2010), Pusey et al. (2004), DAF (2015), DoE (2015)

- denote: no known information on the species

4.6 Turtles

Assessment Methods

In addition to the data and information sources described above, the turtle assessment included a review of Limpus (2008) 'Freshwater turtles in the Mary River: review of biological data for turtles in the Mary River, with emphasis on *Elusor macrurus* and *Elseya albagula*'.

The field survey methods used to collect the turtle data that was collated for this study are presented in Appendix H.

Results

The Mary River has high diversity (six species) and endemism (two regionally endemic species) of freshwater turtles (Limpus 2008):

- Mary River turtle (*Elusor macrurus*) endemic to the Mary River Basin
- white-throated snapping turtle (*Elseya albagula*) endemic to the Mary, Burnett and Fitzroy River Basins
- · Krefft's river turtle (Emydura macquarii krefftii) widespread
- · saw-shelled turtle (Wollumbinia latisternum) widespread
- eastern long-necked turtle (Chelodina longicollis) widespread, and
- · broad-shelled river turtled (*Chelodina expansa*) widespread.

Mary River turtle (endangered) and white-throated snapping turtle (critically endangered) are MNES, listed as threatened species under the Commonwealth's *Environment Protection and Biodiversity Conservation Act 1999* and also under Queensland's *Nature Conservation Act 1992*. Predation of eggs from nesting banks is the most critical threat to the survival of these two species, causing a deficiency of immature turtles and very low recruitment rates in both species, although past harvesting of eggs from Mary River turtle nests has a legacy in the continuing decline of this species (Limpus 2008).

While Six Mile Creek was not surveyed for turtles by Limpus (2008), Mary River turtle and white-throated snapping turtle are known to occur in 'permanent stream and large pool habitats' (Limpus 2008, p.17 and p. 31), whereas the Krefft's river turtle is commonly found in permanent and semi-permanent waterways and man-made waterbodies on ephemeral streams (Limpus 2008, p. 45). Mary River turtle and white-throated snapping turtle were found in several of the larger tributaries of the Mary River, including Tinana Creek, Yabba

Creek and Obi Obi Creek, although white-throated snapping turtle was also found in several additional tributaries and tended to extend further upstream in both tributary streams and the Mary River proper in comparison to Mary River turtle (Limpus 2008). It is therefore possible that both Mary River turtle and white-throated snapping turtle occur infrequently in the lower reaches of Six Mile Creek, with white throated-snapping turtle possibility occurring further upstream than Mary River turtle. All known breeding banks for Mary River turtle and white-throated snapping turtle are along two reaches of the Mary River proper (the first near Tiaro and the second between Traveston and Kenilworth), although white-throated snapping turtle also has several known breeding banks on Yabba Creek (Limpus 2008). Previous survey work for the Project in August and October 2015 found that it is unlikely that there is suitable breeding habitat for either species on Six Mile Creek.

Four of the six turtle species known from the Mary River catchment (Krefft's river turtle, sawshelled turtle, eastern-long neck turtle and broad-shelled river turtle) were caught during the baseline surveys. Only saw-shelled turtle was recorded in Six Mile Creek downstream of Lake Macdonald, with eastern long-necked turtle and Krefft's river turtle recorded from Lake Macdonald, and all four species recorded from upstream of Lake Macdonald (Table 4.12; Appendix I). Saw-shelled turtle and Krefft's river turtle were the most abundant turtle species. Only two juvenile saw-shelled turtles were caught; all other individuals were adults (Appendix I).

Species	Common name	Six Mile Creek DS	Lake Macdonald	Six Mile Creek US
Emydura macquarii krefftii)	Krefft's river turtle		\checkmark	√
Wollumbinia latisternum	Saw-shelled turtle	\checkmark		\checkmark
Chelodina longicollis	Eastern long-necked turtle		\checkmark	\checkmark
Chelodina expansa	Broad-shelled river turtle			✓

Table 4.12 Results of turtle surveys.

4.7 Platypus

Platypus (*Ornithorhynchus anatinus*) is a long-lived (up to 17 years), small, egg-laying, amphibious mammal that is widely distributed in eastern Australia (Grant 1989). It inhabits perennial freshwater streams and connected lentic habitats (including impounded waters), where it spends about 50% of time in water (foraging, moving) and the remaining 50% in one of several short, simple resting burrows (Grant 1989).

Platypus feeds predominantly on benthic invertebrates, with foraging typically occurring at night or dusk, with some individuals foraging during the day in winter. Foraging movements generally within a 1.5 km range, although movements of over 3 km have been recorded, and juveniles are speculated to move over larger distances such as when dispersing from natal areas (Grant 1989).

The breeding season is typically spring, with breeding occurring earlier in norther regions of the species' distribution compared to southern regions. Eggs laid, and young raised, within long (up to 30 m), complex breeding burrows that are maintained by breeding females. Young are weaned after approximately four months and emerge from the burrow in late summer (Grant 1989).

Platypus is a 'Special Least Concern' species under the *Nature Conservation Act* 1992, but is not threatened at National or State levels.

Platypus records from Six Mile Creek and Lake Macdonald were obtained from Atlas of Living Australia (viewed 9 October 2018) and from direct observations of Seqwater rangers. There are confirmed platypus records from Six Mile Creek downstream, within and upstream of Six Mile Dam, including mid-area of Lake Macdonald and within impounded sections of both the Six Mile Creek and Cooroy Creek arms of Lake Macdonald.

4.8 Stygofauna

Stygofauna were assessed using only a desktop approach, including review of relevant literature (e.g. Tomlinson & Boulton 2008; Glanville et al. 2016) and the groundwater study for the Six Mile Creek Upgrade Project (SLR 2018).

Stygofauna are subterranean aquatic animals that live in the pores, voids and cavities of aquifers and other groundwater ecosystems. Many species of stygofauna have specialised adaptations to underground life, including:

- small body size (e.g. many species have a total body length <1 mm)
- · lack of pigmentation
- · absence of eyes, and
- elongated appendages (for tactile sensing of the surrounding environment).

Crustaceans, including copepods, amphipods, isopods and syncarids, typically dominate the composition of stygofaunal communities, although oligochaetes, molluscs, mites,

insects and rotifers are also common. Blind fish and eels are also known from some cave systems, such as those in Western Australia.

Stygofauna taxa are grouped into one of several classes based on the degree of their requirement for subterranean life (Tomlinson & Boulton 2008). For the purpose of this assessment, two classes of stygofauna are considered:

- stygobites: obligate groundwater aquatic fauna that have specialised adaptations to underground life and that live within groundwater systems for their entire life, and
- stygoxenes: aquatic fauna that facultatively use groundwater ecosystems, but are not dependent on groundwater to complete their life cycle.

Stygofauna have no conservation listing at the Queensland level. At the Commonwealth level, the following are listed:

- the Cape Range Remiped (*Kumonga exleyi*) in Western Australia, which is listed as Vulnerable under the Commonwealth's *Environment Protection and Biodiversity Conservation Act 1999* (EPBC Act), and
- stygofauna communities associated with Great Artesian Springs (i.e. the community of native species dependent on natural discharge of groundwater from the Great Artesian Basin), which is listed as Endangered under the EPBC Act.

The Project area is not within the Great Artesian Basin, and is outside the known range of the Cape Range Remiped, therefore listed species of stygofauna do not occur within the Project area.

At the family level stygofauna are widespread and many families that include stygofaunal species also include surface water and marine species (e.g. copepods, rotifers and mites). However, at the species level many taxa have a narrow distributional range (i.e. stygofauna communities contain species that occur exclusively within a small area), and thus stygofauna communities are generally thought to have high endemism (Boulton et al. 2010; Harvey et al. 2011). For example, about one quarter (i.e. 23%) of the stygofauna species that were sampled on two or more occasions in a long-term study of stygofauna in the Pilbara region were sampled from within the same sub-region, and the median area of distribution of these stygofaunal species was 683 km² (Halse et al. 2014), with almost all stygofaunal species having distributions of less than 1 000 km² (Eberhard et al. 2009). A species of Parabathynellidae was recorded from 3 bores in the Burdekin River Alluvial Aquifer in Queensland, with two of these bores located approximately 20 km apart (Cook et al. 2012), suggesting a potential distribution of approximately 400 km². Additionally, studies in both Western Australia and Queensland have found evidence that sub-catchment boundaries can demarcate locations of turn-over of stygofaunal species (Finston et al. 2007;

Little et al. 2016). Therefore, areas of approximately 400 – 600 km² within a single subcatchment may represent reasonable estimates of distribution of most stygofaunal species, acknowledging that site-specific factors (e.g. highly confined aquifers) may impose further restrictions on distribution in some cases, or create strong population subdivision within species on smaller spatial scales (Cook et al. 2012; Little et al. 2016).

A total of 24 described families and 23 described genera have been recorded from Queensland across numerous bioregional areas (Glanville et al. 2016), with representatives of these taxa known from South East Queensland (Glanville et al. 2016; Little et al. 2016). This suggests that stygofauna are likely to be present in the Project area where the habitat of groundwater ecosystems is suitable for their occurrence. The suitability of a groundwater ecosystem to provide habitat for stygofauna is dependent on several environmental factors including:

- geology stygofauna have the potential to occur in aquifers composed of any geological unit with sufficient pore space to complete their life cycle (Tomlinson & Boulton 2008); thus, stygofauna are most common in alluvium, granite, gravel, sand, sandstone, silt, and volcanic geological units (Glanville et al. 2016). Stygofauna are less likely in geological units with relatively small pore spaces, such as those dominated by mudstone, siltstone and clays.
- groundwater hydrology in alluvial aquifers in eastern Australia the average number of stygofauna taxa was higher within 6 m from the water table height, and where the water table height was less than approximately 15 m below the ground (Hancock & Boulton 2008), although stygofauna have been recorded from over 60 m below ground (Glanville et al. 2016), indicating that deep groundwater ecosystems can also support stygofaunal communities.
- · groundwater quality:
 - The mean electrical conductivity of water from which stygofauna have been sampled is less than 4,000 µS/cm, although they have been recorded from a broad range of electrical conductivities (i.e. 11.5 54,800 µS/cm) (Glanville et al. 2016), Tolerance to high electrical conductivity is likely to vary among taxa, with only crustaceans (i.e. copepods and syncarids) reported from the upper end of this range (Glanville et al. 2016)
 - The minimum concentration of dissolved oxygen needed to support stygofauna communities is unknown. Some taxonomic groups are likely to be more tolerant of very low dissolved oxygen, and others more tolerant of very high dissolved oxygen (Halse et al. 2014). However, bores with the highest diversity of stygofauna had dissolved oxygen levels ranging from approximately 20 to 60% saturation (Halse et al. 2014).

- Stygofauna have been recorded from groundwater with pH ranging from 3.5 to 10.3, with diversity highest when pH is between 6.5 and 7.5 (average of 7.0) (Hancock & Boulton 2008).
- Total dissolved solids (TDS) can strongly influence the diversity of stygofauna, with stygofauna almost always absent where TDS is higher than 15 mg/L (Halse et al. 2014).
- Other water quality parameters, such as ionic composition, may also influence the diversity and taxonomic composition of stygofauna (Halse et al. 2014).

The groundwater study for the Six Mile Creek Dam Upgrade Project (SLR 2018) indicated that the:

- Geological units underlying Lake Macdonald, include quaternary alluvium approximately 3 – 21 m deep with high clay content overlaying weathered Triassic Myrtle Creek beds composed of weathered sandstone. Outcrops of Triassic Kin Kin bedrock composed of shale and mudstone occur to the east of Six Mile Creek Dam.
- Hydrology of the underlying geological units is characterised by:
 - low hydraulic conductivity in the alluvium due to high clay content
 - higher hydraulic conductivity of underlying sandstone
- Water quality of groundwater of:
 - alluvium (SMEC 2018):
 - pH: 5.39 6.59
 - dissolved oxygen: 57.4 62.3 mg/L
 - electrical conductivity: 86 140 µS/cm
 - TDS: 59 160 mg/L
 - Kin Kin sandstone considered potable to slightly brackish (SLR 2018).

In summary, the desktop assessment indicated that the alluvium of the Project area is *unlikely to be suitable* for stygofauna due to high clay content, low hydraulic conductivity and high TDS, and the Kin Kin sandstone is *suitable* for stygofauna due to the higher hydraulic conductivity.

4.9 Aquatic Ecology Values Assessment

Assessment Method

The aquatic ecological value of Six Mile Creek was assessed using the criteria in Table 4.13.

Aquatic Ecological Value	Criteria / Description
Very High	Known occurrence and breeding of aquatic MNES and / or threatened species protected under Queensland <i>Nature Conservation Act 1992</i>
High	Known or likely occurrence of aquatic MNES and / or threatened species protected under Queensland <i>Nature Conservation Act 1992</i> and / or HEV Waters under the EPP(Water)
Moderate	Aquatic MNES, threatened species protected under Queensland <i>Nature Conservation Act 1992</i> and HEV waters unlikely to occur, but suitable habitat for non-listed aquatic species of turtles and fish is present. Regulated vegetation categories under the <i>Vegetation Management Act 1999</i> may be present in riparian areas, and watercourses may be important for fish passage under the Queensland <i>Fisheries Act 1994</i> (mapped as having higher than low risk of impact to fish passage by waterway barriers)
Low	Ephemeral watercourse without refugial pools; limited aquatic habitat features present; likely to provide low quality habitat for non-listed aquatic species during high flow events only
Negligible	Site is a drainage feature as per the definition in the Water Act 2000

 Table 4.13
 Criteria used to assess environmental value of each site.

Results

Aquatic MNES (i.e. Mary River cod and Australian lungfish) and platypus are known from Six Mile Creek downstream of Lake Macdonald, and it is possible that Mary River turtle and white-throated snapping turtle occur in the lower reaches of Six Mile Creek. Mary River cod are known to breed in Six Mile Creek downstream of Lake Macdonald. MSES downstream of Lake Macdonald include: waterways providing fish passage, several categories of regulated vegetation (i.e. category R vegetation and vegetation intersecting a watercourse) and HEV (watercourse) waters (4.3 and 4.4).

Within Lake Macdonald, and upstream of Lake Macdonald, platypus are known to occur and likely breed, Mary River cod and Australian lungfish may occur but are unlikely to be breeding, and it is likely that Mary River turtle or white-throated snapping turtle would be rare or absent from Lake Macdonald or upstream of Lake Macdonald. MSES upstream of Lake Macdonald include: waterways providing fish passage, several categories of regulated vegetation (i.e. category R vegetation) and HEV (watercourse) waters (Map 4.2, 4.3 and 4.4).

Using the criteria presented in Table 4.13, the assigned ecological value of:

- Six Mile Creek downstream of Lake Macdonald is very high
- · Lake Macdonald is high
- · Six Mile Creek upstream of Lake Macdonald is high, and
- groundwater ecosystems of the Project area is *low* (i.e. no habitat for conservation significant species).

5 Six Mile Creek Dam Safety Upgrade Project Description

5.1 Project Description

The Project will incorporate the demolition of the existing spillway and embankments and the construction of a new spillway and embankments to improve the safety and performance of the dam to meet current Queensland dam safety regulations. The new spillway will be an uncontrolled dual height labyrinth weir, with the capacity, FSL and inundation area remaining the same as the existing dam following completion of the Project (Table 5.1).

Design Feature	Existing Dam	Upgraded Dam	
Spillway type	Uncontrolled fixed ogee crest	Uncontrolled dual height labyrinth	
Spillway Description	Concrete slab broad crest weir	Mass concrete dual height, multiple cycle labyrinth weir	
Spillway crest elevation (low	Notch/initial: RL 95.32 m AHD	Initial: RL 95.32 m AHD	
level)	Full width: RL 95.35 m AHD	Full width: RL 95.40 m AHD	
Spillway crest elevation (high level)	Not applicable	RL 97.1 m AHD	
Stilling basin floor elevation	RL 83.5 m	RL 86 m	
Energy dissipation method	Plunge pool/stilling basin	Plunge pool/stilling basin	
Full supply level	8,018 ML	8,018 ML	
Dead storage	RL 87.7 m	RL 87.7 m	
Historical No Failure Yield	7,118 ML/y	7,118 ML/y	
Maximum depth	10.5 m	10.5 m	
Area inundated a FSL	260 ha	260 ha	

 Table 5.1
 Design features of existing and upgraded dam.

The major phases of the Project are:

 Drawdown, whereby Lake Macdonald will be drawn-down in stages over a twelveweek period to RL 89 m AHD prior to construction (Figure 5.1). Water levels will be allowed to increase to 89.5 m AHD once construction of the coffer dam is completed (i.e. water levels will be maintained at 89 m AHD for approximately two months), and will be managed within the range of 89-89.5 m AHD during construction. The lake will be drawn down by large scale pumping from the reservoir using a pontoon based

pump station supported by a temporary power supply on the left abutment or adjacent to the right embankment. A pipe system will transfer the water to the existing dam spillway for aeration and energy dissipation before flowing to Six Mile Creek.

- Construction, which will involve:
 - construction of the temporary coffer dam over approximately the first two months of the construction phase, to manage water in Lake Macdonald between 89 m AHD and 89.5 m AHD, and provide a work platform for the main construction phase. The coffer dam will comprise a single row of sheet piles driven into the upstream face of the existing spillway embankment. The design will incorporate a low flow notch no lower than 89.5 m AHD to accommodate low flows, an upper flow level no lower than 90.0 m AHD that is designed to overtop during flood events, and a no-overflow level no lower than 92 m AHD to protect embankment excavations. A width of 30 m has tentatively been adopted for the low flow notch, with position of the notch potentially to be moved during the construction phase by cutting / repairing sheet piles to allow flows to be directed appropriately for site conditions at the time.
 - demolition of the existing dam and spillway, which will be undertaken using rock breakers and excavators. Inert materials from the demolished spillway will be re-used where possible, likely as fish habitat structures to be used in the inundation area.
 - construction of the new dam over approximately 18 to 24 months, which will involve staged construction of the spillway base, spillway wing walls, left and right embankments, outlet tower and labyrinth spillways, and saddle dam
- **Refill and Operation**, whereby the dam will be allowed to be filled to FSL by natural inflows, and normal dam operations, including water supply and provision of environmental flows according to the ROP, re-commence.

The construction period is currently programmed for between August 2019 and October 2021 subject to obtaining approvals and a satisfactory water security situation. Drawdown of Lake Macdonald is currently proposed to begin in May 2019, but this timing may be subject to change. During the construction phase, recreational activities will be temporarily ceased, and the Gerry Cook Hatchery temporarily re-located although some hatchery facilities will be retained through construction and used to support management of Mary River cod and Australian lungfish during the Project.



Figure 5.1 Drawdown schedule.

5.2 Sources of Potential Impact on Aquatic Ecological Values

Review of the Project Description has identified the following sources of potential impact to the aquatic Environmental Values of Six Mile Creek and Lack Macdonald:

- Impacts to water quality in Lake Macdonald and downstream of the lake during the drawdown, construction and refill phases
- · Impacts to aquatic habitat in Lake Macdonald and downstream of the Lake during the drawdown and construction phases
- Impacts to aquatic fauna (injury, mortality or stranding) in Lake Macdonald during the drawdown and construction phases
- Impacts to aquatic flora in Lake Macdonald and downstream of the lake during the drawdown and construction phases
- · Spread of biosecurity matters downstream of Lake Macdonald during the drawdown and construction phases
- Impacts to stygofauna communities in shallow groundwater systems during drawdown and construction phases, and
- · Barriers to fish passage at the dam wall during the refill and operation phases.

6 Risk and Mitigation Assessment

6.1 Impact Assessment Methods

The risks of, and mitigations for, each of the identified potential sources of adverse impact of the Project on the Environmental Values of Six Mile Creek was assessed using a risk-based approach, with the risk score for a potential impact calculated by multiplying the likelihood and consequence scores for that potential impact (Table 6.1, Table 6.2, Table 6.3). The 5 x 3 risk matrix (Table 6.3) gives risk scores ranging between one and 15, with risk being:

- \cdot low, when the score is <5
- medium, when the score is >5 but <10, and
- high, when the score is >10.

Table 6.1	Ratings used to assess the likelihood of potential impacts.
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Rating	Likelihood of occurrence
Very high (5)	Almost certain to occur frequently
High (4)	Probably would happen sometimes to frequently
Moderate (3)	Could happen sometimes
Low (2)	Remote possibility of occurring or not expected to occur
Very low (1)	Definitely would not happen at all

Table 6.2Ratings used to assess the consequence of potential impacts.

Rating	Consequence of occurrence
High (3)	Catastrophic short-term or significant long-term / irreversible environmental harm; significant short- or long-term harm to protected components of the environment, such as MNES.
Moderate (2)	Significant short-term but reversible environmental harm; minor environmental harm to protected components of the environment, such as MNES.
Low (1)	Unfavourable impact with no material harm to the environment; no impact to protected components of the environment.

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		Likelihood				
		Very Low (1)	Low (2)	Moderate (3)	High (4)	Very High (5)
nce	Low (1)	1	2	3	4	5
enbes	Moderate (2)	2	4	6	8	10
Con	High (3)	3	6	9	12	15

Table 6.3Environmental risk matrix, showing risk score and level of risk (green = low;
amber = moderate, red = high).

Assessment of potential impacts to Mary River cod, Mary River turtle, Australian lungfish and white-throated snapping turtle were further assessed using the MNES Significant Impact Criteria Guidelines 1.1.

6.2 Risk-based Impact Assessment

Water Quality

Potential Impacts

Drawdown of waterbodies can have adverse impacts on water quality both at, and downstream of, the discharge site(s) and in the lake. Construction earthworks, and runoff from stock piles of soil, during construction can also adversely impact water quality. Submersion of decomposing organic matter can also adversely affect water quality during dam refill phases and lead to eutrophication. The potential impacts to water quality during the drawdown, construction and refill phases include:

- increasing turbidity and total suspended solids via disturbance of bed sediments and / or the erosion of bed and banks during drawdown and construction; and from disturbance of earth and runoff from soil stockpiles during construction
- reducing pH by exposing or disturbing acidic soils during drawdown and construction and / or where decomposing organic material (e.g. aquatic plants) reduces the pH of water
- reducing dissolved oxygen in the lake and in Six Mile Creek downstream if the source waterbody becomes stratified or eutrophied, such as through submersion of decomposing organic matter (e.g. decomposing Cabomba) during the refill phase

- increasing nutrient concentrations in the lake if drawdown exposes deep sediments below approximately 90 m AHD, which have higher nutrient content than sediments above 90 m AHD (Grinham et al., 2018), and in receiving waters during drawdown if the source water has high nutrient concentrations, or during refill if the lake becomes eutrophied from decomposing organic matter
- increasing dissolved metal concentrations in the lake and receiving waters during drawdown, construction and refilling phases due to mobilisation and oxidation of lake sediments, lateral transport of sediment pore water and ebullition fluxes during drawdown (Grinham, et al., 2018). Drawdown that exposes deep sediments below approximately 90 m AHD, which have higher metal content than sediments above 90 m AHD, will increase the risk of adverse water quality (Grinham et al., 2018), and
- contaminating water from spills of fuels, oils or other chemicals from pumping equipment or other machinery / vehicles during drawdown and construction.

Increased turbidity (and total suspended solids) may negatively impact fish and macroinvertebrates, because highly turbid water reduces respiratory and feeding efficiency. Increased turbidity may also adversely affect submerged aquatic plants as light penetration (required for photosynthesis) is reduced. Reduced light penetration can also lead to a reduction in temperature throughout the water column. Small and brief increases in turbidity, consistent with increases in turbidity that occur during natural flow events, would be unlikely to have a significant impact on aquatic fauna (Dunlop et al., 2005). However, significant increases in turbidity caused by fine silt and clay particles, could adversely impact the health, feeding and breeding ecology of aquatic fauna species (Dunlop et al., 2005). However, significant resuspension of sediments is likely to be limited by cohesive properties of the sediment in the Six Mile Creek Dam (Grinham et al., 2018).

Reduced pH can negatively impact fish health by causing diseases (e.g. lesions and ulcers) and impacting metabolism and reproduction in fish, with very low pH (such as from acidic soil exposure) potentially causing fish kills. While many waterways of the lower Mary River Basin are naturally acidic and stained with tannins and organic compounds, Six Mile Creek is not of this acidic water type; similarly water in Lake Macdonald is tannin stained but is not naturally acidic. Some variation in pH is tolerated by aquatic biota of Six Mile Creek, although significant reductions in pH may have adverse effects on aquatic ecosystem health. The potential for acid sulfate soils in the Project area was assessed and determined to be unlikely (SMEC 2018).

Dissolved oxygen is essential for respiration and metabolism by aquatic biota. Reduced dissolved oxygen can cause stress to fish, and very low dissolved oxygen can cause mass mortality ('fish kills'). Some waterways of the region can have naturally low dissolved oxygen, especially during low flow periods, and thus much of the aquatic biota of the region

can tolerate periods of low dissolved oxygen (i.e. approximately 50 percent saturation), but sustained periods of low dissolved oxygen and / or very low dissolved oxygen will cause mortality in aquatic fauna.

High nutrient concentrations can cause increased growth of phytoplankton, which in turn can deplete dissolved oxygen concentrations, particularly at night when there is no photosynthesis. Benthic algae, including filamentous algae, and aquatic plant growth may increase under high nutrient conditions, especially under high sunlight conditions. Excessive algae and aquatic plant growth can reduce in-stream habitat quality for some aquatic biota.

Fuels, oils and other chemicals (e.g. lubricants and solvents) that may be required for the operation of pumps and other machinery for lake drawdown, including vehicles, are toxic to aquatic flora and fauna at relatively low concentrations. Spilt fuel is most likely to enter watercourses via an accidental spill when activities are adjacent to waterbodies. A significant fuel spill to waterways (in the order of tens or hundreds of litres) is likely to have a locally significant impact on both flora and fauna, with the size of spill and the volume of water in the creeks being the most significant factors influencing the length of stream impacted.

Mitigations

Appropriate mitigation measures should be implemented for the Project to achieve the following objective:

· prevent or reduce potential impacts to water quality.

Suggested mitigation measures are provided below, however alternative measures may also be appropriate to achieve the objective:

- Minimise exposure of deep sediments that have high metal and nutrient concentrations, for example by not lowering water in Lake Macdonald to below 89 m AHD
- Minimise disturbance of unconsolidated bed sediments, which could increase turbidity and TSS, and organic matter on the bed of Lake Macdonald, which could influence dissolved oxygen concentrations and pH (e.g. by using pontoon based pump stations)
- Reduce the likelihood of releasing poor quality water to Six Mile Creek from Lake Macdonald during the drawdown by taking water from mid-depth, or mix of depths, for example with a multi-level intake

- Avoid or manage areas of potential erosion, for example by implementing an Erosion and Sediment Control Plan (ESCP) in accordance with applicable industry standards, monitoring the efficacy of sediment and erosion control management measures, and/or releasing water to the existing concrete apron during drawdown
- Maintain dissolved oxygen concentrations in Lake Macdonald and Six Mile Creek, for example by using aeration units (potentially including the existing destratification unit) within the lake and turbulent release to the existing concrete apron.
- Remove decomposing Cabomba from exposed lake surfaces, where possible, to reduce risk of eutrophication and low dissolved oxygen during refill phase
- Implement real-time water quality monitoring for comparison against suitable objectives for key parameters (i.e. pH, dissolved oxygen, turbidity, TSS, nutrients). The objectives should be consistent with the desired outcomes and trigger corrective action, such as a review and update of existing control measures, if exceeded.
- Reduce the likelihood of chemical spills or leaks, for example through:
 - storing fuels, oils and other chemicals in bunded areas in accordance with Australian Standard 1940 (2004) – The storage and handling of flammable and combustible liquids
 - establishing bunded areas away from water bodies, preferable above the Q100 level
 - only refuelling in bunded areas, and
 - making spill kits available to enable a rapid response to a spill if one was to occur.

Risk Assessment

The *consequence* of impacted water quality is *moderate* because large changes in some water quality parameters, such as drops in pH or the concentration of dissolved oxygen, may have adverse effects for aquatic biota.

The *likelihood* of impacts to water quality is *low* where appropriate mitigation measures such as those described above are implemented, because dissolved oxygen concentrations will be maintained, turbidity and TSS are less likely to increase, and other water quality parameters will be monitored and managed as required (see above).

The mitigated *risk* of impact to water quality is *low*.

Aquatic Habitat

Potential Impacts

Aquatic habitat in the lake will be adversely affected by lowered water levels during the drawdown phase, with the amount of aquatic habitat (i.e. volume of water) in Lake Macdonald reduced to approximately 2.8% of FSL (27 hectares) for the first two months of construction, and allowed to increase to approximately 5.1% of FSL (42 hectares) for the remainder of the construction period. This is a significant temporary reduction in aquatic habitat that will be reversed during the refill and operational phases of the Project.

There may also be potential adverse impacts to aquatic habitat downstream of the lake due to sedimentation and hydrological changes during drawdown and construction phases:

- Sedimentation Fine sediments accumulate on the bed of reservoirs (e.g. >90% of sediment is captured in Six Mile Creek Dam, DNRM 2014), which could be mobilised during drawdown and construction phases and deposited downstream (i.e. sedimentation). Sedimentation smothers benthic habitats, including in-filling pools and interstitial spaces of coarse substrate (e.g. gravels and cobbles), causing cascading impacts to primary producers (i.e. aquatic plants and benthic algae), macroinvertebrates and fishes (Wood and Armitage 1997).
- Potential hydrological impacts The drawdown phase will create a 'flow event' that is of similar magnitude to an ARI 2 year flow, although the duration of this flow will be significantly longer than a natural flow event of similar magnitude. The frequency and duration of low flows will likely increase from current conditions during the construction period, as the reduced dam wall height provides less capacity to reduce or buffer outflows, and thus a greater proportion of in-flows will flow downstream to Six Mile Creek downstream of Lake Macdonald. However, the magnitude, frequency, duration and timing of large flows should not change substantially from current, because the nature of Lake Macdonald and it's catchment are such that attenuation of high flows by the current dam are minimal.

Mitigations

Appropriate mitigation measures should be implemented for the Project to achieve the following objective:

· prevent or reduce potential impacts to aquatic habitat.

Suggested mitigation measures for potential impacts are provided below, however alternative measures may also be appropriate to achieve the objective:

- temporary loss of aquatic habitat in Lake Macdonald
 - where possible, augment aquatic habitat within Lake Macdonald (e.g. by adding physical habitat structures; controlling aquatic weeds) during construction to improve the long-term aquatic habitat values of Lake Macdonald from current condition
- · sedimentation
 - minimise disturbance and downstream transfer of unconsolidated bed sediments, for example by using a pontoon based pump station
 - stabilise exposed sediments, for example by seeding the exposed Lake Macdonald bed with non-invasive grasses following initial drawdown
 - avoid or manage areas of potential erosion, for example by implementing an ESCP in accordance with applicable industry standards, and monitoring the efficacy of management measures
 - slow flow and reduce erosion in the upper reaches of the lake, for example by using physical barriers (e.g. staggered baffles) at key upstream locations
- potential hydrological impacts:
 - avoid releases during natural low flow periods, for example undertake drawdown when flows of moderate magnitude commonly occur
 - avoid major pulse flow events, for example by using a maximum pumping / discharge rate that will not exceed the bank full width of Six Mile Creek downstream of the dam and/or pumping / discharging over an extended period (e.g. 12 weeks)
 - avoid changes to hydrology during the breeding seasons for MNES species known to be in Six Mile Creek downstream of the dam, for example undertake drawdown outside the Mary River cod and Australian lungfish breeding seasons
 - maintain the existing flow regime during the construction period, for example by allowing inflows to pass the construction area, pumping water downstream at a rate consistent with inflows, and/or using water piped from the Mary River to the water treatment plant.

Risk Assessment

Temporary loss of aquatic habitat in Lake Macdonald

The *consequence* of temporary loss of aquatic habitat in Lake Macdonald is *moderate*, because non-breeding habitat for Mary River cod, and habitat for platypus, occurs in Lake Macdonald; however the impact on habitat is reversible, with long-term habitat values to be augmented during the project.

The *likelihood* of temporary loss of habitat is *very high* as it is certain that temporary loss of approximately 97.2% of aquatic habitat (by water volume) in Lake Macdonald will occur.

The *risk* of impact of temporary loss of habitat in Lake Macdonald is therefore *high*, and requires further mitigation measures, specifically:

• developing and implementing a comprehensive aquatic fauna salvage operation (see Aquatic Fauna below).

This additional mitigation reduces the *consequence* to *low*, giving a *residual risk* rating of *moderate*. This is an unavoidable risk given the safety requirements of the Project.

Sedimentation

The *consequence* of sedimentation is *moderate* because breeding habitat for Mary River cod occurs in the downstream reaches of Six Mile Creek; however, the impact is reversible, and subsequent large flows will naturally flush sediment after the construction phase is completed.

The *likelihood* of sedimentation is *low* where appropriate mitigation measures, such as those described above, are applied.

The mitigated *risk* of impact of sedimentation is *low*.

Changes to Downstream Flows

The *consequence* of altered *hydrology* is *low*, because the most notable change to hydrology will be a one-off, short-duration event during the drawdown phase, which is recommended to occur outside the breeding season of Mary River cod and Australian lungfish (the MNES species known to be in Six Mile Creek downstream of Six Mile Creek dam).

The *likelihood* of altered hydrology is *moderate*, because there will be a sustained 12-week release during drawdown and the frequency of flows past the construction site may increase compared to current flows past the dam, which is considered a temporary benefit to the downstream environment.

The mitigated *risk* of impact of altered hydrology is *low*.

Aquatic Fauna

Potential Impacts

Aquatic fauna may become injured in pumping equipment during the drawdown phase, which could make them susceptible to pathogens and disease, or be fatally injured, trapped and subsequently drown. Injury may also occur in the low flow notch during the construction phase, and over the spillway during the refill and operate phase.

Aquatic fauna in Lake Macdonald may become stranded in small isolated pools during the drawdown phase once water levels have lowered, and following large flow events during the construction phase (i.e. when water levels rise and then lower again following significant rainfall), which may increase predation (e.g. predation of smaller fish by larger fish and / or birds), and / or crowding. Crowding may result in reduced dissolved oxygen concentrations in water, increased competition for food and shelter, and increased stress on fauna. As small isolated pools evaporate, or in areas that are dewatered rapidly, there is a risk that aquatic fauna could become stranded on dry areas and perish. Turtles and platypus may also have difficulty moving to the nearest available water through exposed lake bed sediments (i.e. mud) and exposed aquatic vegetation.

Some turtle species have an innate biological response to 'walk out' of waterbodies as water levels lower; however, alternate waters may not be sufficiently proximate, and / or movement to alternate water may involve crossing roads and encountering other hazards.

During the refill and operation phase, injury or mortality to stream fauna could occur by downstream passage over the labyrinth spillway, with the vertical drop to the surface over a vertical fall spillway to the surface below being the predominant source of risk (Berghuis 2017). A vertical fall from a spillway to a deep plunge pool or stilling basin can cause significant injuries to fish when the fall higher is greater than 13 m (Berghuis 2017), with vertical falls to hard surfaces (e.g. concrete, rock) or shallow water likely to result in serious injury at much lower fall heights. The proposed dual height labyrinth spillway incorporates stepped plunge pools on the downstream face, ensuring low vertical fall heights, with plunge pools designed to be at least 30 percent of the vertical fall. Furthermore, tail water levels rise rapidly during overtopping flows, and thus it is expected that the stepped plunge pools

will be drowned out in large flow events. However, entrapment of fish in the stepped plunge pools is possible as overtopping flows subside, where they may be susceptible to predation and poor tailwater quality.

Mitigations

Appropriate mitigation measures should be implemented for the Project to achieve the following objective:

• prevent or reduce potential impacts to aquatic fauna, in particular listed threatened species.

Suggested mitigation measures for potential impacts are provided below, however alternative measures may also be appropriate to achieve the objective:

- Prevent aquatic fauna from being entrained and injured or trapped by pumping equipment, for example by using suitably designed screens with aperture <20 mm, and a pumping rate / in-let housing size that ensures that 'approach velocities'⁸ are < 0.1 m/s (see Boys et al. 2012), and/or monitoring measures implemented to ensure they are functioning correctly
- Enable aquatic fauna to move to areas where water will persist for the duration of the Project, for example lowering water levels slowly during the initial weeks of drawdown
- Where possible, maintain suitable habitat for protected species (e.g. by maintaining refugial pools within platypus home range, or reduce the likelihood of impacts to protected species, for example by implementing a comprehensive aquatic fauna salvage operation in accordance with DAF's Fish Salvage Guidelines (DPI 2004), that includes:
 - establishing quantitative targets for recovery of large bodied fishes, turtles and, if necessary, platypus
 - undertaking salvage and relocation until such time that the quantitative salvage objectives have been achieved
 - selecting relocation sites with consideration of biosecurity management, protecting genetic integrity of populations of all turtle species, Mary cod and

⁸ Approach velocity: velocity of water flowing in the vector perpendicular to, and in front of, the screen face. As velocity declines steadily with increasing distance from the screen, approach velocity is measured 8 cm in front of the screen, which is typical of screening guidelines throughout the world.

Australian lungfish, carrying capacity, and ease of re-introduction of key fauna (especially Mary cod and Australian lungfish) back to Lake Macdonald in the refill and operate phase

- basing the number of individuals released at any one location on the assessed carrying capacity to avoid over-stocking / crowding, and monitoring the success of relocation
- undertaking incidental fish salvage after large flow events during construction until the quantitative salvage objectives have been achieved
- monitoring key water quality parameters and observing fauna in Lake Macdonald during construction to trigger incidental fish salvage, or other mitigation measures, as required
- Prevent turtles dispersing over roads, for example by installing temporary fencing, similar to coarse sediment barriers, between Lake Macdonald and roads, and/or daily surveillance and salvage of turtles during the drawdown phase and weekly during the construction phase
- Restore Lake Macdonald, where possible, once water quality assessments have confirmed suitability, for example by re-stocking salvaged Mary River cod and Australian lungfish, and other species, during the refilling and operation
- Reduce the risk of injury and mortality to aquatic fauna over the spillway and stranding of fauna in plunge pools, for example by ensuring the design of the spillway includes low fall heights to sufficiently deep plunge pools and incorporating recommendations from Berghuis (2017).

Preliminary Risk Assessment

Injury to Fauna

The *consequence* of injury or mortality of aquatic fauna where the proposed mitigation is implemented is *moderate*, as the mitigations will prevent significant harm (e.g. mortality of large numbers) to threatened (and other) species of aquatic fauna that occur in Lake Macdonald.

The *likelihood* of injury or mortality of aquatic fauna is *low* where appropriate mitigation measure, such as those described above, are implemented, because they will prevent threatened species from becoming entrapped in equipment, reduce risks of stranding, and

reduce impacts of injury or mortality to fauna over the spillway and stranding of fauna in plunge pools.

The mitigated *risk* of injury or mortality of aquatic fauna is *low*. Further risk assessment will be implemented once detailed designs of the labyrinth spillway are available.

Stranding of Fauna

The *consequence* of fish and turtle stranding is *moderate* because appropriate mitigations will prevent significant stranding of aquatic fauna, but also because the lake will not be completely drained and refugial habitat will persist through the construction period.

The *likelihood* of fish and turtle stranding is *low* where appropriate mitigations, such as those described above, are applied.

The mitigated *risk* of fish and turtle stranding is *moderate*.

Road Kill of Turtles

The *consequence* of road kill of turtles is *moderate* where appropriate mitigations, such as those described above, are applied, because threatened turtle species are unlikely to occur in the Project area and these species are not known to wander to the same extent as many common species of turtle. Appropriate mitigation measures will also reduce potentially large numbers of common species of turtles walking across nearby roads.

The *likelihood* of road kill of turtles is *low* where suitable mitigations, such as those described, are applied, because they will reduce the frequency of turtles from walking across road.

The *mitigated* risk of impact is *low*.

Aquatic Flora

Potential Impacts

Dewatering will expose aquatic plants above 89.5 m AHD, with those species dependent on standing water expected to perish in these areas.

Mitigations

Water will be retained in Lake Macdonald below 89.5 m AHD, providing refugee habitat for aquatic plants.

Risk Assessment

The *consequence* of impact of dewatering on aquatic plants is *low* because no threatened species of aquatic plant is known from the area, and the aquatic plant community of Lake Macdonald is dominated by *Cabomba*, a restricted biosecurity matter. Furthermore, water will be retained in Lake Macdonald below 89.5 m AHD, and aquatic plant species have effective dispersal capabilities ensuring recolonization by aquatic plants during the refill and operate phase.

The *likelihood* of impact of dewatering on aquatic plants is *moderate*, because some native plants can persist in saturated sediments, but at least some species will perish in dewatered areas, but the impact will be reversed during the refill and operate phase.

The *mitigated risk* of impact is *low*.

Spread of Aquatic Biosecurity Matters

Potential Impacts

An invasive species is a species that is found beyond its natural distribution, and which threatens valued environmental, agricultural or other societal resources. Invasive species that have, or have the potential to have, significant adverse impacts on the ecological, agricultural or economic resources of Queensland are declared under the *Biosecurity Act 2014*, as either:

- · a prohibited matter, not yet present in Queensland, or
- a restricted matter, currently present in Queensland.

The damage caused by invasive species includes:

 Competitive impacts, where the invasive species can reproduce more rapidly, or otherwise out-compete native species, to the extent that a native species declines or is threatened. For example, fish such as eastern gambusia (*Gambusia holbrooki*) and tilapia (*Oreochromis mossambicus*) displace native fish, while aquatic weeds such as water hyacinth (*Eichhornia crassipes*) and Cabomba (*Cabomba caroliniana*) can displace native aquatic plants.

- Degradation of habitat, where feeding or other behaviours result in the degradation of habitat that supports native species. For example, aquatic weeds choke waterways and can reduce the concentration of dissolved oxygen in the water, making it unsuitable for native fauna; and tilapia disturb benthic habitats through their breeding behaviour.
- Predatory impacts, where an invasive species reduces the population size and / or threatens the survival of native species by predation.
- Herbivory impacts, where an invasive species consumes native plants, causing a population decline.

Operational activities of the Project have the potential to spread restricted biosecurity matters, most notably:

- upstream movement of tilapia during the construction period at times when the coffer dam is over-topped by large flows, and
- spread of aquatic plant fragments during fauna relocation during the drawdown phase.

Mitigations

Appropriate mitigation measures should be implemented to achieve the following objective:

 prevent or reduce the potential impacts of the Project on the establishment and spread of aquatic biosecurity matters.

Suggested mitigation measures are provided below, however alternative measures may also be appropriate to achieve the objective:

- To the extent that is possible, reduce the opportunities for upstream movement by tilapia during the lake lowering and construction phases, for example by managing the water level in the lake to reduce the potential for drown out / overtopping (i.e. via pumping), not using or pumping water from Six Mile Creek downstream of the dam for construction or machinery purposes
- When caught, euthanise pest fish humanely in accordance with methods approved by animal ethics, for example, if possible, sort fish during any salvage effort before relocation and euthanise pest fish.

- Control potential increases in cane toad populations, for example by installing cane toad traps, as manufactured by the Mary River Catchment Coordinating Committee
- · Where possible, remove Hygrophila and Cabomba from Lake Macdonald during the construction phase
- Minimise potential spread of aquatic weeds, for example by implementing pest identification training for all relevant construction personnel, only relocating aquatic fauna to waterbodies that are already infested with Cabomba, and/or requiring that vehicles, machinery, equipment and temporary infrastructure are subject to weed hygiene protocols.

Risk Assessment

The *consequence* of establishment or spread of biosecurity matters is *high*, as this would represent both an environmental impact and non-compliance with the *Biosecurity Act 2014*, with establishment of invasive species generally non-reversible.

The *likelihood* of establishment or spread of biosecurity matters is *low* where appropriate mitigations, as described above, are implemented.

The *mitigated risk* of impact from spread and establishment of biosecurity matters in *low*.

Stygofauna

Potential Impacts

The Project has the potential to impact local stygofauna communities in underlying shallow alluvial aquifers through:

- Reducing the rate of aquifer recharge from direct infiltration from Lake Macdonald during dewatering and construction phases, which may result in localised reductions in groundwater levels in the order of 8 – 14 m below ground level (SLR 2018), thereby reducing habitat availability for stygofauna in shallow groundwater systems, and
- Contamination from spills of fuels and oils, which may contaminate shallow groundwater ecosystems and cause lethal or sub-lethal impacts to stygofauna.

Mitigations

Appropriate mitigation measures should be implemented to achieve the following objective:

· prevent or reduce the potential impacts to stygofauna in the Project area.

Suggested mitigation measures are provided below, however alternative measures may also be appropriate to achieve the objective:

- Reduce the likelihood of chemical spills or leaks, for example through:
 - storing fuels, oils and other chemicals in bunded areas in accordance with Australian Standard 1940 (2004) – The storage and handling of flammable and combustible liquids
 - establishing bunded areas away from water bodies, preferable above the Q100 level
 - only refuelling in bunded areas, and
 - making spill kits available to enable a rapid response to a spill if one was to occur.

Risk Assessment

The *consequence* of shallow groundwater drawdown, or contamination of shallow groundwater ecosystems, for stygofauna is *low*, because the alluvial aquifer underlying Six Mile Creek Dam is considered unlikely to be suitable habitat for stygofauna. Furthermore, if stygofauna were present in the alluvial aquifer, potential disturbances to stygofauna habitat associated with the Project would be an order of magnitude smaller than the likely distribution of stygofauna taxa (i.e. speculated to be in the order of 400 km² within a single subcatchment area (see Section 4.8).

The *likelihood* of shallow groundwater drawdown, or contamination of shallow groundwater ecosystems, for stygofauna is *low* where appropriate mitigations, as described above, are implemented.

The *mitigated risk* to stygofauna is *low*.

Fish Passage

Potential Impacts

Almost all freshwater fish species migrate at some spatial scale (Harris et al. in press), with migration being the regular cyclic alteration between different habitats used for spawning, feeding or survival (Northcote 1998).

Migration is a key ecological process that enables species to complete their life history by:

- providing access to foraging, residing and spawning / breeding habitats, which may occur in geographically distinct segments of a river system
- · avoiding predation
- · reducing population density and intra-specific competition, and
- enabling individuals to find refuge from seasonal or inter-annual harsh conditions, such as dry seasons and droughts.

Migration also maintains diverse and abundant fish communities, with both diversity and abundance of fish known to decrease where natural migration patterns are impacted; barriers to fish migration represent a significant threatening process for freshwater fish globally (Harris et al. in press). Maintenance of naturally diverse fish communities (via maintenance of fish passage and other factors, such as habitat quality and natural flow regime) maintains natural ecological processes at the location of the fish community (e.g. natural tropic interactions and food web structure) and contributes significantly to maintaining the EVs of the watercourse.

A permanent waterway barrier in a downstream reach may cause significant changes to fish communities along the whole river if diadromous migration is impeded. Multiple waterway barriers may cause cumulative impacts and can severely change aquatic communities. Secondary ecological impacts such increased rates of benthic algal growth, sedimentation and accumulation of organic matter may result from the exclusion of migratory fish and shrimps due to waterway barrier works.

Six Mile Creek is mapped as a 'major risk of impact' (purple) waterway in DAF's waterway barrier works risk layer, indicating that permanent waterway barriers, such as dams, are likely to have a significant impact on fish passage. The absence of Pacific blue-eyes (a diadromous species) upstream of the dam indicates a likely impact to fish passage from the existing dam (frc environmental 2016). However, many species that undertake diadromous migration in Six Mile Creek are often present in relatively low abundance, and a number of diadromous species that would be expected to occur were absent from surveys (e.g. sea mullet, pink eye mullet, empire gudgeon, striped gudgeon), suggesting the possibility of

cumulative impacts to fish passage between the estuary and Six Mile Creek by barriers in the lower Mary River (e.g. Gympie Weir) (see Walker 2008). Providing fish passage at barriers in the lower Mary River will therefore improve the native fish community in Six Mile Creek below the dam, as well as in other sections of the broader Mary River system.

Management of Fish Passage

Fish passage is currently not provided at the Six Mile Creek dam. While investigations into fishway options have been implemented for the Six Mile Creek dam upgrade (e.g. frc environmental, 2016; Seqwater 2018), preliminary advice from DAF indicates that several factors outweigh the benefit of providing fish passage over the upgraded dam, including site constraints, the risk of aiding upstream dispersal by tilapia, and the anticipated regional benefits of the proposed offsite mitigation option (DAF 2018). Additionally, the limited options available at Six Mile Creek Dam for implementing fish passage were generally not attractive with respect to the need for a mechanical fishway and limitations on available water. Where provision of fish passage is not possible due to overriding constraints, DAF may accept fish passage mitigations. The following mitigation measures are suggested for managing fish passage issues for the Project:

- Provision of fish passage at Gympie Weir by installation of a suitable fishway, and
- Removal of the old culvert material from Six Mile Creek immediately downstream of the Six Mile Creek dam, because during low flows this barrier traps fish in the spillway pool and prevents downstream movement.

6.3 Impacts to Aquatic Matters of National Environmental Significance

The Sensitivity of the Environment that will be Impacted

Six Mile Creek is a sensitive environmental receptor as threatened aquatic species inhabit this waterway, and Mary River cod breeds in Six Mile Creek downstream of Lake Macdonald.

The Timing, Duration and Frequency of the Action and its Impacts

The major phases of the Project are:

- **Drawdown**, whereby Lake Macdonald will be drawn down in stages over a twelveweek period to RL 89 m AHD prior to construction (Figure 5.1). Water levels will be allowed to increase to 89.5 m AHD once construction of the coffer dam is completed (i.e. water levels will be maintained at 89 m AHD for approximately two months) and managed between 89 m AHD and 89.5 m AHD during construction. The lake will be drawn down by large scale pumping from the reservoir using a pontoon based pump station supported by a temporary power supply on the left abutment or adjacent to the right embankment. A pipe system will transfer the water to the existing dam spillway for aeration and energy dissipation before flowing to Six Mile Creek.
- Construction, which will involve:
 - construction of the temporary coffer dam over approximately the first two months of the construction phase, to contain water in Lake Macdonald between 89.0 m AHD and 89.5 m AHD, and provide a work platform for the main construction phase. The coffer dam will comprise a single row of sheet piles driven into the upstream face of the existing spillway embankment. The design will incorporate a low flow notch no lower than 89.0 m AHD to accommodate low flows, an upper flow level no lower than 90.0 m AHD that is designed to overtop during flood events, and a no-overflow level no lower than 92 m AHD to protect embankment excavations. A width of 30 m has tentatively been adopted for the low flow notch, with position of the notch potentially to be moved during the construction phase by cutting / repairing sheet piles to allow flows to be directed appropriately for site conditions at the time.
 - demolition of the existing dam and spillway, which will be undertaken using rock breakers and excavators. Inert materials from the demolished spillway will be re-used where possible, likely as fish habitat structures to be used in the inundation area.
 - construction of the new dam over approximately 18 to 24 months, which will involve staged construction of the spillway base, spillway wing walls, left and right embankments, outlet tower and labyrinth spillways, and saddle dam
- Refill and Operation, whereby the dam will be allowed to be filled to FSL by natural inflows, and normal dam operations, including water supply and provision of environmental flows according to the ROP, re-commence.

The construction period is currently programmed for between August 2019 and October 2021 subject to obtaining approvals and a satisfactory water security situation. Drawdown of Lake Macdonald is currently proposed to begin in May 2019, but this timing may be subject to change. During the construction phase, recreational activities will be temporarily

ceased, and the Gerry Cook Hatchery temporarily re-located although some hatchery facilities will be retained through construction and used to support management of Mary River cod and Australian lungfish during the Project.

On-site and Off-site, and Direct and Indirect Impacts

Potential direct and indirect impacts of the Project are likely to affect both Lake Macdonald and Six Mile Creek downstream. Potential sources of impact are described in Section 5.2, and the risk of adverse impact from each of these sources of potential impact has been assessed (Section 6.2). Most sources of impact were assessed as having a low risk of impact when the appropriate mitigations, such as those identified, are applied. However, the temporary loss of aquatic habitat in Lake Macdonald on aquatic fauna still had a moderate residual impact after mitigation measures were applied.

The Total Impact that can be Attributed to the Action

The Project is replacing an existing dam wall; thus, the long-term impact will be no change from current condition, with the potential exception of improved physical habitat for Mary River cod in Lake Macdonald. Most potential impacts will be temporary (i.e. for the duration of the drawdown and construction periods, approximately 2 to 3 years in total), and where appropriate mitigations, as described above, are applied there will be no ongoing impacts to the aquatic environmental values of Lake Macdonald or Six Mile Creek.

Existing Levels of Impact from Other Sources

Land uses in the catchment of Six Mile Creek include forestry, grazing, horticulture, rural residential and urban areas, which have relatively low levels of impact on the aquatic ecology of Lake Macdonald and Six Mile Creek. Aquatic weeds, especially Cabomba and Hygrophila, are notable existing stressors, with tilapia recently recorded from Six Mile Creek downstream of Lake Macdonald. These biosecurity matters will be managed throughout the Project via an Environmental Management Plan.

The Project will have no impact on the scope or scale of these impacts.

The Degree of Confidence with which the Impacts of the Action are Known and Understood

It is considered that potential impacts of the Project on the aquatic ecology of Six Mile Creek have been assessed with a 'moderate to high' degree of confidence, because:

- the Environmental Values assessment was based on recent survey data supported by database and literature searches, and
- the potential impacts of the Project are generally well understood by specialist aquatic ecologists.

Mitigation Measures

Section 6.2 specifies numerous mitigations that can be applied during one or more phases of the Project to reduce the level of risk of the identified sources of potential impact on aquatic ecological values.

Assessment Against the Significant Impact Criteria

The assessment against the Significant Impact Criteria for critically endangered and endangered aquatic species is presented in Table 6.4. The assessment against the Significant Impact Criteria for vulnerable aquatic species is presented in Table 6.5.

The assessment indicates a significant impact to aquatic MNES species associated with the Project is unlikely.

Significant Impact Criteria	Will the action have a significant impact	Justification
Lead to a long-term decrease in the size of a population of a species	No	Mary River turtle and white-throated snapping turtle are not expected to occur in Six Mile Creek; thus it is unlikely that any population of these species will be in or near the Project area. However, the application of appropriate mitigations will protect water quality, habitat and flows that support these species, as well as ensure any individuals of these species are not injured or stranded should any be present in Lake Macdonald. There will be no long-term decrease in the size of a population of Mary River turtle or white-throated snapping turtle associated with the Project.
		Mary River cod is known from Six Mile Creek downstream of Lake Macdonald, and has been stocked to Lake Macdonald. The application of appropriate mitigations will protect water quality, habitat and flows that support this species, as well as ensure any individuals of Mary River cod are not injured or stranded in Lake Macdonald. Stocking densities at the relocation sites will be carefully managed and monitored. There will be no long-term decrease in the size of a population of Mary River cod associated with the Project.

 Table 6.4
 Assessment of the Project on critically endangered and endangered aquatic MNES species: Mary River cod, Mary River turtle, white-throated snapping turtle.

Significant Impact Criteria	Will the action have a significant impact	Justification
Reduce the area of occupancy of the species	No	The area of occupancy of Mary River turtle and white-throated snapping turtle is unlikely to be influenced by the Project, because these species are not expected to occur in Six Mile Creek. The area of Lake Macdonald will be temporarily reduced during the drawdown and construction phases of the Project, with a fauna salvage and relocation operation implemented during these phases of the Project. Should any Mary River turtle or white-throated snapping turtle be caught and relocated during the salvage operation, they will be returned to Lake Macdonald during the refill and operate phase, ensuring that the area of occupancy of these species after completion of construction is the same as current area of occupancy.
		The area of Lake Macdonald will be temporarily reduced during the drawdown and construction phases of the Project, with a fauna salvage and relocation operation implemented during these phases of the Project. It is expected that Mary River cod will be the primary focus of the salvage operation, with any cod caught during the salvage operation returned to Lake Macdonald during the refill and operate phase, ensuring that the area of occupancy of this species after completion of construction is the same as current area of occupancy.
Fragment an existing population into two or more populations	No change from current	Mary River turtle and white-throated snapping turtle are not expected to occur in Six Mile Creek; thus it is unlikely that any population of these species will be in or near the Project area.
		Mary River cod is known from Six Mile Creek downstream of Lake Macdonald, and has been stocked to Lake Macdonald. Replacement of the dam wall will not change the current level of connectivity (i.e. no connectivity) between upstream and downstream populations of Mary River cod.

Significant Impact Criteria	Will the action have a significant impact	Justification
Adversely affect habitat critical to the survival of a species	No	Mary River turtle and white-throated snapping turtle are not expected to occur in Six Mile Creek; and habitat critical for the survival of these species does not occur in Lake Macdonald or Six Mile Creek. The Project will have no impact on habitat critical to the survival of these species.
		Habitat critical to the survival of Mary River cod occurs in Six Mile Creek downstream of Lake Macdonald. While Mary River cod has been stocked to Lake Macdonald, it is considered that the lake is not habitat critical to the survival of this species. Impacts to downstream habitat and water quality in Six Mile Creek will be mitigated using numerous measures such as those described in Section 6.2. It is unlikely that there will be an impact to habitat critical to the survival of Mary River cod.
Disrupt the breeding cycle of a population	No	Mary River turtle and white-throated snapping turtle are not expected to occur in Six Mile Creek; and breeding habitat for these species in not known from Six Mile Creek or Lake Macdonald. The Project will not adversely impact breeding by Mary River turtle or white-throated snapping turtle.
		Mary River cod are known to breed in Six Mile Creek downstream of Lake Macdonald, but the population in Lake Macdonald is considered to be non-breeding. Impacts to downstream habitat and water quality in Six Mile Creek will be mitigated using numerous measures such as those described in Section 6.2, which will also function to minimise potential impacts to Mary River cod breeding in Six Mile Creek. It is proposed that the drawdown phase will occur outside the breeding season of Mary River cod; thus, this phase of the Project will have no influence on Mary River cod breeding in Six Mile Creek.

Significant Impact Criteria	Will the action have a significant impact	Justification
Modify, destroy, remove, isolate or decrease the availability or quality of habitat to the extent that the species is likely to decline	No	Mary River turtle and white-throated snapping turtle are not expected to occur in Six Mile Creek; and suitable habitat for these species in not known from Six Mile Creek or Lake Macdonald. The Project will not modify, destroy, remove, isolate or decrease the availability or quality of habitat of this species.
		Mary River cod are known to breed in Six Mile Creek downstream of Lake Macdonald, but the population in Lake Macdonald is considered to be non-breeding. Impacts to downstream habitat and water quality in Six Mile Creek will be mitigated using numerous measures described in Section 6.2, which will also function to minimise potential impacts to Mary River cod breeding in Six Mile Creek. Temporary reduction of habitat availability for stocked Mary River cod in Lake Macdonald during the drawdown and construction phases will be mitigated by a salvage, relocation and restocking program, with long- term habitat quality for Mary River cod in Lake Macdonald potentially enhanced during the construction phase. Therefore the Project will not modify, destroy, remove, isolate or decrease the availability or quality of habitat to the extent that Mary River cod is likely to decline
Result in invasive species that are harmful to a critically endangered or endangered species becoming established in the endangered or critically endangered species' habitat	No	Known biosecurity matters of the Project area (e.g. Cabomba, Hygrophila, tilapia) will be carefully managed via the Lake Lowering Plan and the Environmental Management Plan.
Introduce disease that may cause the species to decline	No	The Project will not introduce a disease or pathogen to the Project area as all equipment used should be clean before attending the Project area. Appropriate mitigation measures will also be implemented to minimise injury to aquatic fauna that can leave them susceptible to diseases and pathogens.

Significant Impact Criteria	Will the action have a significant impact	Justification
Interfere with the recovery of the species.	No	White-throated snapping turtle
		Key recovery actions for white-throated snapping turtle include: controlling predators and cattle access to nesting sites to prevent trampling and predation of nests and hatchlings to improve recruitment; managing water releases to avoid inundation of nesting banks during the incubation period while providing adequate environmental flows of good quality water to provide base flows, refugial habitat and geomorphological process to sustain nesting banks; and ensuring that dam and spillway designs minimise injury and mortality of turtles over spillways. As populations and nesting sites for white-throated snapping turtle are not known from Six Mile Creek, the Project will not interfere with recovery actions relating to recruitment, waterway barriers and environmental flows. The final design of the labyrinth spillway will have low fall heights, sufficiently deep plunge pools and adopt the recommendations of Berghuis (2017); thus, the design of the new dam will be consistent with the recovery plan for white-throated snapping turtle.
		Mary River turtle
		There is no recovery plan currently available for Mary River turtle, although the conservation advice statement for the species indicates threats to Mary River turtle are: predation and trampling of nests and hatchlings and lack of recruitment; unfavourable water releases from dams, clearing for agriculture, and impacts to habitat from pollution and invasive weeds. Although recovery action have not currently been developed for Mary River turtle, the Project will not contribute any of

Significant Impact Criteria	Will the action have a significant impact	Justification
		the key threats known for the species. Furthermore, as populations and nesting sites for Mary River turtle are not known from Six Mile Creek, the Project will not interfere with the recovery of this species.
		Mary River cod
		Key recovery recommendations for Mary River cod include: community education and regulatory and administrative initiatives; limiting waterway barrier construction while ensuring that fishways are incorporated into the design of new barriers (and existing barriers where possible), prohibition of stocking of non-indigenous fish in the Mary River systems (especially other species of <i>Maccullochella</i>), environmental flow management, and hatchery and stocking programs. The Project will include a commitment to continue support of the Mary Cod hatchery at Lake Macdonald, with water releases to Six Mile Creek complying with the ROP during the refill and operate phase. While a fishway has been determined to be non-viable at the upgraded Six Mile Creek dam, a fishway will be provided at Gympie Weir (which has been identified as a high priority barrier for retrofitting a fishway, Stockwell et al. 2008). The Project therefore is consistent

Significant Impact Criteria	Will the action have a significant impact	Justification
Lead to a long-term decrease in the size of an important population of a species	n No	Australian lungfish are occur in low numbers in Six Mile Creek (Appendix G; Atlas of Living Australia), with breeding by the species in Six Mile Creek downstream of Lake Macdonald likely to be infrequent and not critical to the species' long-term survival, as breeding habitat (i.e. submerged aquatic plant beds) for lungfish is not present in this reach of Six Mile Creek. Few Australian lungfish are expected to occur in Lake Macdonald or upstream of Lake Macdonald, and the species does not breed in Lake Macdonald. Thus, there is not an important population of Australian lungfish in Six Mile Creek or Lake Macdonald. However, the application of appropriate mitigations will protect water quality, habitat and flows that support this species, as well as ensure any individuals of this species are not injured or stranded should any be present in Lake Macdonald. There will be no long-term decrease in the size of an important population of Australian lungfish associated with the Project.
An important population is a population that is necessary for a species' long-term survival, including source population for breeding or dispersal, necessary for maintaining genetic diversity, and / or populations near the limit of a species' range		
Reduce the area of occupancy of the species	No	The area of Lake Macdonald will be temporarily reduced during the drawdown and construction phases of the Project, with a fauna salvage and relocation operation implemented during these phases of the Project. It is expected that very few Australian lungfish occur in Lake Macdonald, and thus the temporary reduction in lake area will not be a significant impact on the species. However, any lungfish in Lake Macdonald will be salvaged during the drawdown and construction phases and returned to Lake Macdonald during the refill and operate phase, ensuring that the area of occupancy of this species after completion of construction is the same as current area of occupancy.

Table 6.5Assessment of the Project on vulnerable aquatic MNES species: Australian lungfish.

Significant Impact Criteria	Will the action have a significant impact	Justification
Fragment an existing population into two or more populations	No change from current	Australian lungfish are known to occur in low numbers in Six Mile Creek, and may also occur in low numbers in Lake Macdonald. Replacement of the dam wall will not change the current level of connectivity (i.e. no connectivity) between upstream and downstream populations of Australian lungfish.
Adversely affect habitat critical to the survival of a species	No	There is no habitat critical to the survival of Australian lungfish in Six Mile Creek or Lake Macdonald. Impacts to downstream habitat and water quality in Six Mile Creek will be mitigated using numerous measures described in Section 6.2. There will be no impact to habitat critical to the survival of Australian lungfish.
Disrupt the breeding cycle of a population	No	Australian lungfish are known to occur in low numbers in Six Mile Creek, with breeding by the species in Six Mile Creek likely to be infrequent. Impacts to downstream habitat and water quality in Six Mile Creek will be mitigated using numerous measures such as those described in Section 6.2, which will also function to minimise potential impacts to Australian lungfish breeding in Six Mile Creek. The drawdown phase is planned to occur outside the breeding season of Australian lungfish; thus, this phase of the Project will have no influence on any Australian lungfish breeding in Six Mile Creek.
Modify, destroy, remove, isolate or decrease the availability or quality of habitat to the extent that the species is likely to decline	No	Australian lungfish are known to occur in low numbers in Six Mile Creek, and may also occur in low numbers in Lake Macdonald. While neither Six Mile Creek or Lake Macdonald is habitat that is critical to the survival of Australian lungfish, the application of appropriate mitigations as described above will protect water quality, habitat and flows that support this species. There will be no adverse effects to the quality of habitat for Australian lungfish associated with the Project.

Significant Impact Criteria	Will the action have a significant impact	Justification
Result in invasive species that are harmful to a critically endangered or endangered species becoming established in the endangered or critically endangered species' habitat	No	Known biosecurity matters of the Project area (e.g. Cabomba, Hygrophila, tilapia) will be carefully managed via the Aquatic Fauna Salvage and Relocation Plan and the Environmental Management Plan.
Introduce disease that may cause the species to decline	No	The Project will not introduce a disease or pathogen to the Project area.
Interfere with the recovery of the species.	No	There is no recovery plan currently available for Australian lungfish, although the conservation advice for the species identified the main threats as: impoundment of riverine habitat and barriers to movement by dams, reductions in water levels during spawning periods, and pest fish (especially tilapia). Although recovery action have not currently been developed for Australian lungfish, important populations of this species do not occur in Six Mile Creek or Lake Macdonald; thus, the Project will not interfere with the recovery of the species.

7 Conclusions

Two threatened fish and two freshwater turtle species listed under the *Environment Protection and Biodiversity Act 1994* (i.e. aquatic Matters of National Environmental Significance) are known from the Mary River and Six Mile Creek:

- white-throated snapping turtle (*Elseya albagula*), critically endangered
- · Mary River cod (Maccullochella mariensis), endangered
- · Mary River turtle (*Elusor macrurus*), endangered, and
- Australian lungfish (*Neoceratodus forsteri*), vulnerable.

Mary River cod and Australian lungfish are known from Six Mile Creek downstream of Lake Macdonald, and it is possible that Mary River turtle and white-throated snapping turtle sometimes occur in the lower reaches of Six Mile Creek. Mary River cod are known to breed in Six Mile Creek downstream of Lake Macdonald. Within Lake Macdonald, and upstream of Lake Macdonald, platypus are known to occur, Mary River cod and Australian lungfish may occur but are unlikely to be breeding, and Mary River turtle or white-throated snapping turtle are likely to be rare or absent. Matters of State Environmental Significance downstream and upstream of Lake Macdonald include: waterways providing for fish passage, several categories of regulated vegetation and High Ecological Value (watercourse) waters.

The desktop assessment indicated that the alluvium of the Project area is *unlikely to be suitable* for stygofauna due to high clay content, low hydraulic conductivity and high total dissolved solids, and the Kin Kin sandstone is *suitable* for stygofauna due to the higher hydraulic conductivity.

The following sources of potential impact from the Project to the aquatic Environmental Values of Six Mile Creek and Lake Macdonald were identified:

- · Impacts to water quality in Lake Macdonald and downstream of the lake
- · Impacts to aquatic habitat in Lake Macdonald and downstream
- · Impacts to aquatic fauna (injury, mortality or stranding) in Lake Macdonald
- · Impacts to aquatic flora in Lake Macdonald and downstream of the lake
- · Spread of biosecurity matters downstream of Lake Macdonald
- · Impacts to stygofauna communities in shallow groundwater systems, and
- · Barriers to fish passage at the dam wall.

The risks of, and mitigations for, each of the identified potential sources of adverse impact of the Project on the Environmental Values of Six Mile Creek were assessed using a riskbased approach.

Potential direct and indirect impacts of the Project are likely to affect both Lake Macdonald and Six Mile Creek downstream. As the Project is replacing an existing dam wall the longterm impact will be no change from current condition. Most sources of impact during the Project were assessed as having a low risk of impact when appropriate mitigations are applied. Most potential impacts will be temporary (i.e. for the duration of the drawdown and construction periods; approximately 2 to 3 years in total), and if appropriate mitigations are applied there will be no ongoing impacts to the aquatic environmental values of Lake Macdonald or Six Mile Creek.

However, the temporary loss of aquatic habitat in Lake Macdonald due to the drawdown of the lake for safety during construction still resulted in a moderate residual impact after mitigation measures were applied. There will be a temporary loss of approximately 97.2% of aquatic habitat (by water volume) in Lake Macdonald, which requires additional mitigation in the form of a comprehensive aquatic fauna salvage operation. This is an unavoidable risk given the safety requirements of the Project.

Matters of National Environmental Significance were also assessed against the Significant Impact Criteria for critically endangered, endangered and vulnerable aquatic species. The assessment indicates that, while there may be temporary impacts, a significant impact from the Project on aquatic species that are Matters of National Environmental Significance is unlikely.

Fish passage is currently not provided at the Six Mile Creek dam. While fishway options have been considered for the Six Mile Creek dam upgrade (e.g. frc environmental, 2016; Seqwater 2018), preliminary advice from DAF indicates that several factors outweigh the benefit of providing fish passage over the upgraded dam, including site constraints, the risk of aiding upstream dispersal by tilapia, and the anticipated regional benefits of the proposed offsite mitigation option (DAF 2018). Off-site mitigation measures for fish passage are therefore proposed; specifically the provision of fish passage at Gympie Weir by installation of a suitable fishway.

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Appendix A Matters of National Environmental Significance Search Results
Australian Government



Department of the Environment and Energy

EPBC Act Protected Matters Report

This report provides general guidance on matters of national environmental significance and other matters protected by the EPBC Act in the area you have selected.

Information on the coverage of this report and qualifications on data supporting this report are contained in the caveat at the end of the report.

Information is available about <u>Environment Assessments</u> and the EPBC Act including significance guidelines, forms and application process details.

Report created: 27/08/18 11:40:10

Summary Details Matters of NES Other Matters Protected by the EPBC Act Extra Information Caveat Acknowledgements



This map may contain data which are ©Commonwealth of Australia (Geoscience Australia), ©PSMA 2010

Coordinates Buffer: 10.0Km



Summary

Matters of National Environmental Significance

This part of the report summarises the matters of national environmental significance that may occur in, or may relate to, the area you nominated. Further information is available in the detail part of the report, which can be accessed by scrolling or following the links below. If you are proposing to undertake an activity that may have a significant impact on one or more matters of national environmental significance then you should consider the <u>Administrative Guidelines on Significance</u>.

World Heritage Properties:	None
National Heritage Places:	None
Wetlands of International Importance:	1
Great Barrier Reef Marine Park:	None
Commonwealth Marine Area:	None
Listed Threatened Ecological Communities:	3
Listed Threatened Species:	72
Listed Migratory Species:	44

Other Matters Protected by the EPBC Act

This part of the report summarises other matters protected under the Act that may relate to the area you nominated. Approval may be required for a proposed activity that significantly affects the environment on Commonwealth land, when the action is outside the Commonwealth land, or the environment anywhere when the action is taken on Commonwealth land. Approval may also be required for the Commonwealth or Commonwealth agencies proposing to take an action that is likely to have a significant impact on the environment anywhere.

The EPBC Act protects the environment on Commonwealth land, the environment from the actions taken on Commonwealth land, and the environment from actions taken by Commonwealth agencies. As heritage values of a place are part of the 'environment', these aspects of the EPBC Act protect the Commonwealth Heritage values of a Commonwealth Heritage place. Information on the new heritage laws can be found at http://www.environment.gov.au/heritage

A <u>permit</u> may be required for activities in or on a Commonwealth area that may affect a member of a listed threatened species or ecological community, a member of a listed migratory species, whales and other cetaceans, or a member of a listed marine species.

Commonwealth Land:	None
Commonwealth Heritage Places:	1
Listed Marine Species:	46
Whales and Other Cetaceans:	2
Critical Habitats:	None
Commonwealth Reserves Terrestrial:	None
Australian Marine Parks:	None

Extra Information

This part of the report provides information that may also be relevant to the area you have nominated.

State and Territory Reserves:	17
Regional Forest Agreements:	None
Invasive Species:	38
Nationally Important Wetlands:	1
Key Ecological Features (Marine)	None

Details

Matters of National Environmental Significance

Wetlands of International Importance (Ramsar)	[Resource Information]
Name	Proximity
Great sandy strait (including great sandy strait, tin can bay and tin can	30 - 40km upstream

Listed Threatened Ecological Communities

For threatened ecological communities where the distribution is well known, maps are derived from recovery plans, State vegetation maps, remote sensing imagery and other sources. Where threatened ecological community distributions are less well known, existing vegetation maps and point location data are used to produce indicative distribution maps.

Name	Status	Type of Presence
Coastal Swamp Oak (Casuarina glauca) Forest of New South Wales and South East Queensland ecological	Endangered	Community likely to occur within area
Lowland Rainforest of Subtropical Australia	Critically Endangered	Community likely to occur within area
Subtropical and Temperate Coastal Saltmarsh	Vulnerable	Community likely to occur within area
Listed Threatened Species		[Resource Information]
Name	Status	Type of Presence
Birds		
Anthochaera phrygia Regent Honeyeater [82338]	Critically Endangered	Species or species habitat known to occur within area
Botaurus poiciloptilus Australasian Bittern [1001]	Endangered	Species or species habitat known to occur within area
<u>Calidris canutus</u> Red Knot, Knot [855]	Endangered	Species or species habitat may occur within area
<u>Calidris ferruginea</u> Curlew Sandpiper [856]	Critically Endangered	Species or species habitat likely to occur within area

Cyclopsitta diophthalma coxeni

Coxen's Fig-Parrot [59714]

Dasyornis brachypterus Eastern Bristlebird [533]

Diomedea antipodensis Antipodean Albatross [64458]

Diomedea antipodensis gibsoni Gibson's Albatross [82270]

Diomedea exulans Wandering Albatross [89223] Endangered

Species or species habitat likely to occur within area

[Resource Information]

Endangered

Species or species habitat may occur within area

Species or species habitat

may occur within area

Vulnerable

Vulnerable

Species or species habitat may occur within area

Vulnerable

Species or species habitat may occur within area

Name	Status	Type of Presence
Erythrotriorchis radiatus Red Goshawk [942]	Vulnerable	Species or species habitat known to occur within area
<u>Geophaps scripta</u> Squatter Pigeon (southern) [64440]	Vulnerable	Species or species habitat may occur within area
Lathamus discolor Swift Parrot [744]	Critically Endangered	Species or species habitat likely to occur within area
<u>Limosa lapponica baueri</u> Bar-tailed Godwit (baueri), Western Alaskan Bar-tailed Godwit [86380]	Vulnerable	Species or species habitat likely to occur within area
Limosa lapponica menzbieri Northern Siberian Bar-tailed Godwit, Bar-tailed Godwit (menzbieri) [86432]	Critically Endangered	Species or species habitat may occur within area
Macronectes giganteus Southern Giant-Petrel, Southern Giant Petrel [1060]	Endangered	Species or species habitat may occur within area
Macronectes halli Northern Giant Petrel [1061]	Vulnerable	Species or species habitat may occur within area
Numenius madagascariensis Eastern Curlew, Far Eastern Curlew [847]	Critically Endangered	Species or species habitat known to occur within area
Pachyptila turtur subantarctica Fairy Prion (southern) [64445]	Vulnerable	Species or species habitat likely to occur within area
Poephila cincta cincta Southern Black-throated Finch [64447]	Endangered	Species or species habitat may occur within area
Rostratula australis Australian Painted-snipe, Australian Painted Snipe [77037]	Endangered	Species or species habitat likely to occur within area
Thalassarche cauta cauta Shy Albatross, Tasmanian Shy Albatross [82345]	Vulnerable	Species or species habitat may occur within area
Thalassarche cauta steadi White-capped Albatross [82344]	Vulnerable	Species or species habitat likely to occur within area
Thalassarche eremita Chatham Albatross [64457]	Endangered	Species or species habitat may occur within area
<u>Thalassarche impavida</u> Campbell Albatross, Campbell Black-browed Albatross [64459]	Vulnerable	Species or species habitat may occur within area
Thalassarche melanophris Black-browed Albatross [66472]	Vulnerable	Species or species habitat may occur within area
<u>Thalassarche salvini</u> Salvin's Albatross [64463]	Vulnerable	Species or species habitat may occur within area
Turnix melanogaster Black-breasted Button-quail [923]	Vulnerable	Species or species habitat

Fish

Name	Status	Type of Presence
Epinephelus daemelii		
Black Rockcod, Black Cod, Saddled Rockcod [68449]	Vulnerable	Species or species habitat may occur within area
Maccullochella mariensis		
Mary River Cod [83806]	Endangered	Species or species habitat known to occur within area
Neoceratodus forsteri		
Australian Lungfish, Queensland Lungfish [67620]	Vulnerable	Species or species habitat known to occur within area
Frogs		
Litoria olongburensis		
Wallum Sedge Frog [1821]	Vulnerable	Species or species habitat known to occur within area
Mixophyes fleavi		
Fleay's Frog [25960]	Endangered	Species or species habitat likely to occur within area
Mixophyes iteratus		
Giant Barred Frog, Southern Barred Frog [1944]	Endangered	Species or species habitat known to occur within area
Insects		
Argynnis hyperbius inconstans		
Australian Fritillary [88056]	Critically Endangered	Species or species habitat may occur within area
Phyllodes imperialis smithersi		
Pink Underwing Moth [86084]	Endangered	Breeding may occur within area
Mammals		
Chalinolobus dwyeri		
Large-eared Pied Bat, Large Pied Bat [183]	Vulnerable	Species or species habitat likely to occur within area
Dasyurus hallucatus		
Northern Quoll, Digul [Gogo-Yimidir], Wijingadda [Dambimangari], Wiminji [Martu] [331]	Endangered	Species or species habitat likely to occur within area
Dasvurus maculatus maculatus (SE mainland populatio	on)	
Spot-tailed Quoll, Spotted-tail Quoll, Tiger Quoll (southeastern mainland population) [75184]	Endangered	Species or species habitat likely to occur within area

Petauroides volans		
Greater Glider [254]	Vulnerable	Species or species habitat likely to occur within area
Phascolarctos cinereus (combined populations of Qld, I	<u>NSW and the ACT)</u>	
Koala (combined populations of Queensland, New South Wales and the Australian Capital Territory) [85104]	Vulnerable	Species or species habitat known to occur within area
Potorous tridactylus tridactylus		
Long-nosed Potoroo (SE mainland) [66645]	Vulnerable	Species or species habitat likely to occur within area
Pteropus poliocephalus		
Grey-headed Flying-fox [186]	Vulnerable	Roosting known to occur within area
Xeromys myoides		
Water Mouse, False Water Rat, Yirrkoo [66]	Vulnerable	Species or species habitat known to occur within area
Plants		
Acacia attenuata		
[10690]	Vulnerable	Species or species habitat likely to occur within area
Allocasuarina thalassoscopica		
[21927]	Endangered	Species or species habitat may occur within

Name	Status	Type of Presence
Archidendron lovellize		area
Bacon Wood, Tulip Siris [13451]	Vulnerable	Species or species habitat likely to occur within area
<u>Arthraxon hispidus</u> Hairy-joint Grass [9338]	Vulnerable	Species or species habitat likely to occur within area
Baloghia marmorata Marbled Balogia, Jointed Baloghia [8463]	Vulnerable	Species or species habitat likely to occur within area
Bosistoa transversa Three-leaved Bosistoa, Yellow Satinheart [16091]	Vulnerable	Species or species habitat likely to occur within area
Cryptocarya foetida Stinking Cryptocarya, Stinking Laurel [11976]	Vulnerable	Species or species habitat likely to occur within area
<u>Cryptostylis hunteriana</u> Leafless Tongue-orchid [19533]	Vulnerable	Species or species habitat may occur within area
Eucalyptus conglomerata Swamp Stringybark [3160]	Endangered	Species or species habitat likely to occur within area
Floydia praealta Ball Nut, Possum Nut, Big Nut, Beefwood [15762]	Vulnerable	Species or species habitat likely to occur within area
Lepidium peregrinum Wandering Pepper-cress [14035]	Endangered	Species or species habitat may occur within area
Macadamia integrifolia Macadamia Nut, Queensland Nut Tree, Smooth- shelled Macadamia, Bush Nut, Nut Oak [7326]	Vulnerable	Species or species habitat known to occur within area
Macadamia ternifolia Small-fruited Queensland Nut, Gympie Nut [7214]	Vulnerable	Species or species habitat known to occur within area
Phaius australis Lesser Swamp-orchid [5872]	Endangered	Species or species habitat likely to occur within area
Prostanthera spathulata [88266]	Vulnerable	Species or species habitat known to occur within area
<u>Samadera bidwillii</u> Quassia [29708]	Vulnerable	Species or species habitat likely to occur within area
<u>Sophora fraseri</u> [8836]	Vulnerable	Species or species habitat may occur within area
<u>Triunia robusta</u> Glossy Spice Bush [14747]	Endangered	Species or species habitat known to occur within area
Xanthostemon oppositifolius Penda, Southern Penda, Luya's Hardwood [8738]	Vulnerable	Species or species habitat likely to occur within area
Reptiles		
Caretta caretta		
Loggerhead Turtle [1763]	Endangered	Foraging, feeding or related behaviour known

Name	Status	Type of Presence
		to occur within area
<u>Chelonia mydas</u> Green Turtle [1765]	Vulnerable	Foraging, feeding or related behaviour known to occur within area
Delma torquata Adorned Delma, Collared Delma [1656]	Vulnerable	Species or species habitat
		may occur within area
Dermochelys coriacea		
Leatherback Turtle, Leathery Turtle, Luth [1768]	Endangered	Species or species habitat known to occur within area
Eretmochelys imbricata		
Hawksbill Turtle [1766]	Vulnerable	Foraging, feeding or related behaviour known to occur within area
Furina dunmalli		• • • • • • • • •
Dunmall's Snake [59254]	Vulnerable	Species or species habitat may occur within area
Lepidochelys olivacea		
Olive Ridley Turtle, Pacific Ridley Turtle [1767]	Endangered	Foraging, feeding or related behaviour likely to occur within area
Natator depressus		
Flatback Turtle [59257]	Vulnerable	Foraging, feeding or related behaviour known to occur within area
Saiphos reticulatus		
Three-toed Snake-tooth Skink [88328]	Vulnerable	Species or species habitat may occur within area
Sharks		
Pristis zijsron		Dreading may acquir within
[68442]	Vuinerable	area
Listed Migratory Species		[Resource Information]
* Species is listed under a different scientific name on the	he EPBC Act - Threatened	Species list.
Name Migratory Marine Birde	Threatened	Type of Presence
Anous stolidus		
Common Noddy [825]		Species or species habitat may occur within area

Apus pacificus Fork-tailed Swift [678]

Calonectris leucomelas Streaked Shearwater [1077]

Diomedea antipodensis Antipodean Albatross [64458]

Diomedea exulans Wandering Albatross [89223]

<u>Fregata ariel</u> Lesser Frigatebird, Least Frigatebird [1012]

<u>Fregata minor</u> Great Frigatebird, Greater Frigatebird [1013]

<u>Macronectes giganteus</u> Southern Giant-Petrel, Southern Giant Petrel

Endangered

Species or species

Species or species habitat likely to occur within area

Species or species habitat may occur within area

Species or species habitat may occur within area

Species or species habitat may occur within area

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Vulnerable

Vulnerable

Name	Threatened	Type of Presence
[1060]		habitat may occur within area
Macronectes halli		
Northern Giant Petrel [1061]	Vulnerable	Species or species habitat may occur within area
Thalassarche cauta		
Tasmanian Shy Albatross [89224]	Vulnerable*	Species or species habitat may occur within area
Thalassarche eremita		
Chatham Albatross [64457]	Endangered	Species or species habitat may occur within area
Thalassarche impavida		
Campbell Albatross, Campbell Black-browed Albatross [64459]	Vulnerable	Species or species habitat may occur within area
Thalassarche melanophris		
Black-browed Albatross [66472]	Vulnerable	Species or species habitat may occur within area
Thalassarche salvini		
Salvin's Albatross [64463]	Vulnerable	Species or species habitat may occur within area
Thalassarche steadi		
White-capped Albatross [64462]	Vulnerable*	Species or species habitat likely to occur within area
Migratory Marine Species		
Caretta caretta		
Loggerhead Turtle [1763]	Endangered	Foraging, feeding or related behaviour known to occur within area
<u>Chelonia mydas</u>		
Green Turtle [1765]	Vulnerable	Foraging, feeding or related behaviour known to occur within area
Crocodylus porosus		
Salt-water Crocodile, Estuarine Crocodile [1774]		Species or species habitat likely to occur within area
Dermochelys coriacea		
Leatherback Turtle, Leathery Turtle, Luth [1768]	Endangered	Species or species habitat

Eretmochelys imbricata Hawksbill Turtle [1766]

Lamna nasus Porbeagle, Mackerel Shark [83288]

Lepidochelys olivacea Olive Ridley Turtle, Pacific Ridley Turtle [1767]

Manta alfredi

Reef Manta Ray, Coastal Manta Ray, Inshore Manta Ray, Prince Alfred's Ray, Resident Manta Ray [84994]

Manta birostris

Giant Manta Ray, Chevron Manta Ray, Pacific Manta Ray, Pelagic Manta Ray, Oceanic Manta Ray [84995]

Natator depressus Flatback Turtle [59257]

Orcaella brevirostris Irrawaddy Dolphin [45]

Vulnerable

Endangered

Foraging, feeding or related behaviour known to occur within area

Species or species habitat may occur within area

Foraging, feeding or related behaviour likely to occur within area

Species or species habitat may occur within area

Species or species habitat may occur within area

Vulnerable

Foraging, feeding or related behaviour known to occur within area

Species or species

Name	Threatened	Type of Presence
		habitat likely to occur within area
Pristis zijsron		
Green Sawfish, Dindagubba, Narrowsnout Sawfish [68442] Sousa chinensis	Vulnerable	Breeding may occur within area
Indo-Pacific Humpback Dolphin [50]		Species or species habitat likely to occur within area
Migratory Terrestrial Species		
Cuculus optatus		
Oriental Cuckoo, Horsfield's Cuckoo [86651]		Species or species habitat known to occur within area
Hirundapus caudacutus		
White-throated Needletail [682]		Species or species habitat known to occur within area
Monarcha melanopsis		
Black-faced Monarch [609]		Species or species habitat known to occur within area
Monarcha trivirgatus		
Spectacled Monarch [610]		Species or species habitat known to occur within area
Mviagra cyanoleuca		
Satin Flycatcher [612]		Species or species habitat known to occur within area
Dhinidura rufifrana		
Rufous Fantail [592]		Species or species habitat known to occur within area
Migratory Wetlands Species		
Actitis hypoleucos		
Common Sandpiper [59309]		Species or species habitat known to occur within area
Calidris acuminata		
Sharp-tailed Sandpiper [874]		Species or species habitat known to occur within area

Calidris canutus Red Knot, Knot [855]

Endangered

Species or species habitat may occur within area

Calidris ferruginea Curlew Sandpiper [856]

<u>Calidris melanotos</u> Pectoral Sandpiper [858]

Gallinago hardwickii Latham's Snipe, Japanese Snipe [863]

Limosa lapponica Bar-tailed Godwit [844]

Numenius madagascariensis Eastern Curlew, Far Eastern Curlew [847]

Pandion haliaetus Osprey [952]

Tringa nebularia Common Greenshank, Greenshank [832] Critically Endangered Species or likely to oc

Species or species habitat likely to occur within area

Species or species habitat may occur within area

Species or species habitat may occur within area

Species or species habitat known to occur within area

Critically Endangered

Species or species habitat known to occur within area

Breeding known to occur within area

Species or species habitat likely to occur

Name	Threatened	Type of Presence
		within area

Other Matters Protected by the EPBC Act

Commonwealth Heritage Places		[Resource Information]
Name	State	Status
Historic		
Cooroy Post Office	QLD	Listed place
Listed Marine Species		[Resource Information]
* Species is listed under a different scientific name on th	e EPBC Act - Threatened	l Species list.
Name	Threatened	Type of Presence
Birds		
Actitis hypoleucos		
Common Sandpiper [59309]		Species or species habitat known to occur within area
Anous stolidus		
Common Noddy [825]		Species or species habitat may occur within area
Anseranas semipalmata Magpie Goose [978]		Species or species habitat may occur within area

Apus pacificus



Ardea alba Great Egret, White Egret [59541]

Ardea ibis Cattle Egret [59542]

Calidris acuminata Sharp-tailed Sandpiper [874]

Calidris canutus Red Knot, Knot [855]

Calidris ferruginea Curlew Sandpiper [856]

<u>Calidris melanotos</u> Pectoral Sandpiper [858] Species or species habitat likely to occur within area

Species or species habitat known to occur within area

Species or species habitat may occur within area

Species or species habitat known to occur within area

Endangered

Species or species habitat may occur within area

Critically Endangered

Species or species habitat likely to occur within area

Species or species

Name	Threatened	Type of Presence
		habitat may occur within area
Calonectris leucomelas		
Streaked Shearwater [1077]		Species or species habitat may occur within area
Diomedea antipodensis		
Antipodean Albatross [64458]	Vulnerable	Species or species habitat may occur within area
Diomedea exulans		
Wandering Albatross [89223]	Vulnerable	Species or species habitat may occur within area
Diomedea gibsoni		
Gibson's Albatross [64466]	Vulnerable*	Species or species habitat may occur within area
Fregata ariel		
Lesser Frigatebird, Least Frigatebird [1012]		Species or species habitat likely to occur within area
Fregata minor		
Great Frigatebird, Greater Frigatebird [1013]		Species or species habitat likely to occur within area
Gallinago hardwickii		
Latham's Snipe, Japanese Snipe [863]		Species or species habitat may occur within area
Haliaeetus leucogaster		
White-bellied Sea-Eagle [943]		Species or species habitat known to occur within area
Hirundapus caudacutus		
White-throated Needletail [682]		Species or species habitat known to occur within area
Lathamus discolor		
Swift Parrot [744]	Critically Endangered	Species or species habitat likely to occur within area
Limosa lapponica		
Bar-tailed Godwit [844]		Species or species habitat

<u>Macronectes giganteus</u> Southern Giant-Petrel, Southern Giant Petrel [1060]

Macronectes halli Northern Giant Petrel [1061]

Merops ornatus Rainbow Bee-eater [670]

Monarcha melanopsis Black-faced Monarch [609]

Monarcha trivirgatus Spectacled Monarch [610]

Myiagra cyanoleuca Satin Flycatcher [612]

Numenius madagascariensis Eastern Curlew, Far Eastern Curlew [847] Endangered

Species or species habitat may occur within area

known to occur within area

Vulnerable

Species or species habitat may occur within area

Species or species habitat may occur within area

Species or species habitat known to occur within area

Species or species habitat known to occur within area

Species or species habitat known to occur within area

Critically Endangered

Species or species habitat known to occur

Name	Threatened	Type of Presence
Pachyptila turtur		within area
Fairy Prion [1066]		Species or species habitat likely to occur within area
Pandion haliaetus		
Osprey [952]		Breeding known to occur within area
Rhipidura rufifrons		
Rufous Fantail [592]		Species or species habitat known to occur within area
Rostratula benghalensis (sensu lato)		
Painted Snipe [889]	Endangered*	Species or species habitat likely to occur within area
Thalassarche cauta		
Tasmanian Shy Albatross [89224]	Vulnerable*	Species or species habitat may occur within area
Thalassarche eremita		
Chatham Albatross [64457]	Endangered	Species or species habitat may occur within area
Thalassarche impavida		
Campbell Albatross, Campbell Black-browed Albatross [64459]	Vulnerable	Species or species habitat may occur within area
Thalassarche melanophris		
Black-browed Albatross [66472]	Vulnerable	Species or species habitat may occur within area
Thalassarche salvini		
Salvin's Albatross [64463]	Vulnerable	Species or species habitat may occur within area
Thalassarche steadi		
White-capped Albatross [64462]	Vulnerable*	Species or species habitat likely to occur within area
Tringa nebularia		
Common Greenshank, Greenshank [832]		Species or species habitat likely to occur within area

Reptiles

Caretta caretta		
Loggerhead Turtle [1763]	Endangered	Foraging, feeding or related behaviour known to occur within area
Chelonia mydas		
Green Turtle [1765]	Vulnerable	Foraging, feeding or related behaviour known to occur within area
Crocodylus porosus		
Salt-water Crocodile, Estuarine Crocodile [1774]		Species or species habitat likely to occur within area
Dermochelys coriacea		
Leatherback Turtle, Leathery Turtle, Luth [1768]	Endangered	Species or species habitat known to occur within area
Eretmochelys imbricata		
Hawksbill Turtle [1766]	Vulnerable	Foraging, feeding or related behaviour known to occur within area
Lepidochelys olivacea		
Olive Ridley Turtle, Pacific Ridley Turtle [1767]	Endangered	Foraging, feeding or related behaviour likely to occur within area
Natator depressus		
Flatback Turtle [59257]	Vulnerable	Foraging, feeding or related behaviour known to occur within area

Whales and other Cetaceans		[Resource Information]
Name	Status	Type of Presence
Mammals		
Orcaella brevirostris		
Irrawaddy Dolphin [45]		Species or species habitat likely to occur within area
Sousa chinensis		
Indo-Pacific Humpback Dolphin [50]		Species or species habitat likely to occur within area

Extra Information

State and Territory Reserves	[Resource Information]
Name	State
Alyxia	QLD
Cooloothin	QLD
Cooroibah Environmental Reserve	QLD
Great Sandy	QLD
Great Sandy National Park	QLD
Harry Spring	QLD
Johns Property addition to Great Sandy National Park	QLD
Kingsgate Drive	QLD
Mount Cooroy	QLD
Penda Scrub	QLD
Six Mile Creek	QLD
Symplocos	QLD
Tewantin	QLD
Tuchekoi	QLD
Una Corbould	QLD
Woondum	QLD
Yurol	QLD

Invasive Species	[Resource Information]
Weeds reported here are the 20 species of national significance (WoNS), along with	th other introduced plants
that are considered by the States and Territories to pose a particularly significant th	nreat to biodiversity. The

following feral animals are reported: Goat, Red Fox, Cat, Rabbit, Pig, Water Buffalo and Cane Toad. Maps from Landscape Health Project, National Land and Water Resouces Audit, 2001.

Name	Status	Type of Presence
Birds		
Acridotheres tristis		
Common Myna, Indian Myna [387]		Species or species habitat likely to occur within area
Anas platyrhynchos		
Mallard [974]		Species or species habitat likely to occur within area
Columba livia		
Rock Pigeon, Rock Dove, Domestic Pigeon [803]		Species or species habitat likely to occur within area
Lonchura punctulata		
Nutmeg Mannikin [399]		Species or species habitat likely to occur within area
Passer domesticus		
House Sparrow [405]		Species or species habitat likely to occur within area

Name	Status	Type of Presence
Streptopelia chinensis Spotted Turtle-Dove [780]		Species or species habitat likely to occur within area
Sturnus vulgaris Common Starling [389]		Species or species habitat likely to occur within area
Frogs		
Rhinella marina Cane Toad [83218]		Species or species habitat known to occur within area
Mammals		
Bos taurus Domestic Cattle [16]		Species or species habitat likely to occur within area
Canis lupus familiaris Domestic Dog [82654]		Species or species habitat likely to occur within area
Felis catus Cat, House Cat, Domestic Cat [19]		Species or species habitat likely to occur within area
Feral deer Feral deer species in Australia [85733]		Species or species habitat likely to occur within area
Lepus capensis Brown Hare [127]		Species or species habitat likely to occur within area
Mus musculus House Mouse [120]		Species or species habitat likely to occur within area
Oryctolagus cuniculus Rabbit, European Rabbit [128]		Species or species habitat likely to occur within area

Rattus rattus Black Rat, Ship Rat [84]

Species or species habitat likely to occur within area

Sus scrofa Pig [6]

Vulpes vulpes Red Fox, Fox [18]

Plants

Annona glabra Pond Apple, Pond-apple Tree, Alligator Apple, Bullock's Heart, Cherimoya, Monkey Apple, Bobwood, Corkwood [6311] Anredera cordifolia Madeira Vine, Jalap, Lamb's-tail, Mignonette Vine, Anredera, Gulf Madeiravine, Heartleaf Madeiravine, Potato Vine [2643] Asparagus aethiopicus Asparagus Fern, Ground Asparagus, Basket Fern, Sprengi's Fern, Bushy Asparagus, Emerald Asparagus [62425] Asparagus africanus Climbing Asparagus, Climbing Asparagus Fern [66907]

Asparagus plumosus Climbing Asparagus-fern [48993] Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species

Name	Status	Type of Presence
		habitat likely to occur within area
Cabomba caroliniana		
Cabomba, Fanwort, Carolina Watershield, Fish Grass, Washington Grass, Watershield, Carolina Fanwort, Common Cabomba [5171] Chrysanthemoides monilifera		Species or species habitat likely to occur within area
Bitou Bush, Boneseed [18983]		Species or species habitat may occur within area
Chrysanthemoides monilifera subsp. rotundata		
Bitou Bush [16332]		Species or species habitat likely to occur within area
Dolichandra unquis-cati		
Cat's Claw Vine, Yellow Trumpet Vine, Cat's Claw Creeper, Funnel Creeper [85119]		Species or species habitat likely to occur within area
Eichhornia crassipes		
Water Hyacinth, Water Orchid, Nile Lily [13466]		Species or species habitat likely to occur within area
Hymenachne amplexicaulis		
Hymenachne, Olive Hymenachne, Water Stargrass, West Indian Grass, West Indian Marsh Grass [31754]		Species or species habitat likely to occur within area
Lantana camara		
Lantana, Common Lantana, Kamara Lantana, Large- leaf Lantana, Pink Flowered Lantana, Red Flowered Lantana, Red-Flowered Sage, White Sage, Wild Sage [10892]		Species or species habitat likely to occur within area
Prickly Pears [82753]		Species or species habitat likely to occur within area
Parthenium hysterophorus		
Parthenium Weed, Bitter Weed, Carrot Grass, False Ragweed [19566]		Species or species habitat likely to occur within area
Sagittaria platyphylla Delta Arrowhead, Arrowhead, Slender Arrowhead [68483]		Species or species habitat likely to occur within area
Salix spp. except S.babylonica, S.x calodendron & S.x	reichardtii	

Willows except Weeping Willow, Pussy Willow and Sterile Pussy Willow [68497]

Species or species habitat likely to occur within area

Salvinia molesta Salvinia, Giant Salvinia, Aquarium Watermoss, Kariba Weed [13665]

Senecio madagascariensis Fireweed, Madagascar Ragwort, Madagascar Groundsel [2624]

Reptiles

Hemidactylus frenatus Asian House Gecko [1708]

Ramphotyphlops braminus Flowerpot Blind Snake, Brahminy Blind Snake, Cacing Besi [1258]

Nationally Important Wetlands	[Resource Information]
Name	State
Noosa River Wetlands	QLD

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species habitat likely to occur within area

Species or species habitat may occur within area

Caveat

The information presented in this report has been provided by a range of data sources as acknowledged at the end of the report.

This report is designed to assist in identifying the locations of places which may be relevant in determining obligations under the Environment Protection and Biodiversity Conservation Act 1999. It holds mapped locations of World and National Heritage properties, Wetlands of International and National Importance, Commonwealth and State/Territory reserves, listed threatened, migratory and marine species and listed threatened ecological communities. Mapping of Commonwealth land is not complete at this stage. Maps have been collated from a range of sources at various resolutions.

Not all species listed under the EPBC Act have been mapped (see below) and therefore a report is a general guide only. Where available data supports mapping, the type of presence that can be determined from the data is indicated in general terms. People using this information in making a referral may need to consider the qualifications below and may need to seek and consider other information sources.

For threatened ecological communities where the distribution is well known, maps are derived from recovery plans, State vegetation maps, remote sensing imagery and other sources. Where threatened ecological community distributions are less well known, existing vegetation maps and point location data are used to produce indicative distribution maps.

Threatened, migratory and marine species distributions have been derived through a variety of methods. Where distributions are well known and if time permits, maps are derived using either thematic spatial data (i.e. vegetation, soils, geology, elevation, aspect, terrain, etc) together with point locations and described habitat; or environmental modelling (MAXENT or BIOCLIM habitat modelling) using point locations and environmental data layers.

Where very little information is available for species or large number of maps are required in a short time-frame, maps are derived either from 0.04 or 0.02 decimal degree cells; by an automated process using polygon capture techniques (static two kilometre grid cells, alpha-hull and convex hull); or captured manually or by using topographic features (national park boundaries, islands, etc). In the early stages of the distribution mapping process (1999-early 2000s) distributions were defined by degree blocks, 100K or 250K map sheets to rapidly create distribution maps. More reliable distribution mapping methods are used to update these distributions as time permits.

Only selected species covered by the following provisions of the EPBC Act have been mapped:

- migratory and
- marine

The following species and ecological communities have not been mapped and do not appear in reports produced from this database:

- threatened species listed as extinct or considered as vagrants
- some species and ecological communities that have only recently been listed
- some terrestrial species that overfly the Commonwealth marine area
- migratory species that are very widespread, vagrant, or only occur in small numbers

The following groups have been mapped, but may not cover the complete distribution of the species:

- non-threatened seabirds which have only been mapped for recorded breeding sites
- seals which have only been mapped for breeding sites near the Australian continent

Such breeding sites may be important for the protection of the Commonwealth Marine environment.

Coordinates

-26.38065 152.93177

Acknowledgements

This database has been compiled from a range of data sources. The department acknowledges the following custodians who have contributed valuable data and advice:

-Office of Environment and Heritage, New South Wales -Department of Environment and Primary Industries, Victoria -Department of Primary Industries, Parks, Water and Environment, Tasmania -Department of Environment, Water and Natural Resources, South Australia -Department of Land and Resource Management, Northern Territory -Department of Environmental and Heritage Protection, Queensland -Department of Parks and Wildlife, Western Australia -Environment and Planning Directorate, ACT -Birdlife Australia -Australian Bird and Bat Banding Scheme -Australian National Wildlife Collection -Natural history museums of Australia -Museum Victoria -Australian Museum -South Australian Museum -Queensland Museum -Online Zoological Collections of Australian Museums -Queensland Herbarium -National Herbarium of NSW -Royal Botanic Gardens and National Herbarium of Victoria -Tasmanian Herbarium -State Herbarium of South Australia -Northern Territory Herbarium -Western Australian Herbarium -Australian National Herbarium, Canberra -University of New England -Ocean Biogeographic Information System -Australian Government, Department of Defence Forestry Corporation, NSW -Geoscience Australia -CSIRO -Australian Tropical Herbarium, Cairns -eBird Australia -Australian Government – Australian Antarctic Data Centre -Museum and Art Gallery of the Northern Territory -Australian Government National Environmental Science Program

-Australian Government National Environmental Scien

-Australian Institute of Marine Science

-Reef Life Survey Australia

-American Museum of Natural History

-Queen Victoria Museum and Art Gallery, Inveresk, Tasmania

-Tasmanian Museum and Art Gallery, Hobart, Tasmania

-Other groups and individuals

The Department is extremely grateful to the many organisations and individuals who provided expert advice and information on numerous draft distributions.

Please feel free to provide feedback via the Contact Us page.

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Appendix B Detailed Description of Aquatic Matters of National Environmental Significance

1 Critically Endangered and Endangered Species

White-throated snapping turtle (listed as critically endangered under the EPBC Act), the Mary River cod (listed as endangered), and the Mary River turtle (listed as endangered), are the only critically endangered or endangered aquatic species that are known from the vicinity of the proposed development, and that could potentially be impacted.

An action is likely to have a significant impact on a critically endangered or endangered species if there is a real chance or possibility that it will:

- · lead to a long-term decrease in the size of a population
- · reduce the area of occupancy of the species
- · fragment an existing population into two or more populations
- · adversely affect habitat critical to the survival of a species
- · disrupt the breeding cycle of a population
- modify, destroy, remove, isolate or decrease the availability or quality of habitat to the extent that the species is likely to decline
- result in invasive species that are harmful to a critically endangered or endangered species becoming established in the endangered or critically endangered species' habitat
- \cdot introduce disease that may cause the species to decline, or
- · interfere with the recovery of the species.

1.1 White-throated snapping turtle

Status

White-throated snapping turtle (*Elseya albagula*) is listed as critically endangered under the EPBC and endangered under the NC Act.

Population

The population of white-throated snapping turtles is dominated by adults, because predation of eggs and hatchlings, and trampling of nests by cattle, have greatly reduced recruitment and the juvenile proportion of the population (Hamann et al. 2007; Limpus 2008;

Limpus et al. 2011; DotEE 2017). This likely represents a significant reduction in population size, because an unimpacted population of freshwater turtles would be expected to have a high proportion of juveniles (Thompson 1983). While population size is responsive to adult survivorship due to relatively high life-time fecundity of long-lived adults (Heppell et al. 1996), and thus population size would likely increase if recruitment rates increase, population size will decline further if recruitment rates continue to be low (Threatened Species Scientific Committee 2014a).

Population genetic analyses indicate that the population of white-throated snapping turtle in the Fitzroy River Basin is highly divergent at both mitochondrial and nuclear genetic markers from southern populations in the Burnett, Mary and Kolan Rivers (Todd et al. 2013; Todd 2013). However, populations from the Burnett, Mary and Kolan Rivers are highly differentiated from each other at nuclear genetic markers; hence, each river contains a genetically distinct population (Todd et al., 2013; Todd 2013). There is little genetic differentiation between populations within basins (Todd et al. 2013; Todd 2013), suggesting that significant barriers would fragment populations of this species.

As shown for other aquatic species in the Mary River (e.g. Mary River cod, lungfish and Mary River turtle), the estuarine reach that separates Tiana Creek from the remainder of the Mary River is likely a significant natural long-term barrier to connectivity, and thus the population of white-throated snapping turtles in Tiana Creek is likely differentiated from the population in the main Mary River basin (noting this precautionary population assessment requires validating with samples from Tiana Creek).

Area of Occupancy

The white-throated snapping turtle is restricted to the Fitzroy, Mary and Burnett River basins, and adjacent small, coastal systems, including the Kolan and Burrum Rivers (Hamann et al., 2007). White-throated snapping turtle is widely distributed in the main stem and major tributaries of the Fitzroy, Burnett and Mary Rivers (Todd 2013). The tributaries of the Mary River from which white-throated snapping turtle have been reported are Tinana, Wide Bay, Obi Obi and Yabba creeks (Limpus 2008). There are no records of white-throated snapping turtle from Six Mile Creek.

Habitat

White-throated snapping turtles are habitat specialists that prefer permanent, clear, well oxygenated water that is flowing and contains shelter (e.g. large woody debris and undercut banks) (Todd 2013). The species has also been recorded in non-flowing waters, such as

impoundments (e.g. Borumba Dam, Imbil Weir, Mary River Barrage) but only in low numbers (Threatened Species Scientific Committee 2014a). Within the greater Fitzroy, Burnett and Mary River basins, this species has been recorded almost exclusively in close association with permanent flowing stream reaches that are typically characterised by a sand-gravel substrate with submerged rock crevices, undercut banks and / or submerged logs and fallen trees, and are rarely found in reaches without such refuge. (Hamann et al. 2007; Limpus et al. 2007; Limpus et al. 2011). Across its distribution, individuals have been recorded from both shallow and deep, slow flowing pools (Hamann et al. 2007).

White-throated snapping turtles are rarely present in water bodies that are isolated from flowing streams, such as farm dams or sewage treatment ponds, suggesting that the species does not move extended distances over dry land (Hamann et al. 2007). However, white-throated snapping turtles have been observed walking short distances from drying waterholes to nearby water bodies (Limpus et al. 2007).

Water Quality

There are no published water quality tolerances for white-throated snapping turtle, although they are known from flowing streams with generally clear, well-oxygenated water (i.e. low turbidity and high dissolved oxygen). The species is one of several turtle species that can absorb oxygen from water via cloacal respiration, which reduces energy expenditure and reduce exposure to threats, especially for juveniles (DotEE 2017a). Thus high dissolved oxygen concentrations is a key water quality characteristic for this species. It is also suggested that high suspended sediment concentrations and high water temperatures (the latter of which limits dissolved oxygen concentrations) likely decrease diving duration, which may increase exposure to predation, especially juveniles (DotEE 2017a)

Flow Requirements

White-throated snapping turtle generally prefer flowing waterways with clear water of variable depth (i.e. shallow riffles, shallow runs and deep pools). The species is uncommon in non-flowing waters and isolated waterbodies that are not connected to flowing water habitats. As the species is a cloacal ventilating species, it is thought that it would not function well in deeper habitats of impounded waters where dissolved oxygen concentrations are likely low (Limpus 2008), although could occur in shallow upper reaches of impoundments where there are inflows.

Foraging and Movement

White-throated snapping turtles feed primarily on aquatic plants, and fruits and leaves from overhanging riparian vegetation (Limpus et al. 2007; DotEE 2017a). They may also eat

periphyton, freshwater bivalves and insects, particularly when plant food resources are limited (Limpus et al. 2007). The diet of juveniles is dominated by invertebrates, whereas the diet of larger individuals (i.e. standard carapace length < 6 cm) is dominated by plant material (Limpus 2008).

The species generally has small home ranges of less than 1 km with some individuals recorded as moving up to 10 km (DotEE 2017a), although population genetic analyses indicate high rates of gene flow within basins (Todd 2013), indicating that at least some individuals move larger distances within basins. There longer-range dispersal events often exceed the distance of flowing riverine habitat between dams and weirs, indicating that barriers to long-range movement is a significant impact on the species (DotEE 2017a).

The estuarine reach that separates Tiana Creek from the remainder of the Mary River is a significant natural long-term barrier to connectivity for Mary River cod, lungfish and Mary River turtle, and thus movement between Tiana Creek and the Mary River is also unlikely for white-throated snapping turtles, but to date has not been confirmed for this species.

Habitat Critical to Survival

Habitat critical to the survival of natural populations of white-throated snapping turtle includes (DotEE 2017a):

- Riverine systems with permanent generally clear, well-oxygenated flowing water, and deeper waterholes, in the Fitzroy, Burnett, Kolan, Burrum and Mary River Basins
- all currently known and potential nesting sites (i.e. sandy banks within 50 m of the channel)

Breeding Cycle

The life history of white-throated snapping turtles is characterised by a long life span and slow growth to maturity (Threatened Species Scientific Committee 2014a). The age at first breeding is approximately 15 to 20 years (Limpus et al. 2011; DotEE 2017a). Breeding occurs once per year, mostly during autumn and winter, with adult females breeding in each successive year unless the turtle has been injured or debilitated, or riverine habitat has been altered (e.g. water extraction, drought or weeds) (Threatened Species Scientific Committee 2014a). Females generally nest on sandy or loam alluvial banks over an extended breeding season of some 7 months from autumn to spring (DotEE 2017a). Females lay a single clutch of eggs during the breeding season, with an average of 14 eggs

per clutch (Hamann et al. 2007; Limpus et al. 2011). Nests are generally laid in areas of low canopy cover and in areas of dense grass cover; however, dense weeds at the water's edge may limit suitability of potential nesting banks (Hamann et al. 2007; Limpus et al. 2011). Nests are an average of 16.6 m from the water's edge, with eggs laid in deep chambers (greater than 20 cm in depth) and on banks with a slope of up to 26.5° (Hamann et al. 2007; Limpus et al. 2007; Limpus et al. 2011). However, nests have been recorded up to 60 m from the water (Hamann et al. 2007). White-throated snapping turtles will repeatedly use specific areas of banks over multiple years (Limpus et al. 2007).

There is no parental care, and egg and small juvenile survival is typically low (Heppell et al. 1996; Hamann et al. 2007). There is abundant evidence of nesting in all three river basins (i.e. Fitzroy, Burnett and Mary River Basins), but most eggs are lost to predation or trampling by stock (Hamann et al. 2007; Limpus et al. 2011).

Threats

The principal threat to white-throated snapping turtles is the excessive loss of eggs and hatchlings due to predation (Threatened Species Scientific Committee 2014a; DotEE 2017a). Primary predators include feral (e.g. foxes, dogs, pigs and cats) and native (e.g. water rats and goannas) animals. Trampling of nests by cattle is also a major threat.

Suitable habitat for white-throated snapping turtle is often limited, has often become fragmented by dams and weirs, and reduced in quality by inappropriate water allocation. Mortality of turtles in dam outlet works and over dam spillways is also a threat to this species (Hamann et al. 2007).

Other threats to this species are:

- stocking of fish into dam impoundments for recreational fishing
- · recreational fishing resulting in hook injuries
- · boat strike
- · loss of nesting habitat to weed infestation in the riparian zone
- · dense aquatic weeds in the waterways, and
- water extraction for agriculture and irrigation (Limpus et al. 2011).

No introduced diseases have been recorded as a threat to this species (DotEE 2017a).

Summary of Draft Recovery Plan

The objectives of the draft recovery plan (DotEE 2017a) are to:

- ensure a self-sustaining healthy population structure in all catchments in which the white-throated snapping turtle occurs; and
- ensure an ecologically functional wild population of white-throated snapping turtle that, with limited species-specific management, has a high likelihood of persistence in nature.

The strategies to achieve the plan's objectives are to:

- · Substantially improve the recruitment of hatchlings into the population;
- · Reduce the incidence of adult mortality and injury;
- Maintain and/or improve stream flow and habitat quality throughout the species' distribution;
- · Maintain and/or improve the connectivity within populations throughout each catchment; and
- · Increase public awareness and participation in conservation of the species and its habitat.

1.2 Mary River Cod

Status

The Mary River cod (*Maccullochella mariensis*) is listed as endangered under the EPBC Act. It is also listed as a 'no take' species under the Queensland *Fisheries Act 1994,* with the exception of specific impoundments nominated by DAF where it has been stocked.

Population

The species is restricted to the Mary River basin, where fewer than 600 individuals may occur, primarily in three main tributaries of the Mary River (i.e. Tiana-Coondoo, Obi Obi and Six Mile Creeks (Simpson & Jackson 1996; Threatened Species Scientific Committee 2016), although recent survey work suggests that the main stem of the Mary River, and potentially other significant tributaries, contains a larger population of cod than previously recognised (SKM 2007 and references therein). Nonetheless, the population size appears

to be decreasing ((Simpson & Jackson 1996), with effective population size estimates being very low (18 - 56), suggesting the breeding population of Mary River cod is very small (Huey et al. 2013).

Population genetic analyses indicate two genetically distinct populations of Mary River cod – main Mary River system and Tiana-Coondoo Creek, which are separated by estuarine waters (Huey et al., 2013). While past stocking from Tiana Creek to the Mary River has not appeared to influence population genetic structure, the genetic results indicate that stocking programs should keep populations from the Mary River (including Six Mile Creek) totally separate from those of Tiana Creek (Huey et al., 2013).

Area of Occupancy

The Mary River cod is endemic to the Mary River system. While of low abundance in most reaches, Mary River cod is reported to occur in the main stem of the Mary River (SKM 2007 and references therein) and several tributaries, with Six Mile Creek, Obi Obi Creek and Tiana Creek likely having larger numbers of cod than Yabba Creek, Munna Creek and Booloumba Creek (Simpson & Jackson 1996; Threatened Species Scientific Committee 2016). The population in Six Mile Creek is distributed from the confluence with the Mary River to well upstream of Lake Macdonald, and has historically considered to be in a stable condition (Simpson & Jackson 1996). In Tinana-Coondoo Creek, Mary River cod occur up to seventy kilometres upstream of the confluence with the Mary river, of which only 25-30% is considered suitable habitat. In Obi Obi creek, the species range extends approximately 10 km upstream from the confluence with the Mary River (Simpson & Jackson 1996). A number of impoundments in South East Queensland have been stocked with Mary River cod for recreational fishing (Huey et al. 2013).

Habitat

The pool habitats within Obi Obi, Six Mile and Tinana-Coondoo Creeks are known strongholds for Mary River cod, although pools along the main Mary River, and other significant tributaries of the Mary River, may contain more cod than previously recognised (SKM 2007 and references therein). Suitable pools are of variable depth (up to 3 m) and usually occur along pool and shallow riffle or run sequences (Simpson & Jackson 1996). The in-stream habitat features used for foraging and nesting by Mary River cod include submerged large woody debris, undercut banks, rock ledges and boulders (DoE 2014b). Reaches of creek with intact riparian vegetation are also favoured by the species as it provides shade and a supply of woody debris (Hydrobiology 2008a; Threatened Species Scientific Committee 2016).) Larval and juvenile fish habitat preferences are variable between the tributaries of its distribution in the Mary River, with no single habitat type (small wood debris, large woody debris, marginal vegetation, leaf litter) or substrate type (sand or mud) dominant (Dunlop 2016).

Water Quality

The reported water quality ranges for sites where Mary River cod have been caught are:

- pH = 6.0 7.3
- · conductivity (μ S/cm) = 100 800
- temperature (°C) = 15.7 29.0, and
- · dissolved oxygen (mg/L) = 3.9 9.7 (Hydrobiology 2008a).

Of the listed water quality parameters, temperature and dissolved oxygen are the most important, as high temperatures and low dissolved oxygen levels can be lethal to the species (DNRM, pers. Com.).

Flow Requirements

Adults of the species typically prefer low flowing water of suitable depth (i.e. 1 - 3 m) (Hydrobiology 2008a, 2008b), and generally avoid shallow (<1 m) areas. During periods of high flow, they shelter amongst woody debris and undercut banks, although movement may be triggered by moderate flows (Hydrobiology 2008b). Stable base flows maintain shallow riffles, which also maintain dissolved oxygen levels (Hydrobiology 2008b). Mary River cod have been observed dispersing from 10 km to 70 km over several months following high flows (Simpson & Jackson 1996). Spawning is triggered by light episodic rainfall (up to 20 mm) and associated flows, when it coincides with a water temperature of at least 19 °C and the full moon phase (R Manning 2014, pers. comm.).

Foraging and Movement

Mary River cod typically have relatively narrow home ranges, occupying a particular pool for extended periods (Simpson & Jackson 1996). Within their ranges, movement tends to be upstream during the summer months when rainfall and flows are higher, connecting pools, and downstream or into larger tributaries during the winter months (Threatened Species Scientific Committee 2016). Population genetic analyses indicate some level of connectivity throughout the main Mary River (Huey et al. 2013), suggesting at least some individuals move larger distances from home ranges, although it is not known if longer-range dispersal is by juveniles or adults (or both).

Mary River cod are predatory and generally feed on smaller fish and crustaceans, most commonly during dawn and dusk; but the species is also known to occasionally consume waterbirds and other fauna (Threatened Species Scientific Committee 2016). The species often forages on prey immediately downstream of riffles, presumably due to a constriction of the watercourse and the concentration of prey items (SKM 2007 and references therein) This suggests that shallower riffle habitats is important habitats for Mary River cod, although this habitat may not be commonly occupied by the species.

Habitat Critical to Survival

Habitat critical to the survival of natural populations of Mary Cod includes the following features in streams and waterways within the Mary River:

- Deep, slow-flowing, clay-lined pools with submerged large woody debris and other complex submerged habitat features (e.g. boulders, undercut banks) along the Mary River and major tributaries, especially Six Mile Creek, Obi Obi Creek and Tiana Creek
- · Shallow flowing riffle habitats connecting the pool habitats, to maintain dissolved oxygen concentrations
- For breeding: water temperature of at least 19°C coinciding with a rise (e.g. approximately 30cm) in discharge from early spring to early summer

Breeding Cycle

Mary River cod mature at approximately 38 cm and are considered to be a large, slow growing, long-lived fish with relatively low fecundity (Threatened Species Scientific Committee 2016).). The cod is presumed to spawn more than once a year, during spring and into early summer (Simpson & Jackson 1996). Spawning is triggered by light episodic rainfall (up to 20 mm) and associated increases in flows, when it coincides with a water temperature of at least 19 °C (Hydrobiology 2008b).

Eggs are typically deposited in a nest formed by a hollow log or similar habitat features (e.g. submerged open pipe) (Simpson & Jackson 1996). The male will subsequently guard the eggs until they begin to hatch towards the end of the fourth day at 20°C. The male will continue to guard the brood until they are ready to search for food between seven and nine days after hatching (Simpson & Jackson 1996). In the event that conditions do not coincide i.e. water temperature of at least 19°C, and light episodic rainfall up to 20 mm, female Mary

River cod will reabsorb their eggs and will not spawn (Threatened Species Scientific Committee 2016).

Threats

Threats to Mary River cod, include:

- Overfishing overfishing during the late 1800's and early 1900's saw the removal of large numbers of fish (Simpson & Jackson 1996). Currently, fishing for Mary River cod is prohibited in the Mary River, however there is evidence that illegal capture of the Mary River cod still occurs (Simpson & Jackson 1996).
- Habitat degradation specifically clearing of riparian zones, which exposes bank soil to erosion and leads to sedimentation of pool habitats (Simpson and Jackson 1996). Loss of riparian vegetation also reduces the input of branches and other habitat elements that are preferred by Mary River cod
- Dams and weirs impose barriers to movement; while long-range dispersal by cod is not frequent as they generally have relatively small home ranges, periodic movement over longer distances is likely to be important for the long-term survival of the species (Simpson & Jackson 1996). Cold-water releases from dams and altered flows downstream of dams may also impact breeding and / or survival of larvae (DoE SPRAT Profile)
- Pollution various sources of pollution may impact the suitability of water quality for Mary River cod, with increased nutrients and reduced dissolved oxygen known water quality issued for Mary River cod (Simpson & Jackson 1996), and
- Introduced and invasive species may increase competition for food or habitat resources, or may prey on larval and juvenile cod (Simpson & Jackson 1996).

No introduced diseases have been recorded as a threat to this species (Simpson & Jackson 1996).

Summary of Recovery Plan

- 1. Establish a program of community involvement and education.
- 2. Review and develop regulations and administrative procedures to ensure protection of the Mary River cod and its habitats, including:
 - develop a translocation strategy to minimise impacts of non-endemic fish introductions.

- maintain and restore fish passage past weirs/dams
- develop and implement environmental flow guidelines.
- 3. Develop a plan to improve hatchery production of the Mary River cod, and restock throughout the former range.
- 4. Undertake research on key aspects of Mary River cod ecology and captivebreeding techniques.
- 5. Restore degraded Mary River cod habitats:
 - 5.1. Develop a strategic plan for restoration of cod habitats.
 - 5.2. Implement pilot habitat rehabilitation programs in key areas of the Mary River.

1.3 Mary River Turtle

Status

The Mary River turtle (*Elusor macrurus*) is listed as endangered under the EPBC Act, and endangered under the NC Act.

Population

There is limited data available on the population size of the Mary River Turtle (Limpus 2008), although effective population size estimates for the species in the wild were between 136 and 158 (Schmidt et al., 2017), suggesting that the breeding population of this species is very small. This is consistent with estimates that the nesting population has declined by 95% since 1974 (Flakus 2002). The current population is dominated by aging adults, but studies suggest that the proportion of immature turtles may be increasing (Limpus 2008).

Population genetic analyses indicate that most of the Mary River contains a genetically connected population of Mary River turtle, with the exception of Tinana Creek, which is a genetically unique population (Schmidt et al., 2017).

Area of Occupancy

The Mary River turtle has been recorded in the Mary River and several of its tributaries (e.g. Yabba Creek and Tiana Creek) between Kenilworth (260 km from the river mouth) and the

Mary River tidal barrage at Tiaro (Flakus 2002; Limpus 2008); Threatened Species Scientific Committee 2008, although has not been recorded from Six Mile Creek.

Habitat

The species is regularly associated with areas of submerged habitat, including sparse to dense aquatic plant coverage, woody debris and rock crevices (SKM 2007 and references cited therein). Similar to other reptiles, the Mary River turtle often basks on emerging rock and logs within the waterbody and along its banks (Cann & Legler 1994).

Much of the Mary River turtle habitat is surrounded by cleared grazing and agricultural land, although in such reaches the species has been caught in areas where the river is wide and there is trailing vegetation and in-stream habitat (Cann & Legler 1994). Some areas of Mary River turtle habitat retain some riparian and catchment vegetation, especially in upper catchment areas and along several tributaries.

Water Quality

There are no specific water quality tolerances that have been published for Mary River turtle; but like other turtles with aquatic cloacal respiration, Mary River turtle requires flowing, well-oxygenated sections of streams (Threatened Species Scientific Committee 2008). Declines in water quality may reduce the efficiency of cloacal respiration by Mary River turtle, which can reduce foraging efficiency and more frequently expose juveniles to predators at the water surface.

Flow Requirements

The Mary River turtle prefers habitats characterised by shallow, flowing streams with riffle zones and well-oxygenated water with connecting shallow runs and deep pools (depth ranging from approximately 1 m - 5 m) (Flakus & Connell 2008; Threatened Species Scientific Committee 2008). During flooding, the Mary River turtle takes refuge in backwaters until flow decreases to pre-flood levels (Sadlier et al. 2004). They are also known to swim upstream during moderate to high flow events, returning to the same pool once water levels recede (Flakus & Connell 2008).

Foraging and Movement

The Mary River turtle is omnivorous and feeds on aquatic plants (including algae) and invertebrates (including bivalves) (Cann & Legler 1994). Juvenile Mary River turtles eat

aquatic insect larvae, supplemented by freshwater sponges, aquatic plants including green algae, and fruits of some terrestrial trees (Flakus 2002; Micheli-Campbell et al. 2013).

Individuals of the species have well defined home ranges and many show strong site fidelity (Limpus 2008; Micheli-Campbell et al. 2013), although low spatial genetic variability through the main Mary River (excluding Tiana Creek) (Schmidt et al., 2017) suggest that at least some individuals undertake longer-range movement. Movement may be seasonal, with limited movement in winter but up to 2 km in search of breeding sand banks have been recorded during the early summer months (Sadlier et al. 2004).

Habitat Critical to Survival

Habitat critical to the survival of natural populations of Mary River turtle includes the following features in streams and waterways within their range:

- · wide sections of the river with trailing vegetation and in-stream habitat
- flowing streams with riffle zones and well-oxygenated water, and reaches with deep connected pools
- aquatic plants (including algae) and invertebrates (including bivalves)
- · sparsely vegetated sandy river banks in close proximity to riffles and pools.

Breeding Cycle

Mary River turtles live for between 30 and 80 years and do not breed until between 15 and 25 years of age (Limpus 2008). Sparsely vegetated sandy river banks in close proximity to riffles and pools are preferred nesting habitats, with these sites revisited across decades by the same individual (Flakus 2002; Limpus 2008). Breeding occurs only once every year with a clutch size of approximately 13 eggs. Successful hatching is dependent on 50 consecutive days of non-inundation after nesting (Cann & Legler 1994; Flakus 2002). For this reason, nests are typically located 5 m above the water level and up to 30 m inland from the watercourse. Nesting occurs in late October to December after the first significant summer rain (Cann & Legler 1994; Flakus 2002). Eggs have an incubation period of 50 - 56 days, depending on sand temperature (Cann & Legler 1994).

Threats

Major threats to the Mary River turtle include:

- Nest predation and reduced success of recruitment for twelve years during the 1960's and 1970's large numbers of Mary River turtle eggs were collected for commercial purposes (Flakus 2002; Limpus 2008; Schmidt et al., 2017). As a result, little to no recruitment occurred during this time and this resulted in poor breeding success of Mary River turtle for four decades (Flakus 2002; Schmidt et al., 2017). Furthermore, pressure from predation in nesting areas by foxes, goannas and wild dogs has continued to cause low hatching success of the Mary River (Flakus 2002; Limpus 2008).
- Dams impoundments do not provide suitable habitat for the Mary River turtle, having typically still water with low levels of dissolved oxygen that reduces the efficiency of cloacal respiration (Threatened Species Scientific Committee 2008).
 Dam also do not provide favourable conditions for food resources of Mary River turtles, not suitable nesting habitats.
- Habitat degradation including clearing of the riparian zone, weed invasion and trampling of nest sites by cattle (Threatened Species Scientific Committee 2008).

Summary of Recovery Plan

As there is no recovery plan for the Mary River turtle. The priority actions outlined in the Approved Conservation Advice (Threatened Species Scientific Committee 2008) have been listed below.

Habitat Loss, Disturbance and Modification

- · Monitor known populations to identify key threats.
- Monitor the progress of recovery, including the effectiveness of management actions and the need to adapt them if necessary.
- · Identify and protect areas critical to the survival of the species, such as nesting sites.
- · Involve sand mining lessees in protection of critical habitat.
- Ensure road widening and maintenance activities (or other infrastructure or development activities as appropriate) in areas where the Mary River Turtle occurs do not adversely impact on known populations.
- Manage any disruptions to water flows.
- Suitably control access to areas of critical habitat to minimise impacts on habitat and nesting.
- · Continue protection of Mary River Turtle eggs from illegal collection.

· Investigate formal conservation arrangements such as the use of covenants, conservation agreements or inclusion in reserve tenure.

Trampling, Browsing or Grazing

· Continue protection of Mary River Turtle eggs from trampling.

Animal Predation

· Undertake predator control in nesting areas of the Mary River Turtle.

Conservation Information

· Raise awareness of the Mary River Turtle within the local community.

Enable Recovery of Additional Sites and/or Populations

 Develop a headstart program to increase hatchling survival and allow recruitment into the population, including moving clutches to safe incubation sites, creating new sandbanks for nesting, re-planting macrophytes after flood scouring, and introducing snags to pools.

2 Vulnerable Species

An action is likely to have a significant impact on a vulnerable species if there is a real chance or possibility that it will:

- · lead to a long-term decrease in the size of an important population of a species
- · reduce the area of occupancy of an important population
- fragment an existing important population into two or more populations
- adversely affect habitat critical to the survival of a species
- · disrupt the breeding cycle of an important population
- modify, destroy, remove or isolate or decrease the availability or quality of habitat to the extent that the species is likely to decline
- · result in invasive species that are harmful to a vulnerable species becoming established in the vulnerable species' habitat
- · introduce disease that may cause the species to decline, or
- · interfere substantially with the recovery of the species.

2.1 Australian Lungfish

Status

Australian lungfish (*Neoceratodus forsteri*) is listed as vulnerable under the EPBC Act. It is also listed as a 'no take' species under the Queensland *Fisheries Act 1994*.

Population

As a result of significant changes to core lungfish habitat recruitment to the adult breeding population is considered to be unsustainable low, and thus population reductions over the next few generations are likely (Threatened Species Scientific Committee 2014b; DotEE 2017b).

Population genetic analyses indicate three natural and genetically discrete populations of Australian lungfish: Burnett River, main Mary River and Tiana Creek in the lower Mary River (Hughes et al. 2015). Genetic analyses confirm populations in the Brisbane River and Pine River have been translocated from the main Mary River population (Hughes et al. 2015).

Area of Occupancy

The main stem and major tributaries of the Burnett and Mary Rivers comprise the natural distribution of Australian lungfish (Hughes et al. 2015; DotEE 2017b), although translocated populations also occur in the Brisbane, Pine, Albert and Coomera Rivers (Threatened Species Scientific Committee 2014b). Australian lungfish have been periodically recorded in low abundance from Six Mile Creek.

Habitat

Australian lungfish are generally found in wide, slow-flowing reaches with deep pools (i.e. 2-3 m) and shallower sections (i.e. 1m deep) with abundant aquatic plant cover and overhaning riparian vegetation (Threatened Species Scientific Committee 2014b; DotEE 2017b). Riffles and runs may also be present among deeper pool habitats (Arthington 2009). Australian lungfish tend to inhabit reaches with structurally complex submerged habitat, including submerged logs, high aquatic plant cover and underwater crevices formed by rock scouring and / or undercut banks (Arthington 2009). Open water with an absence of complex in-stream structures, such as impounded waters, is not preferred habitat of the species, although the species is known from shallow, headwater areas of impoundments where more complex habitats are available (Arthington 2009).

Water Quality

The water quality ranges of sites where Australian lungfish have been caught are:

- · pH = 7.0 9.1
- · conductivity (μ S/cm) = 421 1165
- temperature (°C) = 10 30, and
- dissolved Oxygen (mg/L) = 6.9 15.6 (Hydrobiology 2008a).

Australian lungfish are not tolerant of, and will not disperse through, saline water (Pusey et al. 2004); hence, the genetic distinctness of the population in Tiana Creek, which is separated from the main Mary River by estuarine waters.

Flow Requirements

The specific flow requirements for lungfish are only partly understood. Adults of the species are found mostly in permanent still or slow flowing deep pools, or in shallow pools with high cover of submerged aquatic plants. Breeding and recruitment occurs under low flow conditions (i.e. water levels between 10 and 30 cm above cease to flow levels) (Hydrobiology 2008b).

Foraging and Movement

Lungfish are largely sedentary, having home ranges of less than 2 km, although long-term recoding of movement patterns shows that some individuals may move up to 5 km over a number of years (Kind 2002; Threatened Species Scientific Committee 2014b). Genetic connectivity in the Burnett and Mary Rivers, respectively, suggests that at least some individuals undertake longer-range movements (Hughes et al. 2015). Most movement is reported to occur during the summer months (Kind 2002), with individuals in upper reaches of impounded waters known to move larger distances to find suitable habitat than individuals from non-impounded waters (Kind 2002; Threatened Species Scientific Committee 2014b; DotEE 2017b).

Lungfish feed mostly at night on benthic invertebrates, amphibian larvae and aquatic plants, with juveniles consuming mostly benthic invertebrates (Threatened Species Scientific Committee 2014b). There are also reports of lungfish ingesting terrestrial plant leaves and fruits (DotEE 2017b).

Habitat Critical to Survival

Habitat critical to the survival of natural populations of Australian Lungfish includes the following features main stem rivers and significant tributaries of the Burnett and Mary Rivers:

 wide, slow-flowing or still permanent reaches with deep pools and shallower sections with abundant aquatic plant cover and structurally complex submerged habitat in the Burnett and Mary River basins, but also any breeding or foraging habitat in areas to which it has been translocated (e.g. Brisbane and North Pine Rivers)
Breeding Cycle

Male Australian lungfish begin breeding at approximately 15 years of age while females first breed at approximately 20 years of age (Threatened Species Scientific Committee 2014b). Most spawning occurs at night in flowing streams between August and December, before summer rains commence, and when water temperatures are higher than 20°C (Threatened Species Scientific Committee 2014b; DotEE 2017b). Spawning habitat is primarily dense aquatic plant meadows in shallow water with high dissolved oxygen (Threatened Species Scientific Committee 2014b; DotEE 2017b), although trailing roots and other habitats are occasionally used for spawning. Hatching of eggs occurs approximately one month after fertilisation (McGrouther 2013), with larval lungfish thought to have poor swimming abilities, and thus take refuge in aquatic plant meadows where water velocity is low and dissolved oxygen is high. Growth rates are typically slow, attaining 40 cm in length over approximately five years, although under optimal conditions growth rates are much faster (Arthington 2009).

Threats

The main threats to Australian lungfish are (Threatened Species Scientific Committee 2014b; DotEE 2017b):

- Dams and weirs which modify core habitat (i.e. convert flowing river habitats to impounded waters) and impact migration, including breeding migrations in search of suitable breeding locations. Stranding of lungfish below tidal barrages is reported, as is crowing in impoundments at times of very low water
- Altered Hydrology water extraction and water reductions during spawning times can expose shallow aquatic plant beds resulting of death of fertilised eggs, but similarly large water releases at the times may also have adverse impacts in recruitment
- Habitat degradation and impacted water quality such as through clearing of riparian zones, which exposes bank soil to erosion and leads to sedimentation of pool habitats, and invasive aquatic plants can reduce spawning habitat quality.
- Invasive species including stocked and pest fish which may compete with, and prey on eggs of , lungfish, with tilapia impacting the quality of spawning habitats by disturbing aquatic plants; invasive aquatic plants also impact the quality of lungfish spawning habitat.
- · Fishing recreational fishers are known to unintentionally catch the Australian lungfish

Summary of Draft Recovery Plan

Draft Recovery Plan objectives:

Enhance Australian lungfish populations throughout their range (particularly populations within the Burnett, Mary, Brisbane and North Pine River catchments) to a point where there can be assurance that the species no longer meets the criteria for listing and can be delisted from the national threatened species list under the EPBC Act.

Recovery strategies:

The strategies to achieve the plans' objectives are to:

- · Reduce the impacts of, and remove any redundant, artificial barriers
- · Manage waterways to optimise breeding and recruitment opportunities
- · Limit habitat degradation and maintain or enhance water quality
- · Reduce the impacts of introduced pest and weed species
- · Manage the impacts of water-based recreational activities
- · Address key knowledge gaps to improve Australian lungfish management
- Facilitate high levels of community participation and support in the implementation of Australian lungfish management strategies

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Appendix C Detailed Habitat Results

Six Mile Creek – Downstream of Lake Macdonald

Site: SMC4			Years surveyed		
	October 2013	November 2014	November 2015	October 2016	November 2017
Photograph (upstream view)					
Habitat features					
Adjacent land use	native forest, road	native forest	native forest, grazing	native forest, railroad, rural residential	native forest, grazing
Riparian vegetation cover	moderate	extensive	extensive	moderate	low to moderate
Dominant type of riparian vegetation	grass, shrubs, trees	shrubs, trees	grass, shrubs, trees	grass and trees	grass and trees
Mean wetted width (m)	6 m	8 m	9 m	12 m	9 m
Flow habitats	-	connected pool, run	connected pool, riffle, run	connected pool	connected pool, riffle, run
Mean water velocity (m/s)	no flow	0.01 m/s	0.12 m/s	0.015 m/s	0.155 m/s
Water level	low	at watermark	low	low	at watermark
Bed substrate composition	sand (60%), silt/clay (40%)	bedrock (5%), sand (85%), silt/clay (5%)	bedrock (5%), sand (80%), silt/clay (15%)	sand (90%), silt/clay (10%)	sand (90%), silt/clay (10%)
Stream bed stability	stable	stable	stable	stable	moderate erosion
Bank material	sand	sand, silt/clay	sand, silt/clay	sand, silt/clay	sand, silt, bedrock
Bank slope	steep to vertical	steep	moderate to steep	steep	flat and steep
Bank shape	convex and undercut	convex and concave	stepped and convex	convex and concave	CONVEX
erosion	moderale	low to high	none to low	IOW	IOW
Aquatic Plant cover in water	none	none	none	none	none
Aquatic plant cover on banks	-	none	low	low	low to moderate
Physical in-stream habitat features	individual logs and log jams, 5% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation	individual branches and branch piles, individual logs, terrestrial leaves and twigs, 5% cover trailing bank vegetation
In-stream barriers to aquatic fauna passage	bridge	bridge	bridge	bridge	bridge
Other disturbances	erosion, roads	erosion, roads	erosion, roads	erosion, roads	weeds, erosion, roads

no data

Site: SMC5			Years surveyed		
	October 2013	November 2014	November 2015	October 2016	November 2017
Photograph (upstream view)					
Habitat features					
Adjacent land use	native forest	native forest, grazing	native forest, grazing	native forest, rural residential	native forest, grazing
Riparian vegetation cover	moderate	low to moderate	extensive	moderate	moderate
Dominant type of riparian vegetation	shrubs, trees	shrubs, trees	shrubs, trees	grass and trees	grass, shrubs, trees
Mean wetted width (m)	7 m	10 m	8 m	10 m	10 m
Flow habitats	_	connected pool, riffle, run	connected pool, riffle, run	connected pool, riffle, run	connected pool, riffle, run
Mean water velocity (m/s)	slow	0.05 m/s	0.45 m/s	0.034 m/s	0.35 m/s
Water level	low	low	low	low	at watermark
Bed substrate composition	boulder (2%), cobble (5%), pebble (3%), gravel (20%), sand (70%)	pebble (5%), gravel (25%), sand (50%), silt/clay (20%)	boulder (7%), cobble (5%), pebble (5%), gravel (5%), sand (60%), silt/clay (18%)	boulder (2%), cobble (3%), pebble (5%), gravel (5%), sand (75%), silt/clay (10%)	bedrock (5%), boulder (15%), cobble (10%), pebble (5%), sand (50%), silt/clay (5%)
Stream bed stability	stable	stable	stable	stable	moderate erosion
Bank material	boulder, cobble, sand	pebbles, gravel, sand, silt	sand, boulder (deposited by trucks)	sand, silt/clay	gravel, sand, clay
Bank slope	steep	moderate to steep	moderate to steep	steep	steep to vertical
Bank shape	convex	stepped	convex and concave	convex and concave	concave
Extent of bank erosion	low	none to low	low	low	low
Aquatic Plant cover in water	none	none	none	none	low
Aquatic plant cover on banks	-	none	low	low	low
Physical in-stream habitat features	large woody debris, terrestrial leaves and twigs, 5% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 50% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation	branch piles, individual logs and log jams, terrestrial leaves and twigs, 15% cover trailing bank vegetation
In-stream barriers to aquatic fauna passage	bridge, build up of rocks and logs at the base of the bridge	bridge	bridge	bridge	bridge
Other disturbances	erosion, roads	erosion, roads, access tracks	erosion, roads, weeds, cleared vegetation	erosion, weeds, roads	weeds, roads

no data

Site: SMCDS05	Years surveyed		
	August 2015		
Photograph (upstream view)			
Habitat features			
Adjacent land use	industrial, native forest, camping		
Riparian vegetation cover	low		
Dominant type of riparian vegetation	grass, shrubs, trees		
Mean wetted width (m)	4 m		
Flow habitats	connected pool, riffle, run		
Mean water velocity (m/s)	2.0 m/s		
Water level	low		
Bed substrate composition	boulder (5%), cobble (10%), pebble (15%), gravel (15%), sand (45%), silt/clay (10%)		
Stream bed stability	moderate erosion		
Bank material	cobble, pebble, gravel, sand, silt/clay		
Bank slope	steep		
Bank shape	concave		
Extent of bank erosion	moderate		
Aquatic Plant cover in water	none		
Aquatic plant cover on banks	low		
Physical in-stream	individual branches and branch		
habitat features	piles, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation		
In-stream barriers to aquatic fauna passage	nil		
Other disturbances	erosion, roads, weeds		

Site: SMCDS04	Years surveyed		
	August 2015	October 2015	February 2018
Photograph (upstream view			
Habitat features			
Adjacent land use	native forest	native forest	native forest, road
Riparian vegetation cover	moderate	moderate	moderate
Dominant type of riparian vegetation	shrubs, ferns, trees	trees	grass, trees, shrubs
Mean wetted width (m)	15 m	15 m	10 m
Flow habitats	connected pool, run	connected pool	connected pool
Mean water velocity (m/s)	0.1 m/s	0.01 m/s	0.05 m/s
Water level	low	at watermark	at watermark
Bed substrate composition	sand (20%), silt/clay (80%)	sand (10%), silt/clay (90%)	artificial boulder (5%), cobble (5%), pebble (5%), gravel (5%), sand (15%), silt/clay (65%)
Stream bed stability	moderate aggradation	moderate aggradation	stable
Bank material	silt	silt	clay/silt
Bank slope	steep	vertical	moderate to steep
Bank shape	convex	wide lower bench, undercut 20– 30%	convex
Extent of bank erosion	low	moderate	some
Aquatic Plant cover in water	none	none	none
Aquatic plant cover on banks	low	low	low to moderate
Physical in-stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 15% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 10% cover trailing bank vegetation	individual branches and branch piles, dense terrestrial leaves and twigs, individual logs and log jams, 10% trailing bank vegetation, turtle basking spots present
In-stream barriers to aquatic fauna passage	nil	nil	nil
Other disturbances	roads	roads	roads, erosion

Sito: SMCDS03		Voars survovod	
Sile. Silic DS03	August 2015	October 2015	February 2018
Photograph (upstream view)	August 2019	October 2013	Image: Constraint of the second se
Habitat features			
Adjacent land use	native forest	native forest	native forest
Riparian vegetation cover	extensive	extensive	extensive
Dominant type of riparian vegetation	shrubs, trees	trees	shrubs, trees
Mean wetted width (m)	6 m	3 m	8 m
Flow habitats	connected pool, run	connected pool, riffle, run	connected pool, riffle, run
Mean water velocity (m/s)	0.2 m/s	0.2 m/s	0.02 m/s
Water level	low	at watermark	at watermark
Bed substrate composition	boulder, cobble, pebble, gravel, sand, silt/clay	boulder (3%), cobble (6%), pebble (5%), gravel (5%), sand (50%), silt/clay (31%)	gravel (5%), sand (15%), silt/clay (80%)
Stream bed stability	stable to moderate aggradation	moderate aggradation	moderate aggradation
Bank material	gravel, sand, silt/clay	sand, silt	silt/clay
Bank slope	steep	steep	steep to vertical
Bank shape	concave	concave	concave
Extent of bank erosion	low	moderate	moderate
Aquatic Plant cover in water	none	none	none
Aquatic plant cover on banks	low	low	low
Physical in-stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 10% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 10% cover trailing bank vegetation	individual branches and branch piles, sparce terrestrial leaves and twigs, individual logs and log jams, 5% cover trailing vegetation, turtle basking spots present
In-stream barriers to aquatic fauna passage	bridge	bridge	bridge
Other disturbances	access tracks	erosion, roads	erosion

Site: SMCDS02		Years surveyed	
	August 2015	October 2015	February 2018
Photograph (upstream view)			
Habitat features			
Adjacent land use	native forest	native forest, residential	native forest
Riparian vegetation cover	low	moderate	mocerate
Dominant type of riparian vegetation	shrubs, trees	shrubs, trees	shrubs, trees
Mean wetted width (m)	4 m	4 m	4 m
Flow habitats	connected pool, riffle, run	connected pool, riffle, run	connected in-channel pool, riffle, run
Mean water velocity (m/s)	0.5 m/s	0.01 m/s	0.05 m/s
Water level	low	at watermark	at watermark
Bed substrate composition	boulder (10%), cobble (10%), pebble (10%), gravel (15%), sand (50%), silt/clay (5%)	cobble (5%), pebble (2%), gravel (10%), sand (77%), silt/clay (10%)	cobble (5%), pebble (20%), gravel (25%), sand (25%), silt/clay (25%)
Stream bed stability	moderate erosion	moderate aggradation	stable
Bank material	sand, silt	cobble, sand, silt	clay/silt
Bank slope	moderate to steep	moderate	moderate
Bank shape	convex	concave and undercut 20–25%	uncercut 10%, wide lower brank, convex and concave on the left bank; convex and concave on the right bank
Extent of bank erosion	low	moderate	-
Aquatic Plant cover in water	none	none	low
Aquatic plant cover on banks	low	low	low to moderate
Physical in-stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 10% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 15% cover trailing bank vegetation	individual branches and branch piles, individual ogs and log jams, 10% cover trailing bank vegetation
In-stream barriers to aquatic fauna passage	nil	nil	nil
Other disturbances	erosion, roads	cleared vegetation, roads	tyre debris, roads

no data

Site: SMCDS01	Years surveyed			
	August 2015	October 2015	February 2018	February 2018
Photograph (upstream view)				
Habitat features				
Adjacent land use	native forest, rural	native forest, recreation, SEQ Water	dam, native forest	native forest, national park
Riparian vegetation cover	moderate	moderate	little upstream, extensive downstream	extensive
Dominant type of riparian vegetation	grass, shrubs, trees	shrubs, trees	grasses and trees upstream, trees and others downstream	trees, shrubs
Mean wetted width (m)	6 m	5 m	17 m	12 m
Flow habitats	connected pool, run	connected pool, riffle, run	connected pool, riffle	isolated in-channel pool
Mean water velocity (m/s)	0.5 m/s	0.15 m/s	0.01 m/s	0.01 m/s
Water level	low	at watermark	at watermark	-
Bed substrate composition	boulder (5%), cobble (5%), pebble (10%), gravel (10%), sand (30%), silt/clay (4%)	boulder (2%), pebble (3%), gravel (10%), sand (70%), silt/clay (15%)	boulder (15%), cobble (10%), pebble (15%), gravel (25%), sand (10%) silt/clay (25%)	gravel (10%), sand (10%), silt/clay (80%)
Stream bed stability	stable	moderate aggradation	stable	high
Bank material	pebble, gravel, sand, silt	gravel, sand, silt	clay, silt, pebble, cobble, concrete boulder	silt/clay
Bank slope	steep	moderate	steep, vertical	moderate to steep
Bank shape	convex	concave and undercut 20–25%	convex	-
Extent of bank erosion	none	low	moderate	-
Aquatic Plant cover in water	low	moderate	low	moderate
Aquatic plant cover on banks	low	high	low to moderate	moderate
Physical in-stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 15% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 25% cover trailing bank vegetation	individual branches, terrestrial leaves and twigs, 5% cover trailing bank vegetation	large woody debris, 50% cover trailing bank vegetation
In-stream barriers to aquatic fauna passage	dam wall upstream of site	dam wall upstream of site	dam wall upstream of site	old culvert upstream
Other disturbances	weeds	cleared vegetation, roads	roads	culvert

no data

Six Mile Creek – In Lake Macdonald

Site: DS Lake	Years surveyed			
	August 2015	October 2015	February 2018	February 2018
Photograph (upstream view)				
Habitat features				
Adjacent land use	residential	native forest, residential	recreation, water treatment	recreation, water treatment
Riparian vegetation cover	extensive	extensive	little	some
Dominant type of riparian vegetation	grass, shrubs, trees	grass, trees	grass, trees, shrubs	grass, trees, shrubs
Mean wetted width (m)	100 m	300 m	>100 m	>100 m
Flow habitats	connected pool	connected pool	connected in-channel pool	connected pool
Mean water velocity (m/s)	<0.1 m/s	0.01 m/s	<0.1 m/s	<0.1 m/s
Water level	low	at watermark	at watermark	at watermark
Bed substrate composition	gravel (5%), sand (15%), silt/clay (80%)	gravel (3%), sand (90%), silt/clay (7%)	boulder (5%), cobble (10%), pebble (10%), gravel (10%), sand (55%), silt/clay (10%)	pebble (5%), gravel (10%), sand (75%), silt/clay (10%)
Stream bed stability	stable	moderate aggradation	moderate aggradation	moderate aggradation
Bank material	sand, silt/clay	sand, silt	clay/silt	clay/silt and sand
Bank slope	low	low	low to moderate	low
Bank shape	concave	concave and undercut 2%	concave	concave
Extent of bank erosion	none	none to low	moderate	low
Aquatic Plant cover in water	low	moderate	high	high
Aquatic plant cover on banks	low	low	low	low
Physical in-stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 5–25% cover trailing bank vegetation	individual branches, individual logs and log jams, terrestrial leaves and twigs, 5% cover trailing bank vegetation	individual branches, sparce terrestrial leaves and twigs, individual logs, 25% cover trailing vegetation	individual branches, sparce terrestrial leaves and twigs, individual logs, 50% cover trailing vegetation,
In-stream barriers to aquatic fauna passage	dam wall downstream of site	dam wall downstream of site	dam wall downstream of site	dam wall downstream of site
Other disturbances	weeds, cleared vegetation	weeds, cleared vegetation	cleared vegetation, roads, weeds, cleared vegetation	weeds, cleared vegetation, access tracks

Site: US Lake	Years surveyed		
	August 2015	October 2015	
Photograph (upstream view)			
Habitat features			
Adjacent land use	native forest, rural residential	native forest, residential	
Riparian vegetation cover	low	low to moderate	
Dominant type of riparian vegetation	grass, trees	grass, trees	
Mean wetted width (m)	>30 m	120 m	
Flow habitats	connected pool	connected pool	
Mean water velocity (m/s)	<0.1 m/s	0.02 m/s	
Water level	low	at watermark	
Bed substrate	cobble (5%), pebble (5%), gravel	sand (90%), silt/clay (10%)	
composition	(5%), sand (15%), silt/clay (70%)		
Stream bed stability	stable	moderate aggradation	
Bank material	sand, silt/clay	sand, silt	
Bank slope	low to moderate	low	
Bank shape	convex	concave	
Extent of bank erosion	none to low	none	
Aquatic Plant cover in water	low	moderate	
Aquatic plant cover on banks	low	low	
Physical in-stream	individual branches and logs, terrestrial leaves and twice 5-25%	individual branches and logs, terrestrial leaves and twice, 10	
nabitat isatures	cover trailing bank vegetation	15% cover trailing bank vegetation	
In-stream barriers	dam wall downstream of site	dam wall downstream of site	
to aquatic fauna passage			
Other disturbances	cleared vegetation, weeds	cleared vegetation, weeds	

Six Mile Creek – Upstream of Lake Macdonald

Site: CU02	Years surveyed		
	August 2015	October 2015	
Photograph (upstream view)			
Habitat features			
Adjacent land use	native forest	native forest	
Riparian vegetation cover	extensive	moderate to extensive	
Dominant type of riparian vegetation	grass, trees	grass, trees	
Mean wetted width (m)	19 m	13 m	
Flow habitats	connected pool	connected pool	
Mean water velocity (m/s)	<0.1 m/s	0.01 m/s	
Water level	low	at watermark	
Bed substrate composition	gravel (5%), sand (10%), silt/clay (85%)	sand (20%), silt/clay (80%)	
Stream bed stability	stable	moderate erosion	
Bank material	silt	silt	
Bank slope	low	low and steep	
Bank shape	convex	concave, wide lower bench, undercut 20%	
Extent of bank erosion	low	low	
Aquatic Plant cover in water	low	moderate	
Aquatic plant cover on banks	low	low	
Physical in-stream	individual branches and branch	individual logs, terrestrial leaves	
habitat features	piles, individual logs and log jams, terrestrial leaves and twigs, 25– 75% cover trailing bank vegetation	and twigs, 25–50% cover trailing bank vegetation	
In-stream barriers to aquatic fauna passage	nil	nil	
Other disturbances	weeds	cleared vegetation, weeds	

Site: CU01	Years s	urveyed
	August 2015	October 2015
Photograph (upstream view		
Habitat features		
Adjacent land use	native forest	native forest
Riparian vegetation cover	extensive	moderate
Dominant type of riparian vegetation	grass, trees	grass, trees
Mean wetted width (m)	13 m	13 m
Flow habitats	connected pool	connected pool
Mean water velocity (m/s)	<0.1 m/s	0.01 m/s
Water level	low	at watermark
Bed substrate composition	pebble (5%), gravel (5%), sand (10%), silt/clay (80%)	sand (30%), silt/clay (70%)
Stream bed stability	moderate erosion	moderate erosion
Bank material	silt	silt
Bank slope	moderate	steep to vertical
Bank shape	convex, concave and undercut 60%	wide lower bench, undercut 60%
Extent of bank erosion	low	moderate
Aquatic Plant cover in water	low	moderate
Aquatic plant cover on banks	low	low
Physical in-stream	individual branches and branch	individual branches and branch
habitat features	piles, individual logs and log jams, terrestrial leaves and twigs, 75% cover trailing bank vegetation	piles, individual logs and log jams, terrestrial leaves and twigs, 15– 25% cover trailing bank vegetation
In-stream barriers	nil	nil
to aquatic fauna passage		
Other disturbances	weeds, access tracks, erosion	cleared vegetation, access tracks, weeds

Site: CU03	Years surveyed			
	August 2015	October 2015		
Photograph (upstream view)				
Habitat features				
Adjacent land use	native forest, rural	native forest, residential, recreational park		
Riparian vegetation cover	moderate to extensive	extensive		
Dominant type of riparian vegetation	grass, shrubs, trees	shrubs, trees		
Mean wetted width (m)	9 m	10 m		
Flow habitats	connected pool	connected pool		
Mean water velocity (m/s)	0.1 m/s	0.01 m/s		
Water level	low	at watermark		
Bed substrate	pebble (5%), gravel (10%), sand	sand (30%), silt/clay (70%)		
composition	(25%), silt/clay (60%)			
Stream bed stability	moderate aggradation	moderate aggradation		
Bank material	sand, silt	sand, silt		
Bank slope	low to moderate	low and steep		
Bank shape	convex	concave and undercut 10–30%		
Extent of bank erosion	moderate	low		
Aquatic Plant cover in water	high	moderate		
Aquatic plant cover on banks	high	moderate		
Physical in- stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 25% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 25% cover trailing bank vegetation		
In-stream barriers to aquatic fauna passage	nil	nil		
Other disturbances	weeds, roads	cleared vegetation, roads		

Site: SMCUS02	Years s	urveyed
	August 2015	October 2015
Photograph (upstream view		
Habitat features		
Adjacent land use	rural residential	native forest, park, recreation
Riparian vegetation cover	extensive	low to moderate
Dominant type of riparian vegetation	grass, shrubs, trees	grass, trees
Mean wetted width (m)	24 m	12 m
Flow habitats	connected pool	connected pool
Mean water velocity (m/s)	0.1 m/s	_
Water level	low	at watermark
Bed substrate composition	gravel (5%, sand (15%), silt/clay (80%)	sand (20%), silt/clay (80%)
Stream bed stability	stable	moderate aggradation
Bank material	sand, silt/clay	silt
Bank slope	low	low
Bank shape	convex	concave
Extent of bank erosion	none	none
Aquatic Plant cover	high	moderate
in water	hish	
Aquatic plant cover on banks	nign	IOW
Physical in-stream	individual branches and branch	individual branches and logs,
nabitat features	terrestrial leaves and twigs, 5–15% cover trailing bank vegetation	cover trailing bank vegetation
In-stream barriers to aquatic fauna passage	bridge	bridge
Other disturbances	cleared vegetation, weeds, roads	cleared vegetation, roads

no data

Site: SMCUS01		Years surveyed	
	August 2015	October 2015	February 2018
Photograph (upstream view)			
Habitat features			
Adjacent land use	native forest	native forest, conservation	native vegetation, rainforest
Riparian vegetation cover	extensive	moderate	extensive
Dominant type of riparian vegetation	grass, shrubs, trees	shrubs, trees	shrubs, trees
Mean wetted width (m)	7 m	5 m	8 m
Flow habitats	connected pool	connected pool	connected pool, riffle
Mean water velocity (m/s)	<0.1 m/s	0.01 m/s	no flow
Water level	low	at watermark	high
Bed substrate composition	gravel (5%), sand (25%), silt/clay (70%)	sand (40%), silt/clay (60%)	gravel (5%), sand (75%), silt/clay (20%)
Stream bed stability	stable	moderate aggradation	stable
Bank material	sand, silt	sand, silt	sand, silt/clay
Bank slope	moderate	vertical	moderate to vertical
Bank shape	convex and undercut 30%	undercut 60–70%	25% undercut, convex to concave
Extent of bank erosion	low	moderate	some
Aquatic Plant cover in water	none	moderate	none
Aquatic plant cover on banks	low	low	low
Physical in-stream habitat features	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 50% cover trailing bank vegetation	individual branches and branch piles, individual logs and log jams, terrestrial leaves and twigs, 15% cover trailing bank vegetation	individual branches and branch piles, sparce terrestrial leaves and twigs, individual logs and log jams, 90% cover trailing bank vegetation, limited basking spots and nesting habitat for turtles
to aquatic fauna passage			
Other disturbances	weeds	nıl	access tracks and roads

Site: SMCUS03	Years s	urveyed
	August 2015	October 2015
Photograph (upstream view		
Habitat features		
Adjacent land use	rural residential	native forest, residential
Riparian vegetation cover	extensive	moderate
Dominant type of riparian vegetation	shrubs, trees, ferns	shrubs, trees
Mean wetted width (m)	10 m	8 m
Flow habitats	connected pool	connected pool, riffle, run
Mean water velocity (m/s)	<0.1 m/s	0.05 m/s
Water level	low	at watermark
Bed substrate composition	boulder (2%), cobble (3%), pebble (10%), gravel (15%), sand (50%), silt/clay (20%)	pebble (20%), gravel (50%), sand (20%), silt/clay (10%)
Stream bed stability	stable	moderate erosion
Bank material	sand, silt/clay	sand, silt
Bank slope	moderate to steep	vertical
Bank shape	convex	undercut 90%
Extent of bank erosion	low	moderate
Aquatic Plant cover in water	none	none
Aquatic plant cover on banks	low	low
Physical in-stream habitat features	individual branches and logs, terrestrial leaves and twigs, 10% cover trailing bank vegetation	individual branches and logs, terrestrial leaves and twigs, 25– 50% cover trailing bank vegetation
In-stream barriers to aquatic fauna passage	culvert	culvert
Other disturbances	cleared vegetation, roads	nil

Appendix D Summary of Seqwater Water Quality Monitoring Data for Lake Macdonald

Water quality monitoring data collected by Seqwater was collated and summarised for:

- · Lake Macdonald at the dam wall
- · Lake Macdonald, mid lake
- Lake Macdonald, Six Mile Creek Arm
- · Cooroy Creek at Cooroy-Noosa Road, and
- · Lake Macdonald tailwater.

Lake Macdonald – Dam Wall

Parameter			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Physico-Chemical																
Temperature - Field	°C	_	121	14.9	20.1	23.3	26.5	29	31.9	6	18.5	19.4	25.15	27.3	27.75	27.9
pH - Field	pH unit	6.5-8.0	32	6.4	6.7	7.07	7.512	7.972	9.28	6	6.5	6.6	6.6	6.7	6.85	6.9
pH - Lab	ph Unit	6.5-8.0	129	6.2	6.7	6.8	6.9	7.1	8.9	1	6.5	6.5	6.5	6.5	6.5	6.5
Conductivity - Field	µS/cm	626	28	69	106.2	117	131	137.3	143	2	57	61	67	73	76	77
Conductivity	µS/cm	626	133	33	82	100	124.6	140	178	5	29	45.8	57	72	111	124
Dissolved Oxygen - Field	mg/L	_	121	3.4	4.5	5.8	7.4	8.19	8.9	6	1.8	2.8	4.3	5.2	5.65	5.8
Dissolved Oxygen - Field	% Saturation	85-110	26	60.3	77.2	85.3	93	98.675	108.5	0	_	_	_	_	_	_
Turbidity - Field	NTU	50	26	1.2	2.76	3.6	4.3	7.15	10	0	_	_	_	_	_	_
Turbidity - Lab	NTU	50	130	1.6	2.6	3.35	4.4	7.865	14	7	2.9	4.12	4.6	9.8	12.4	13
Suspended Solids	mg/L	6	160	1	2.5	4	5	10.05	40	81	1	4	5	6	8	17
Total Dissolved Solids	mg/L	_	0	_	_	_	_	_	_	2	70	73.6	79	84.4	87.1	88
Alkalinity - Total	mg/L	_	0	_	_	_	_	_	_	1	9	9	9	9	9	9
BGA - Field	cells/mL	_	1	279384	279384	279384	279384	279384	279384	0	_	_	_	_	_	_
BGA - Field	RFU	_	1	132.8	132.8	132.8	132.8	132.8	132.8	0	_	_	_	_	_	_
Eutrophic Depth	mg/L	_	5	0.6	0.84	1	1	1	1	0	_	_	_	_	_	_
Redox Potential	MV	_	6	54.6	57.7	131.9	153.3	160.65	163.1	0	_	_	_	_	_	_
SUVA	L/mg-M	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Nutrients																
Total Nitrogen as N	mg/L	0.5	162	0	0.41	0.52	0.82	1.1	2.43	88	0.09	0.39	0.48	0.59	0.8165	1.08
Total Dissolved Nitrogen	mg/L	-	1	0.2	0.2	0.2	0.2	0.2	0.2	67	0.09	0.3	0.39	0.46	0.709	0.8
Kjeldahl Nitrogen as N	mg/L	_	83	0	0.4	0.51	0.8	1.1	2.4	1	0.42	0.42	0.42	0.42	0.42	0.42
Total Oxidised N	mg/L	0.06	135	0	0.0029	0.027	0.05	0.122	0.986	81	0.001	0.001	0.002	0.011	0.043	0.2
Nitrate as N	mg/L	0.7	1	0.001	0.001	0.001	0.001	0.001	0.001	43	0.001	0.001	0.001	0.0106	0.0523	0.19
Nitrite as N	mg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001	43	0.001	0.001	0.001	0.004	0.0109	0.011
Organic Nitrogen	mg/L	0.42	120	0	0.388	0.57	0.832	1.07	2.33	0	_	_	_	-	_	_
Ammonia - Total as N	mg/L	0.02	135	0	0.0025	0.02	0.05	0.09	0.181	81	0.0025	0.007	0.023	0.07	0.1	0.795
Total Phosphorus	mg/L	0.05	161	0	0.014	0.02	0.04	0.057	0.27	87	0.0025	0.017	0.022	0.0328	0.0444	0.079
Total Dissolved Phosphorus	mg/L	_	1	0.006	0.006	0.006	0.006	0.006	0.006	67	0.0025	0.006	0.01	0.015	0.025	0.049
Filterable Reactive Phosphorus	mg/L	0.02	129	0	0.0016	0.004	0.01	0.029	0.18	75	0.0005	0.0005	0.002	0.0052	0.008	0.011
Phosphate	mg/L	_	5	0.0025	0.0025	0.0025	0.0038	0.0077	0.009	5	0.0025	0.0025	0.0025	0.0036	0.0069	0.008

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Other Organics																
DOC	mg/L	_	0	_	_	_	_	_	_	74	3.8	5.6	6.3	8.24	10	11.1
ТОС	mg/L	_	0	_	_	_	_	_	_	74	3.8	6	6.8	8.34	10.07	13.1
Major Cations & Anions																
Fluoride - Total	mg/L	_	0	_	_	_	_	_	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Chloride	mg/L	_	0	_	_	_	_	_	_	1	21	21	21	21	21	21
Magnesium - Total	mg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Silica - Reactive as SiO3	mg/L	_	0	_	_	_	_	_	_	53	0.34	1.592	3.54	5.844	7.464	8.49
Sulphide	mg/L	_	0	_	_	_	_	_	_	0	_	_	_	-	_	_
Sulphur-Total	mg/L	_	0	_	_	_	_	_	_	6	2	3	3.5	4	6.25	7
Total Metals																
Aluminium	µg/L	55	1	41	41	41	41	41	41	13	7	19	31	176.6	431.2	676
Antimony	µg/L	_	0	_	_	_	_	_	_	14	0.5	0.5	0.5	0.5	0.5	0.5
Arsenic	µg/L	13	0	_	_	_	-	-	-	13	0.25	0.25	0.5	0.6	0.78	0.9
Barium	µg/L	_	0	_	_	_	_	_	-	13	6	7	9	12.4	20.94	24
Beryllium	µg/L	_	0	_	_	_	_	_	_	13	0.5	0.5	0.5	0.5	0.5	0.5
Boron	µg/L	370	0	_	_	_	_	_	_	13	15	15	15	15	15	15
Cadmium	µg/L	0.2	0	_	_	_	_	_	_	13	0.05	0.05	0.05	0.05	0.05	0.05
Chromium	µg/L	1	0	_	_	_	_	_	_	13	0.5	0.5	0.5	0.5	0.7	1
Cobalt	µg/L	_	0	_	_	_	_	_	_	12	0.5	0.5	0.5	0.5	0.5	0.5
Copper	µg/L	1.4	0	_	_	_	_	_	_	13	0.5	0.5	0.5	1	1.4	2
Iron	µg/L	_	15	359	543	1100	1440	1857	2025	78	189	622.4	1000	1568	2151.5	7670
Lead	µg/L	3.4	0	_	_	_	_	_	_	13	0.5	0.5	0.5	0.5	1.9	4
Lithium	µg/L	_	0	_	_	_	_	_	_	12	0.5	0.5	0.5	0.9	1	1
Manganese	µg/L	1900	81	9	16	52	130	174	209	78	43	90.34	137.5	209.6	326.2	1750
Mercury	µg/L	0.06	0	_	_	_	_	_	_	14	0.05	0.05	0.05	0.05	0.05	0.05
Molybdenum	µg/L	11	0	_	_	_	_	_	_	13	0.5	0.5	0.5	0.5	0.5	0.5
Nickel	µg/L	_	0	_	_	_	_	_	_	13	0.5	0.5	0.5	0.5	0.5	0.5
Selenium	µg/L	5	0	_	_	_	_	_	_	14	1	1	1	1	1	1
Silver	µg/L	0.05	0	_	_	-	_	_	_	13	0.5	0.5	0.5	0.5	0.5	0.5
Strontium	µg/L	_	0	_	_	_	_	_	_	12	12	18	19.5	22	24.35	26
Thorium	µg/L	_	0	_	_	_	_	_	_	12	0.5	0.5	0.5	0.5	0.5	0.5
Tin	µg/L	_	0	_	_	_	_	_	_	12	0.5	0.5	0.5	0.5	3.875	8
Uranium	µg/L	_	0	_	_	_	_	_	_	13	0.5	0.5	0.5	0.5	0.5	0.5

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Vanadium	µg/L	_	0	_	_	_	_	_	_	12	5	5	5	5	5	5
Zinc	µg/L	8	0	_	-	_	_	_	-	13	2.5	2.5	2.5	7.8	24.2	38
Soluble Metals																_
Aluminium	µg/L	55	7	2.5	7	11	42.6	56.2	61	6	2.5	2.5	12	40	91	108
Arsenic	µg/L	13	0	-	-	_	_	-	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Barium	µg/L	-	0	-	-	-	-	-	-	1	5.8	5.8	5.8	5.8	5.8	5.8
Beryllium	µg/L	_	0	-	-	_	_	-	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Boron	µg/L	370	0	_	_	_	_	_	-	1	15	15	15	15	15	15
Cadmium	µg/L	0.2	0	-	-	_	_	-	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Chromium	µg/L	1	0	_	_	_	_	_	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Cobalt	µg/L	_	0	-	-	_	_	-	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Copper	µg/L	1.4	0	_	-	_	_	_	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Iron	µg/L	_	16	52	147	354	566	765	930	80	55	199	432	771.4	1321	6410
Lead	µg/L	3.4	0	_	_	_	_	_	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Lithium	µg/L	_	0	_	-	_	_	_	_	1	0.5	0.5	0.5	0.5	0.5	0.5
Manganese	µg/L	1900	81	0	5	10	25	81	104	80	1	6	18.6	60.6	153.4	1740
Molybdenum	µg/L	11	0	_	-	_	_	_	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Nickel	µg/L	_	0	_	-	_	_	_	_	1	0.5	0.5	0.5	0.5	0.5	0.5
Silver	µg/L	0.05	0	_	_	_	_	_	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Strontium	µg/L	_	0	_	-	_	_	_	_	1	19	19	19	19	19	19
Thorium	µg/L	_	0	_	_	_	_	_	-	1	0.5	0.5	0.5	0.5	0.5	0.5
Tin	µg/L	_	0	_	-	_	_	_	_	1	0.5	0.5	0.5	0.5	0.5	0.5
Uranium	µg/L	_	0	_	-	_	_	_	_	1	0.5	0.5	0.5	0.5	0.5	0.5
Vanadium	µg/L	_	0	_	-	_	_	_	_	1	5	5	5	5	5	5
Zinc	µg/L	8	0	_	-	_	_	_	_	1	2.5	2.5	2.5	2.5	2.5	2.5
Algae																
Chlorophyll a - Field	µg/L	5	1	10.8	10.8	10.8	10.8	10.8	10.8	0	-	_	_	_	_	_
Chlorophyll a	µg/L	5	132	0	6	10	14	20	37	6	9.1	11	16	18	20.25	21
Microbiological																
Cryptosporidium - Corrected	MPN/100mL	_	5	0.5	0.5	0.5	0.5	0.5	0.5	0	-	-	_	_	_	_
Cryptosporidium - Detected	MPN/100mL	_	18	0.5	0.5	0.5	0.5	1.0375	1.25	0	-	_	_	_	_	_
E.coli - Colilert	MPN/100mL	_	77	0.5	1.2	3	9	16.2	170	0	-	-	_	_	_	_
E.coli - MF	MPN/100mL	_	2	8	17.8	32.5	47.2	54.55	57	0	-	-	_	_	_	_
Enterococci - MF	MPN/100mL	_	79	0.5	0.5	2	4	23.5	440	0	_	_	_	_	_	_

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Giardia - Corrected	MPN/100mL	_	5	0.5	0.5	0.5	0.5	0.5	0.5	0	_	_	_	_	_	_
Giardia - Detected	MPN/100mL	_	19	0.5	0.5	0.5	0.7	3.625	25	0	_	_	_	_	_	_
Total Coliforms - Colilert	MPN/100mL	_	74	140	452	690	1600	2335	3100	0	_	_	_	_	_	_
Pesticide																
2.4-D	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4-DB	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4-DP	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4.5-T	µg/L	36	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4.6-T	µg/L	_	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05
2.6-D	µg/L	280	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05
3-Hydroxy Carbofuran	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
4-Chlorophenoxy acetic acid	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
4.4-DDD (pp-DDD)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
4.4-DDE (pp-DDE)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
4.4-DDT (pp-DDT)	µg/L	0.006	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Acrolein	µg/L	_	0	_	_	_	_	_	_	1	1	1	1	1	1	1
Aldicarb	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Aldrin (Eldrin)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
alpha-BHC (lindane)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Ametryn	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Amitrole	µg/L	_	0	_	_	_	_	_	_	1	0.05	0.05	0.05	0.05	0.05	0.05
AMPA	µg/L	_	0	_	_	_	_	_	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Asulam	µg/L	_	0	_	_	_	_	_	_	2	1	1	1	1	1	1
Atrazine	µg/L	13	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Azinphos methyl	µg/L	0.01	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Azinphos-ethyl	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Bendiocarb	µg/L	_	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Benomyl	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.006	0.0075	0.009	0.00975	0.01
beta-BHC (lindane)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Bioresmethrin	µg/L	_	0	_	_	_	_	_	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Brodifacoum	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Bromacil	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Bromophos-ethyl	µg/L	-	0	_	_	_	_	-	-	2	0.05	0.05	0.05	0.05	0.05	0.05
Bromoxynil	µg/L	_	0	-	-	-	-	_	-	2	0.025	0.025	0.025	0.025	0.025	0.025

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Parameter	Unite	WOO	Surface							Deep						
	Units	ngo	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Carbaryl	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Carbofuran	µg/L	0.06	0	-	-	-	-	-	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Carbophenothion	µg/L	_	0	-	-	-	-	-	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Chlordane - Total	µg/L	0.03	0	_	_	-	-	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Chlorfenvinphos - Total	µg/L	_	0	_	_	-	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Chlorothalonil	µg/L	_	0	_	_	-	-	_	_	2	1	1	1	1	1	1
Chlorpyrifos	µg/L	0.01	0	_	_	-	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Chlorpyrifos-methyl	µg/L	_	0	_	_	-	_	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1
Chlorsulfuron	µg/L	_	0	_	_	_	_	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1
cis-Chlordane	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Clopyralid	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Coumaphos	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Cyanazine	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Cyproconazole	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Cyprodinil	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Cyromazine	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
delta-BHC	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Demeton-O & Demeton-S	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Demeton-S methyl	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Diazinon	µg/L	0.01	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Dicamba	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Dichlorvos (DDVP)	µg/L	_	0	_	_	_	_	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1
Diclofop-methyl	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Dieldrin	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Difenoconazole	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Diflufenican	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Dimethoate	µg/L	0.15	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Diquat	µg/L	1.4	0	_	_	_	_	_	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Disulfoton	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Diuron	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Endosulfan - Total	µg/L	0.03	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan I (alpha isomer)	µg/L	-	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan II (beta isomer)	µg/L	-	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan sulfate	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001

Parameter			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Endrin	µg/L	0.01	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endrin aldehyde	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endrin ketone	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
EPN	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Ethion	µg/L	_	0	_	_	-	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Ethoprophos (Prophos)	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Fenamiphos	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Fenarimol	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Fenchlorphos (Ronnel)	µg/L	_	0	_	_	_	_	_	_	2	5	5	5	5	5	5
Fenitrothion	µg/L	0.2	0	_	_	_	_	_	_	2	1	1	1	1	1	1
Fensulphothion	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Fenthion	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Fenvalerate & Esfenvalerate	µg/L	_	0	-	-	_	-	-	-	1	0.25	0.25	0.25	0.25	0.25	0.25
Fipronil	µg/L	_	0	-	-	_	-	-	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Fluometuron	µg/L	_	0	_	_	-	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Fluroxypyr	µg/L	_	0	_	_	-	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Flusilazole	µg/L	_	0	_	_	-	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Glyphosate	µg/L	370	0	_	_	-	_	_	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Heptachlor	µg/L	0.01	0	_	_	-	_	_	_	2	0.0005	0.0005	0.0005	0.0005	0.0005	0.0005
Heptachlor epoxide	µg/L	_	0	_	_	_	-	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Hexachlorobenzene (HCB)	µg/L	_	0	_	_	-	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Hexazinone	µg/L	_	0	_	_	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Iprodione	µg/L	_	0	_	_	_	-	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Irgarol	µg/L	_	0	_	_	_	-	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Lindane (gamma-BHC)	µg/L	0.2	0	_	_	_	-	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Malathion (Maldison)	µg/L	0.05	0	_	_	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
MCPA	µg/L	_	0	_	_	_	-	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
MCPB	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Mecoprop	µg/L	_	0	-	_	_	-	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Methiocarb	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Methomyl	µg/L	3.5	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Methoxychlor	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Metolachlor	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Metribuzin	µg/L		0	_	_	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01

Parameter	Unite	WOO	Surface							Deep						
	Units	WQU	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Metsulfuron-methyl	µg/L	-	0	-	_	_	-	_	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Mevinphos	µg/L	-	0	-	-	-	-	-	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Molinate	µg/L	3.4	0	-	_	_	_	-	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Monocrotophos	µg/L	-	0	-	-	-	-	-	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Omethoate	µg/L	-	0	-	_	_	_	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Oryzalin	µg/L	-	0	-	-	-	-	-	-	2	0.025	0.025	0.025	0.025	0.025	0.025
Oxamyl	µg/L	-	0	-	_	_	_	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Oxychlordane	µg/L	-	0	-	_	_	_	-	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Oxyfluorfen	µg/L	-	0	-	_	_	_	-	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Paclobutrazol	µg/L	_	0	_	_	_	-	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Paraquat	µg/L	_	0	_	_	_	-	_	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Parathion	µg/L	0.004	0	_	_	_	-	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1
Parathion-methyl	µg/L	-	0	-	_	_	_	-	_	2	1	1	1	1	1	1
Penconazole	µg/L	-	0	-	_	_	_	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Pendimethalin	µg/L	_	0	_	_	_	-	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Permethrin	µg/L	_	0	_	_	_	-	_	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Phorate	µg/L	_	0	_	_	_	-	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Picloram	µg/L	_	0	_	_	_	-	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Piperonyl Butoxide	µg/L	_	0	_	_	_	-	_	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Pirimiphos-ethyl	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Pirimiphos-methyl	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Profenofos	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Prometryn	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Propazine	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Propiconazole	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Prothiophos (Tokuthion)	µg/L	_	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Pyrimethanil	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Silvex (2,4,5-TP/Fenoprop)	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Simazine	µg/L	3.2	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Sulprofos	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Sum of DDD + DDE + DDT	µg/L	-	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Tebuconazole	µg/L	_	0	-	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Tebuthiuron	µg/L	2.2	0	-	_	-	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Temephos	µg/L	-	0	_	_	_	_	-	_	2	0.01	0.01	0.01	0.01	0.01	0.01

Parameter			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Terbufos	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Terbuthylazine	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Terbutryn	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Tetrachlorvinphos	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Tetraethyl dithiopyrophos	µg/L	_	0	_	_	_	_	_	_	2	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Thiamethoxam	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Thiobencarb	µg/L	2.8	0	_	_	_	_	-	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Thiodicarb	µg/L	_	0	_	_	_	_	-	-	2	0.005	0.005	0.005	0.005	0.005	0.005
trans-Chlordane	µg/L	_	0	_	_	_	_	-	-	2	0.001	0.001	0.001	0.001	0.001	0.001
Triazophos	µg/L	_	0	_	_	_	_	_	_	2	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Trichlorfon	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Trichloronate (Trichloronat)	µg/L	_	0	_	_	_	_	_	_	2	0.25	0.25	0.25	0.25	0.25	0.25
Triclopyr	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Trifluralin	µg/L	2.6	0	_	_	_	_	_	_	2	5	5	5	5	5	5
Polyaromatic Hydrocarbon																
2-Chloronaphthalene	µg/L	_	0	_	_	_	_	_	_	3	0.05	0.05	0.05	0.05	0.05	0.05
2-Methylnapthalene	µg/L	_	0	_	_	_	_	_	_	3	0.05	0.05	0.05	0.05	0.05	0.05
Acenaphthene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Acenapthylene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Anthracene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[a]anthracene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[a]pyrene	µg/L	_	0	_	_	_	_	_	_	3	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Benzo[b+j]fluoranthene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[e]pyrene	µg/L	_	0	_	_	_	_	_	_	3	0.05	0.05	0.05	0.05	0.05	0.05
Benzo[ghi]perylene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[k]fluoranthene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Chrysene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Dibenzo[ah]anthracene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Fluoranthene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Fluorene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Indeno(1.2.3-cd)pyrene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Napthalene	µg/L	16	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Perylene	µg/L	_	0	_	_	_	_	_	_	3	0.05	0.05	0.05	0.05	0.05	0.05
Phenanthrene	µg/L	_	0	_	_	_	_	_	-	3	0.01	0.01	0.01	0.01	0.01	0.01

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Pyrene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Total PAH	µg/L	_	0	_	_	-	_	_	_	3	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Taste and Colour																
2-Methylisoborneol	ng/L	_	0	_	_	-	_	_	_	66	1	3.1	11.7	20.3	24.325	33.4
2-methylisoborneol + Geosmin	ng/L	_	0	_	_	-	_	_	_	28	2.1	4.68	12.05	15.68	22.64	24.2
Geosmin	ng/L	_	0	-	_	_	-	-	-	66	1	1	2.1	4.2	7.6	19.1
Volatile Organic Compound																
Benzene	µg/L	950	0	_	_	-	_	_	_	3	0.25	0.25	0.25	0.25	0.25	0.25
Toluene	µg/L	_	0	_	_	-	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Ethylbenzene	µg/L	_	0	_	_	-	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
meta & para-Xylene	µg/L	_	0	_	_	-	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
ortho-Xylene	µg/L	350	0	_	_	-	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Total Xylenes	µg/L	_	0	-	_	_	-	-	-	3	0.5	0.5	0.5	0.5	0.5	0.5
Hexaconazole	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01

Grey shading denotes parameters that exceed the water quality object / trigger value

Gold shading denotes parameters that have an LOR higher than the water quality objective / trigger value

Lake Macdonald Mid Lake

_		-	Surface						Беер								
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	
Physico-Chemical																	
Temperature - Field	°C	-	40	16.37	19.752	23.985	27.32	28.905	30	0	_	_	_	_	-	_	
pH - Field	pH unit	6.5-8.0	32	6.43	6.7	7	7.332	7.79	7.9	0	_	_	_	_	-	_	
pH - Lab	ph Unit	6.5-8.0	128	6.4	6.7	6.8	6.9	7.165	8.9	0	_	_	_	_	-	_	
Conductivity - Field	µS/cm	626	28	63	106	116.5	128.6	137.3	143	0	_	_	_	_	_	_	
Conductivity	µS/cm	626	132	35	81	100	124	143.6	192	0	_	_	_	_	_	_	
Dissolved Oxygen - Field	mg/L	_	40	4.9	6.34	7.64	8.46	9.14	9.28	0	_	_	_	_	_	_	
Dissolved Oxygen - Field	% Saturation	85-110	26	71.1	88.2	93.3	99.3	103.8	107.1	0	_	_	_	_	_	_	
Turbidity - Field	NTU	50	26	1.6	2.3	3.05	4	7.375	10.4	0	_	_	_	_	_	_	
Turbidity - Lab	NTU	50	129	1.3	2.5	3.4	4.24	7.9	20.4	0	_	_	_	_	_	_	
Suspended Solids	mg/L	6	135	0	2	3	4	8	13	67	1	3.2	5	6.8	8.7	20	
Total Dissolved Solids	mg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_	
Alkalinity - Total	mg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_	
BGA - Field	cells/mL	_	1	279400	279400	279400	279400	279400	279400	0	_	_	_	_	_	_	
BGA - Field	RFU	_	1	132.8	132.8	132.8	132.8	132.8	132.8	0	_	_	_	_	_	_	
Eutrophic Depth	mg/L	_	5	0.6	0.84	1.1	1.12	1.18	1.2	0	_	_	_	_	_	_	
Redox Potential	MV	_	6	45.1	49.5	76.65	135.9	150.975	156	0	_	_	_	_	_	-	
SUVA	L/mg-M	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_	
Nutrients							_										
Total Nitrogen as N	mg/L	0.5	137	0	0.412	0.6	0.868	1.2	2.11	76	0.09	0.37	0.475	0.58	0.7625	0.91	
Total Dissolved Nitrogen	mg/L	_	3	0.3	0.336	0.39	0.42	0.435	0.44	64	0.07	0.29	0.4	0.46	0.62	0.76	
Kjeldahl Nitrogen as N	mg/L	_	81	0	0.4	0.6	0.8	1.2	2.1	1	0.36	0.36	0.36	0.36	0.36	0.36	
Total Oxidised N	mg/L	0.06	136	0	0.0025	0.021	0.047	0.13475	0.945	70	0.001	0.001	0.001	0.01	0.03875	0.066	
Nitrate as N	mg/L	0.7	2	0.001	0.001	0.001	0.001	0.001	0.001	41	0.001	0.001	0.001	0.007	0.039	0.061	
Nitrite as N	mg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001	41	0.001	0.001	0.001	0.004	0.008	0.011	
Organic Nitrogen	mg/L	0.42	119	0	0.39	0.6	0.82	1.068	2.06	0	_	_	_	_	_	_	
Ammonia - Total as N	mg/L	0.02	136	0	0.0025	0.0125	0.03	0.07	0.091	70	0.0025	0.0025	0.0155	0.0504	0.0762	0.12	
Total Phosphorus	mg/L	0.05	136	0	0.01	0.02	0.04	0.06325	0.2	76	0.0025	0.017	0.022	0.034	0.054	0.262	
Total Dissolved Phosphorus	mg/L	_	3	0.006	0.0076	0.01	0.0124	0.0136	0.014	63	0.0025	0.006	0.01	0.0156	0.0227	0.043	
Filterable Reactive Phosphorus	mg/L	0.02	130	0	0.001	0.005	0.008	0.0231	0.09	70	0.0005	0.0005	0.002	0.005	0.0101	0.017	
Phosphate	mg/L	_	5	0.0025	0.0025	0.0025	0.0038	0.0077	0.009	0	_	_	_	_	_	_	

			Surface [Беер							
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum		
Other Organics																		
DOC	mg/L	-	2	5.4	5.54	5.75	5.96	6.065	6.1	69	3.9	5.5	6.3	8.54	9.64	10		
ТОС	mg/L	-	2	5.4	5.72	6.2	6.68	6.92	7	69	3.9	5.9	6.8	8.7	10	10.5		
Major Cations & Anions																		
Fluoride - Total	mg/L	_	0	_	-	_	_	_	_	0	_	_	-	_	_	_		
Chloride	mg/L	_	0	_	-	_	_	_	_	0	_	_	-	_	_	_		
Magnesium - Total	mg/L	_	0	-	-	_	_	_	_	0	_	_	_	_	_	-		
Silica - Reactive as SiO3	mg/L	_	2	4.24	4.632	5.22	5.808	6.102	6.2	47	0.45	2.554	3.67	5.488	7.777	9.59		
Sulphide	mg/L	_	0	-	-	_	_	_	_	0	_	_	_	_	_	-		
Sulphur-Total	mg/L	_	0	_	_	_	_	_	_	4	2	2.6	3	3.4	3.85	4		
Total Metals																		
Aluminium	µg/L	55	0	_	_	_	_	_	_	11	7	12	29	99	741.5	996		
Antimony	µg/L	_	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Arsenic	µg/L	13	0	_	_	_	_	_	_	11	0.25	0.25	0.6	0.6	0.7	0.8		
Barium	µg/L	_	0	_	_	_	_	_	_	11	4	6	7.2	10.9	48.05	75		
Beryllium	µg/L	-	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Boron	µg/L	370	0	_	_	_	_	_	_	11	15	15	15	15	15	15		
Cadmium	µg/L	0.2	0	_	_	_	_	_	_	11	0.05	0.05	0.05	0.05	0.05	0.05		
Chromium	µg/L	1	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	1.25	2		
Cobalt	µg/L	_	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Copper	µg/L	1.4	0	_	_	_	_	_	_	11	0.5	0.5	0.5	1	1.5	2		
Iron	µg/L	_	3	247	509	902	966.8	999.2	1010	67	340	754	1100	1628	2267	2930		
Lead	µg/L	3.4	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Lithium	µg/L	_	0	_	_	_	_	_	_	11	0.5	0.5	0.5	1	1.05	1.1		
Manganese	µg/L	1900	5	45	52.2	100	108.4	115.6	118	67	41	88.48	130	230.6	341.5	642		
Mercury	µg/L	0.06	0	-	_	-	_	-	-	11	0.05	0.05	0.05	0.05	0.05	0.05		
Molybdenum	µg/L	11	0	-	_	_	_	_	-	11	0.5	0.5	0.5	0.5	0.5	0.5		
Nickel	µg/L	-	0	-	_	_	-	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Selenium	µg/L	5	0	-	_	_	_	_	_	11	1	1	1	1	1	1		
Silver	µg/L	0.05	0	-	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Strontium	µg/L	_	0	_	_	_	_	_	_	11	13	18	19	21	33	44		
Thorium	µg/L	_	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Tin	µg/L	_	0	-	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		
Uranium	µg/L	_	0	_	_	_	_	_	_	11	0.5	0.5	0.5	0.5	0.5	0.5		

			Surface						Беер							
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Vanadium	µg/L	_	0	_	_	_	_	_	_	11	5	5	5	5	5	5
Zinc	µg/L	8	0	_	_	_	_	_	_	11	2.5	2.5	2.5	2.5	6.25	10
Soluble Metals																_
Aluminium	µg/L	55	0	_	_	_	-	_	_	5	2.5	2.5	2.5	30.2	113.3	141
Arsenic	µg/L	13	0	_	_	_	-	_	_	0	_	-	-	_	_	_
Barium	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Beryllium	µg/L	_	0	_	_	_	-	_	_	0	_	-	-	_	_	_
Boron	µg/L	370	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Cadmium	µg/L	0.2	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Chromium	µg/L	1	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Cobalt	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Copper	µg/L	1.4	0	_	_	_	-	_	_	0	_	-	-	_	_	_
Iron	µg/L	_	3	59	192.6	393	691.2	840.3	890	67	60	225.4	450	842.8	1249	1840
Lead	µg/L	3.4	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Lithium	µg/L	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Manganese	µg/L	1900	5	2.1	2.9	5	44.4	48.6	50	67	0.5	7.52	25	75.56	233.4	582
Molybdenum	µg/L	11	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Nickel	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Silver	µg/L	0.05	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Strontium	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Thorium	µg/L	-	0	-	-	-	-	-	-	0	-	-	-	-	-	-
Tin	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Uranium	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Vanadium	µg/L	-	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Zinc	µg/L	8	0	-	-	_	-	_	_	0	_	_	_	_	-	-
Algae							_									
Chlorophyll a - Field	µg/L	5	1	8.2	8.2	8.2	8.2	8.2	8.2	0	-	-	-	-	-	-
Chlorophyll a	µg/L	5	134	0	6	9	12	18	36	0	_	_	_	_	_	_
Microbiological																
Cryptosporidium - Corrected	MPN/100mL	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Cryptosporidium - Detected	MPN/100mL	_	0	_	-	-	-	_	_	0	_	_	_	_	_	-
E.coli - Colilert	MPN/100mL	-	78	0.5	1	3	7.6	16.6	550	0	-	-	-	-	-	-
E.coli - MF	MPN/100mL	-	2	6	8	11	14	15.5	16	0	-	-	-	-	-	-
Enterococci - MF	MPN/100mL	_	80	0.5	0.5	1	3	10	320	0	_	_	_	_	_	_

			Surface						Беер								
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	
Giardia - Corrected	MPN/100mL	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_	
Giardia - Detected	MPN/100mL	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_	
Total Coliforms - Colilert	MPN/100mL	-	73	99	390	730	1360	1940	2400	0	_	-	-	-	-	-	
Pesticide																	
2.4-D	µg/L	_	0	-	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
2.4-DB	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
2.4-DP	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
2.4.5-T	µg/L	36	0	-	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
2.4.6-T	µg/L	_	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05	
2.6-D	µg/L	280	0	-	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05	
3-Hydroxy Carbofuran	µg/L	_	0	-	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
4-Chlorophenoxy acetic acid	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
4.4-DDD (pp-DDD)	µg/L	_	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
4.4-DDE (pp-DDE)	µg/L	_	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
4.4-DDT (pp-DDT)	µg/L	0.006	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Acrolein	µg/L	_	0	_	_	_	_	_	_	1	1	1	1	1	1	1	
Aldicarb	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Aldrin (Eldrin)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
alpha-BHC (lindane)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Ametryn	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Amitrole	µg/L	_	0	_	_	_	_	_	_	1	0.05	0.05	0.05	0.05	0.05	0.05	
AMPA	µg/L	_	0	_	_	_	_	_	_	2	0.5	0.5	0.5	0.5	0.5	0.5	
Asulam	µg/L	_	0	_	_	_	_	_	_	2	1	1	1	1	1	1	
Atrazine	µg/L	13	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Azinphos methyl	µg/L	0.01	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Azinphos-ethyl	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Bendiocarb	µg/L	_	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05	
Benomyl	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.006	0.0075	0.009	0.00975	0.01	
beta-BHC (lindane)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Bioresmethrin	µg/L	_	0	_	_	_	_	_	_	1	0.25	0.25	0.25	0.25	0.25	0.25	
Brodifacoum	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Bromacil	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Bromophos-ethyl	µg/L	_	0	_	_	_	_	_	_	2	0.05	0.05	0.05	0.05	0.05	0.05	
Bromoxynil	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	

			Surface														
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	
Carbaryl	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Carbofuran	µg/L	0.06	0	-	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Carbophenothion	µg/L	_	0	-	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Chlordane - Total	µg/L	0.03	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Chlorfenvinphos - Total	µg/L	_	0	-	_	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Chlorothalonil	µg/L	_	0	-	_	_	_	_	_	2	1	1	1	1	1	1	
Chlorpyrifos	µg/L	0.01	0	-	_	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Chlorpyrifos-methyl	µg/L	_	0	-	_	_	-	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1	
Chlorsulfuron	µg/L	_	0	-	_	_	_	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1	
cis-Chlordane	µg/L	_	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Clopyralid	µg/L	_	0	-	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Coumaphos	µg/L	_	0	-	-	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Cyanazine	µg/L	-	0	-	-	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Cyproconazole	µg/L	-	0	-	-	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Cyprodinil	µg/L	_	0	-	-	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Cyromazine	µg/L	-	0	-	-	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
delta-BHC	µg/L	_	0	-	-	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Demeton-O & Demeton-S	µg/L	_	0	-	-	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Demeton-S methyl	µg/L	_	0	-	-	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Diazinon	µg/L	0.01	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Dicamba	µg/L	_	0	-	-	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Dichlorvos (DDVP)	µg/L	_	0	_	_	_	_	_	_	2	0.1	0.1	0.1	0.1	0.1	0.1	
Diclofop-methyl	µg/L	_	0	-	-	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Dieldrin	µg/L	-	0	-	-	_	-	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Difenoconazole	µg/L	-	0	-	-	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Diflufenican	µg/L	_	0	-	-	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Dimethoate	µg/L	0.15	0	-	-	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Diquat	µg/L	1.4	0	_	_	_	_	_	_	1	0.025	0.025	0.025	0.025	0.025	0.025	
Disulfoton	µg/L	_	0	-	-	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Diuron	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Endosulfan - Total	µg/L	0.03	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Endosulfan I (alpha isomer)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Endosulfan II (beta isomer)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Endosulfan sulfate	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	

			Surface						Deen								
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	
Endrin	µg/L	0.01	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Endrin aldehyde	µg/L	_	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Endrin ketone	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
EPN	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Ethion	µg/L	_	0	-	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Ethoprophos (Prophos)	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Fenamiphos	µg/L	_	0	-	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Fenarimol	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Fenchlorphos (Ronnel)	µg/L	_	0	_	_	_	_	_	_	2	5	5	5	5	5	5	
Fenitrothion	µg/L	0.2	0	-	_	_	_	_	_	2	1	1	1	1	1	1	
Fensulphothion	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Fenthion	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Fenvalerate & Esfenvalerate	µg/L	_	0	_	_	_	_	_	_	1	0.25	0.25	0.25	0.25	0.25	0.25	
Fipronil	µg/L	_	0	-	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Fluometuron	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Fluroxypyr	µg/L	_	0	_	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Flusilazole	µg/L	_	0	-	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Glyphosate	µg/L	370	0	_	_	_	_	_	_	2	0.5	0.5	0.5	0.5	0.5	0.5	
Heptachlor	µg/L	0.01	0	_	_	_	_	_	_	2	0.0005	0.0005	0.0005	0.0005	0.0005	0.0005	
Heptachlor epoxide	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Hexachlorobenzene (HCB)	µg/L	_	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Hexazinone	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Iprodione	µg/L	_	0	-	_	_	_	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Irgarol	µg/L	_	0	-	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Lindane (gamma-BHC)	µg/L	0.2	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Malathion (Maldison)	µg/L	0.05	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
MCPA	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
МСРВ	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Mecoprop	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Methiocarb	µg/L	_	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Methomyl	µg/L	3.5	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Methoxychlor	µg/L	-	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Metolachlor	µg/L	-	0	_	_	_	_	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Metribuzin	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	

			Curford														
Parameter	Units	WOO	Surrace							Беер							
	Units	ngo	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	
Metsulfuron-methyl	µg/L	-	0	-	-	-	-	-	-	1	0.05	0.05	0.05	0.05	0.05	0.05	
Mevinphos	µg/L	-	0	-	-	-	-	-	-	2	0.01	0.01	0.01	0.01	0.01	0.01	
Molinate	µg/L	3.4	0	-	-	-	-	-	_	2	0.05	0.05	0.05	0.05	0.05	0.05	
Monocrotophos	µg/L	-	0	-	-	-	-	-	-	2	0.01	0.01	0.01	0.01	0.01	0.01	
Omethoate	µg/L	_	0	-	_	_	-	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Oryzalin	µg/L	-	0	-	-	-	-	-	-	2	0.025	0.025	0.025	0.025	0.025	0.025	
Oxamyl	µg/L	_	0	-	_	_	-	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Oxychlordane	µg/L	_	0	-	_	_	-	-	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Oxyfluorfen	µg/L	_	0	-	_	_	-	-	_	2	0.5	0.5	0.5	0.5	0.5	0.5	
Paclobutrazol	µg/L	_	0	-	_	_	-	-	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Paraquat	µg/L	-	0	-	-	-	-	-	-	1	0.05	0.05	0.05	0.05	0.05	0.05	
Parathion	µg/L	0.004	0	-	_	_	-	-	_	2	0.1	0.1	0.1	0.1	0.1	0.1	
Parathion-methyl	µg/L	_	0	-	-	-	-	-	_	2	1	1	1	1	1	1	
Penconazole	µg/L	_	0	-	-	-	_	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Pendimethalin	µg/L	_	0	-	_	_	-	-	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Permethrin	µg/L	-	0	-	-	-	-	-	-	1	0.25	0.25	0.25	0.25	0.25	0.25	
Phorate	µg/L	-	0	-	-	-	-	-	-	2	0.05	0.05	0.05	0.05	0.05	0.05	
Picloram	µg/L	_	0	-	_	_	-	-	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Piperonyl Butoxide	µg/L	_	0	-	_	_	-	-	_	1	0.25	0.25	0.25	0.25	0.25	0.25	
Pirimiphos-ethyl	µg/L	_	0	_	_	_	-	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Pirimiphos-methyl	µg/L	_	0	-	_	_	-	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Profenofos	µg/L	_	0	-	_	_	-	-	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Prometryn	µg/L	_	0	-	_	_	-	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Propazine	µg/L	-	0	-	-	-	-	-	-	2	0.005	0.005	0.005	0.005	0.005	0.005	
Propiconazole	µg/L	_	0	-	_	_	-	-	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Prothiophos (Tokuthion)	µg/L	_	0	-	_	_	-	-	_	2	0.05	0.05	0.05	0.05	0.05	0.05	
Pyrimethanil	µg/L	_	0	-	_	_	-	-	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Silvex (2,4,5-TP/Fenoprop)	µg/L	-	0	_	_	_	-	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Simazine	µg/L	3.2	0	-	_	_	-	-	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Sulprofos	µg/L	-	0	_	_	_	-	_	_	2	0.025	0.025	0.025	0.025	0.025	0.025	
Sum of DDD + DDE + DDT	µg/L	-	0	_	_	_	_	_	_	2	0.001	0.001	0.001	0.001	0.001	0.001	
Tebuconazole	µg/L	-	0	_	_	_	-	_	_	2	0.005	0.005	0.005	0.005	0.005	0.005	
Tebuthiuron	µg/L	2.2	0	_	_	_	-	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01	
Temephos	µg/L	-	0	-	_	_	_	-	-	2	0.01	0.01	0.01	0.01	0.01	0.01	

Surface Deep Parameter Units WQO 20th 80th 95th Count minimum 20th median 80th 95th maximum 20th 80th 95th percentile percentile percentile percentile maximum Count minimum 80th 95th	aximum															
Parameter Units WQO 20th 80th 95th 20th 80th 95th 20th 95th 80th 95th 20th 95th 80th 95th 80th 95th 80th 95th 95th 95th 95th 95th 95th 95th 95	aximum															
Terbufos µg/L – 0 – – – – – – – 2 0.005 0.	.005															
Terbuthylazine µg/L - 0 2 0.005 0.00	.005															
Terbutryn µg/L – 0 – – – – – – 2 0.005 0.0	.005															
Tetrachlorvinphos µg/L - 0 2 0.005 0	.005															
Tetraethyl dithiopyrophos µg/L – 0 – – – – – – – – 2 0.0025 0.002	.0025															
Thiamethoxam µg/L - 0 2 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Thiobencarb µg/L 2.8 0 2 0.005	.005															
Thiodicarb µg/L - 0 2 0.005 0.	.005															
trans-Chlordane µg/L - 0 2 0.001 0.001 0.001 0.001 0.001 0.001 0.001 0.001	.001															
Triazophos µg/L – 0 – – – – – – – 2 0.0025 0	0025															
Trichlorfon µg/L – 0 – – – – – – – 2 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Trichloronate (Trichloronat) µg/L – 0 – – – – – – – 2 0.25 0.25 0.25 0.25 0.25 0.25 0.25 0.	.25															
Triclopyr µg/L – 0 – – – – – – 2 0.005 0.005 0.005 0.005 0.005 0.005 0.005	.005															
Trifluralin µg/L 2.6 0 2 5 5 5 5 5 5 5 5																
Polyaromatic Hydrocarbon																
2-Chloronaphthalene µg/L - 0 3 0.05 0.05 0.05 0.05 0.05 0.05 0.05	.05															
2-Methylnapthalene µg/L - 0 3 0.05 0.05 0.05 0.05 0.05 0.05 0.05	.05															
Acenaphthene µg/L - 0 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Acenapthylene µg/L - 0 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Anthracene µg/L – 0 – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Benzo[a]anthracene µg/L – 0 – – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Benzo[a]pyrene µg/L – 0 – – – – – – – 3 0.0025 0.00	0025															
Benzo[b+j]fluoranthene µg/L – 0 – – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Benzo[e]pyrene µg/L - 0 3 0.05 0.05 0.05 0.05 0.05 0.05 0.05	.05															
Benzo[ghi]perylene µg/L – 0 – – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Benzo[k]fluoranthene µg/L – 0 – – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Chrysene µg/L - 0 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Dibenzo[ah]anthracene µg/L – 0 – – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Fluoranthene µg/L – 0 – – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Fluorene µg/L – 0 – – – – – – 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Indeno(1.2.3-cd)pyrene µg/L - 0 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Napthalene µg/L 16 0 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01	.01															
Perylene µg/L - 0 3 0.05 0.05 0.05 0.05 0.05 0.05 0.05	.05															
Phenanthrene µg/L - 0 3 0.01 0.01 0.01 0.01 0.01 0.01 0.01 0.	.01															
			Surface							Deep						
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Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Pyrene	µg/L	_	0	_	_	_	_	_	_	3	0.01	0.01	0.01	0.01	0.01	0.01
Total PAH	µg/L	_	0	_	_	_	_	_	_	3	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Taste and Colour																
2-Methylisoborneol	ng/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
2-methylisoborneol + Geosmin	ng/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Geosmin	ng/L	_	0	-	_	-	-	-	-	0	_	-	_	_	_	-
Volatile Organic Compound																
Benzene	µg/L	950	0	_	_	_	_	_	_	3	0.25	0.25	0.25	0.25	0.25	0.25
Toluene	µg/L	_	0	_	_	_	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Ethylbenzene	µg/L	_	0	_	_	_	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
meta & para-Xylene	µg/L	_	0	_	_	_	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
ortho-Xylene	µg/L	350	0	_	_	_	_	_	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Total Xylenes	µg/L	-	0	-	_	_	_	-	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Hexaconazole	µg/L	_	0	_	_	_	_	_	_	2	0.01	0.01	0.01	0.01	0.01	0.01

Grey shading denotes parameters that exceed the water quality object / trigger value

Gold shading denotes parameters that have an LOR higher than the water quality objective / trigger value

Six Mile Creek Arm

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Physico-Chemical																
Temperature - Field	°C	_	24	14.3	17.8584	21.295	25.008	26.565	27.3	0	_	_	_	_	_	_
pH - Field	pH unit	6.5-8.0	24	6.07	6.5	6.88	7.278	7.7575	8	0	_	_	_	_	_	_
pH - Lab	ph Unit	6.5-8.0	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Conductivity - Field	µS/cm	626	24	108	116.6	123	136.4	166.35	187	0	_	_	_	_	_	_
Conductivity	µS/cm	626	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Dissolved Oxygen - Field	mg/L	_	24	2.17	4.728	5.815	7.212	8.2905	8.87	0	_	_	_	_	_	_
Dissolved Oxygen - Field	% Saturation	85-110	24	0.209	51.16	67.4	80.46	97.91	102.1	0	_	_	_	_	_	_
Turbidity - Field	NTU	50	24	1.2	2.16	3.75	6.168	16.51	25.2	0	_	_	_	_	_	_
Turbidity - Lab	NTU	50	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Suspended Solids	mg/L	6	30	0.5	3	4	6	8	10	1	3	3	3	3	3	3
Total Dissolved Solids	mg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Alkalinity - Total	mg/L	-	0	_	_	_	_	_	_	0	_	_	_	_	_	_
BGA - Field	cells/mL	-	1	279400	279400	279400	279400	279400	279400	0	_	_	_	_	_	_
BGA - Field	RFU	-	1	132.8	132.8	132.8	132.8	132.8	132.8	0	_	_	_	_	_	_
Eutrophic Depth	mg/L	-	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Redox Potential	MV	_	5	68.7	78.86	97.8	127.14	127.56	127.7	0	_	_	_	-	_	_
SUVA	L/mg-M	_	2	3.8	3.9	4.05	4.2	4.275	4.3	0	_	_	_	-	_	_
Nutrients							_									
Total Nitrogen as N	mg/L	0.5	33	0.09	0.47	0.61	0.878	0.974	1.1	1	0.28	0.28	0.28	0.28	0.28	0.28
Total Dissolved Nitrogen	mg/L	-	27	0.07	0.412	0.5	0.646	0.741	1	1	0.26	0.26	0.26	0.26	0.26	0.26
Kjeldahl Nitrogen as N	mg/L	_	1	0.3	0.3	0.3	0.3	0.3	0.3	1	0.27	0.27	0.27	0.27	0.27	0.27
Total Oxidised N	mg/L	0.06	29	0.001	0.001	0.001	0.0054	0.0498	0.202	1	0.006	0.006	0.006	0.006	0.006	0.006
Nitrate as N	mg/L	0.7	24	0.001	0.001	0.001	0.0024	0.019	0.053	1	0.006	0.006	0.006	0.006	0.006	0.006
Nitrite as N	mg/L	-	25	0.001	0.001	0.001	0.0034	0.0148	0.196	1	0.001	0.001	0.001	0.001	0.001	0.001
Organic Nitrogen	mg/L	0.42	0	-	-	-	-	-	-	0	-	-	-	-	-	-
Ammonia - Total as N	mg/L	0.02	31	0.0025	0.0025	0.008	0.019	0.0725	0.105	1	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Total Phosphorus	mg/L	0.05	33	0.006	0.0208	0.031	0.052	0.0832	0.175	1	0.02	0.02	0.02	0.02	0.02	0.02
Total Dissolved Phosphorus	mg/L	-	27	0.0025	0.0072	0.011	0.0176	0.0321	0.067	1	0.01	0.01	0.01	0.01	0.01	0.01
Filterable Reactive Phosphorus	mg/L	0.02	31	0.0005	0.002	0.003	0.005	0.012	0.027	1	0.003	0.003	0.003	0.003	0.003	0.003
Phosphate	mg/L	_	0	_	_	_	_	-	_	0	_	_	_	_	_	_

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Other Organics																
DOC	mg/L	-	30	5.3	6.6	7.9	10	15.2	25	0	_	_	_	_	_	_
ТОС	mg/L	-	30	5.3	6.84	8.3	10.52	14.2	23	0	-	-	-	_	_	_
Major Cations & Anions																
Fluoride - Total	mg/L	-	0	_	_	_	_	_	_	0	_	_	_	_	-	_
Chloride	mg/L	-	0	_	_	_	_	-	_	0	_	_	_	_	_	_
Magnesium - Total	mg/L	-	0	_	_	_	_	-	_	0	_	_	_	_	_	_
Silica - Reactive as SiO3	mg/L	-	16	1.26	2.75	4.91	7.51	8.0525	9.26	0	-	-	-	_	_	_
Sulphide	mg/L	-	0	_	_	_	_	_	_	0	_	_	_	_	-	_
Sulphur-Total	mg/L	-	0	_	_	_	-	-	_	0	-	-	-	_	_	_
Total Metals																
Aluminium	µg/L	55	4	2.5	14.2	26.5	86.6	149.15	170	6	11	40	120.5	509	765.5	851
Antimony	µg/L	-	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Arsenic	µg/L	13	4	0.6	0.66	0.8	1.1	1.325	1.4	6	0.8	0.8	1.2	1.5	1.575	1.6
Barium	µg/L	-	4	2.3	2.72	5	10.6	14.65	16	6	6	9	14.15	17.9	22.7	24.3
Beryllium	µg/L	-	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Boron	µg/L	370	4	15	15	15	15	15	15	6	15	15	15	15	15	15
Cadmium	µg/L	0.2	4	0.05	0.05	0.05	0.05	0.05	0.05	6	0.05	0.05	0.05	0.05	0.05	0.05
Chromium	µg/L	1	4	0.5	0.5	0.5	0.7	0.925	1	6	0.5	0.5	0.75	1.2	1.35	1.4
Cobalt	µg/L	_	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Copper	µg/L	1.4	4	0.5	0.5	0.5	1.1	1.775	2	6	0.5	0.5	0.5	1	1.75	2
Iron	µg/L	-	28	460	632.4	1120	1700	1972.5	2600	21	577	1160	1500	3610	4700	5580
Lead	µg/L	3.4	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Lithium	µg/L	-	4	0.5	0.5	0.5	1.1	1.775	2	6	1	1	1.25	2	2	2
Manganese	µg/L	1900	28	10	28.16	55.5	84.54	125.1	160	21	22.3	36	95	256	581	620
Mercury	µg/L	0.06	4	0.05	0.05	0.05	0.05	0.05	0.05	6	0.05	0.05	0.05	0.05	0.05	0.05
Molybdenum	µg/L	11	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Nickel	µg/L	-	4	0.5	0.5	0.5	1.1	1.775	2	6	0.5	0.5	0.5	0.5	1.625	2
Selenium	µg/L	5	4	1	1	1	1	1	1	6	1	1	1	1	1	1
Silver	µg/L	0.05	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Strontium	µg/L	_	4	16	16.6	17.5	21.2	24.8	26	6	17	19	20	27	27	27
Thorium	µg/L	_	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Tin	µg/L	_	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5
Uranium	µg/L	_	4	0.5	0.5	0.5	0.5	0.5	0.5	6	0.5	0.5	0.5	0.5	0.5	0.5

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Vanadium	µg/L	_	4	5	5	5	5	5	5	6	5	5	5	5	5	5
Zinc	µg/L	8	4	2.5	2.5	4.75	7.4	7.85	8	6	2.5	2.5	2.5	7	10	11
Soluble Metals																
Aluminium	µg/L	55	1	2.5	2.5	2.5	2.5	2.5	2.5	1	8	8	8	8	8	8
Arsenic	µg/L	13	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Barium	µg/L	-	0	-	-	_	-	_	_	0	-	_	_	-	_	-
Beryllium	µg/L	_	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Boron	µg/L	370	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Cadmium	µg/L	0.2	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Chromium	µg/L	1	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Cobalt	µg/L	_	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Copper	µg/L	1.4	0	_	_	-	_	_	_	0	_	-	_	_	_	_
Iron	µg/L	_	29	162	306.8	633	993	1276	1320	22	239	430	1120	1850	3877.5	4100
Lead	µg/L	3.4	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Lithium	µg/L	_	0	_	_	-	_	_	_	0	_	-	_	_	_	_
Manganese	µg/L	1900	29	1.3	4	8.5	14.88	26.8	130	22	4.2	21.46	37	247.2	535.7	550
Molybdenum	µg/L	11	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Nickel	µg/L	_	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Silver	µg/L	0.05	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Strontium	µg/L	_	0	_	_	-	_	_	_	0	_	-	_	_	_	_
Thorium	µg/L	_	0	-	_	-	_	_	_	0	_	-	_	_	_	_
Tin	µg/L	_	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Uranium	µg/L	_	0	_	_	-	_	_	_	0	_	-	_	_	_	_
Vanadium	µg/L	_	0	_	_	-	_	_	_	0	_	_	_	_	_	_
Zinc	µg/L	8	0	_	_	-	_	_	_	0	_	-	_	_	_	_
Algae							_									
Chlorophyll a - Field	µg/L	5	1	5.7	5.7	5.7	5.7	5.7	5.7	0	_	-	_	_	_	_
Chlorophyll a	µg/L	5	7	4	7.2	11	18.2	22.1	23	0	_	_	_	_	_	_
Microbiological																
Cryptosporidium - Corrected	MPN/100mL	_	0	_	_	-	_	_	_	0	_	-	_	_	_	_
Cryptosporidium - Detected	MPN/100mL	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
E.coli - Colilert	MPN/100mL	_	70	1	16	32	96.8	300.5	520	0	_	_	_	_	_	_
E.coli - MF	MPN/100mL	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Enterococci - MF	MPN/100mL	_	70	0.5	2.8	11	24	130	340	0	_	_	_	_	_	

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Giardia - Corrected	MPN/100mL	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Giardia - Detected	MPN/100mL	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Total Coliforms - Colilert	MPN/100mL	_	70	280	1600	2400	2400	4565	12000	0	_	-	_	-	_	_
Pesticide																
2.4-D	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
2.4-DB	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
2.4-DP	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
2.4.5-T	µg/L	36	0	-	-	_	_	_	_	0	_	-	_	-	_	_
2.4.6-T	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
2.6-D	µg/L	280	0	_	-	_	_	_	_	0	-	_	_	_	_	_
3-Hydroxy Carbofuran	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
4-Chlorophenoxy acetic acid	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
4.4-DDD (pp-DDD)	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
4.4-DDE (pp-DDE)	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
4.4-DDT (pp-DDT)	µg/L	0.006	0	-	-	_	_	_	_	0	_	-	_	-	_	_
Acrolein	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Aldicarb	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Aldrin (Eldrin)	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
alpha-BHC (lindane)	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Ametryn	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
Amitrole	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
AMPA	µg/L	_	0	_	-	_	_	_	-	0	-	_	_	_	_	_
Asulam	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Atrazine	µg/L	13	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Azinphos methyl	µg/L	0.01	0	-	-	_	_	_	_	0	_	-	_	-	_	_
Azinphos-ethyl	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
Bendiocarb	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Benomyl	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
beta-BHC (lindane)	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Bioresmethrin	µg/L	_	0	-	-	_	_	_	_	0	_	-	_	-	_	_
Brodifacoum	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Bromacil	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Bromophos-ethyl	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Bromoxynil	µg/L	-	0	-	_	-	_	_	_	0	_	-	_	_	_	

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Carbaryl	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	-
Carbofuran	µg/L	0.06	0	-	_	_	_	_	_	0	-	_	-	-	_	_
Carbophenothion	µg/L	_	0	-	_	_	_	_	_	0	-	_	_	_	_	_
Chlordane - Total	µg/L	0.03	0	-	_	_	_	_	_	0	-	_	-	-	_	_
Chlorfenvinphos - Total	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Chlorothalonil	µg/L	_	0	-	_	_	_	_	_	0	-	_	_	_	_	_
Chlorpyrifos	µg/L	0.01	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Chlorpyrifos-methyl	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Chlorsulfuron	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
cis-Chlordane	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Clopyralid	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Coumaphos	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Cyanazine	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Cyproconazole	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Cyprodinil	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Cyromazine	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
delta-BHC	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Demeton-O & Demeton-S	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Demeton-S methyl	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Diazinon	µg/L	0.01	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Dicamba	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Dichlorvos (DDVP)	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Diclofop-methyl	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Dieldrin	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Difenoconazole	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Diflufenican	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Dimethoate	µg/L	0.15	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Diquat	µg/L	1.4	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Disulfoton	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Diuron	µg/L	_	0	-	_	-	-	_	_	0	_	-	-	-	_	_
Endosulfan - Total	µg/L	0.03	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Endosulfan I (alpha isomer)	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Endosulfan II (beta isomer)	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Endosulfan sulfate	µg/L	_	0	-	_	_	_	_	_	0	_	-	_	-	-	-

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Endrin	µg/L	0.01	0	_	_	_	_	_	_	0	_	_	_	_	-	_
Endrin aldehyde	µg/L	_	0	-	_	_	_	_	_	0	_	_	_	_	_	_
Endrin ketone	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
EPN	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Ethion	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Ethoprophos (Prophos)	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Fenamiphos	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Fenarimol	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Fenchlorphos (Ronnel)	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Fenitrothion	µg/L	0.2	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Fensulphothion	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Fenthion	µg/L	_	0	_	-	_	-	_	_	0	_	_	_	-	-	_
Fenvalerate & Esfenvalerate	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Fipronil	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Fluometuron	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Fluroxypyr	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Flusilazole	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Glyphosate	µg/L	370	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Heptachlor	µg/L	0.01	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Heptachlor epoxide	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Hexachlorobenzene (HCB)	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Hexazinone	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Iprodione	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Irgarol	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Lindane (gamma-BHC)	µg/L	0.2	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Malathion (Maldison)	µg/L	0.05	0	_	_	_	_	_	_	0	_	-	_	_	_	_
MCPA	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
МСРВ	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Mecoprop	µg/L	_	0	_	_	_	_	_	_	0	_	-	_	_	_	_
Methiocarb	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	_	_	_
Methomyl	µg/L	3.5	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Methoxychlor	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Metolachlor	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Metribuzin	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Metsulfuron-methyl	µg/L	_	0	_	_	_	_	_	-	0	_	_	_	_	-	_
Mevinphos	µg/L	_	0	-	_	_	-	_	-	0	-	-	_	-	-	_
Molinate	µg/L	3.4	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Monocrotophos	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Omethoate	µg/L	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Oryzalin	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Oxamyl	µg/L	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Oxychlordane	µg/L	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Oxyfluorfen	µg/L	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Paclobutrazol	µg/L	_	0	-	_	_	-	_	-	0	-	-	_	-	-	_
Paraquat	µg/L	_	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Parathion	µg/L	0.004	0	_	_	_	-	_	_	0	_	_	_	_	_	_
Parathion-methyl	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Penconazole	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Pendimethalin	µg/L	_	0	-	_	_	-	_	-	0	-	-	_	-	-	_
Permethrin	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Phorate	µg/L	_	0	-	_	_	-	_	-	0	-	-	_	-	-	_
Picloram	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Piperonyl Butoxide	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Pirimiphos-ethyl	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Pirimiphos-methyl	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Profenofos	µg/L	-	0	-	-	-	-	-	-	0	-	-	-	-	-	-
Prometryn	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Propazine	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Propiconazole	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Prothiophos (Tokuthion)	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Pyrimethanil	µg/L	-	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Silvex (2,4,5-TP/Fenoprop)	µg/L	-	0	-	_	-	-	-	-	0	-	-	_	-	-	-
Simazine	µg/L	3.2	0	-	_	-	-	-	-	0	-	-	-	-	-	-
Sulprofos	µg/L	-	0	_	_	-	-	_	_	0	_	-	-	_	-	_
Sum of DDD + DDE + DDT	µg/L	-	0	_	_	-	-	_	_	0	_	-	-	_	-	_
Tebuconazole	µg/L	-	0	_	_	-	-	_	_	0	_	-	-	_	-	_
Tebuthiuron	µg/L	2.2	0	_	_	-	-	_	_	0	_	-	-	_	-	_
Temephos	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_

			Surface							Doon						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Terbufos	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Terbuthylazine	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Terbutryn	µg/L	_	0	_	_	_	_	_	-	0	_	-	_	-	_	_
Tetrachlorvinphos	µg/L	_	0	_	_	_	_	_	-	0	_	-	_	-	_	_
Tetraethyl dithiopyrophos	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Thiamethoxam	µg/L	_	0	_	-	_	_	_	_	0	_	_	_	-	_	_
Thiobencarb	µg/L	2.8	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Thiodicarb	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
trans-Chlordane	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Triazophos	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Trichlorfon	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Trichloronate (Trichloronat)	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Triclopyr	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Trifluralin	µg/L	2.6	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Polyaromatic Hydrocarbon																
2-Chloronaphthalene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
2-Methylnapthalene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Acenaphthene	µg/L	_	0	-	_	_	-	-	_	0	-	_	_	-	-	-
Acenapthylene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Anthracene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Benzo[a]anthracene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Benzo[a]pyrene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Benzo[b+j]fluoranthene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Benzo[e]pyrene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Benzo[ghi]perylene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Benzo[k]fluoranthene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Chrysene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Dibenzo[ah]anthracene	µg/L	_	0	-	_	_	-	_	_	0	-	_	_	_	_	-
Fluoranthene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Fluorene	µg/L	-	0	-	_	-	-	-	_	0	-	_	-	-	-	-
Indeno(1.2.3-cd)pyrene	µg/L	_	0	-	_	_	-	_	_	0	-	_	_	_	_	-
Napthalene	µg/L	16	0	-	_	-	-	-	_	0	-	_	-	-	-	-
Perylene	µg/L	-	0	-	_	-	-	-	_	0	-	_	-	-	-	-
Phenanthrene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_

			Surface							Deep						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Pyrene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	_
Total PAH	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	-	-
Taste and Colour																
2-Methylisoborneol	ng/L	_	0	_	_	_	_	_	_	0	_	_	_	_	-	-
2-methylisoborneol + Geosmin	ng/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Geosmin	ng/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Volatile Organic Compound																
Benzene	µg/L	950	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Toluene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Ethylbenzene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
meta & para-Xylene	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	_	-
ortho-Xylene	µg/L	350	0	_	_	_	_	_	_	0	_	_	_	_	_	-
Total Xylenes	µg/L	_	0	_	_	_	_	_	_	0	_	_	_	_	-	_
Hexaconazole	µg/L	_	0	_	-	_	-	-	-	0	-	-	_	_	_	-

Grey shading denotes parameters that exceed the water quality object / trigger value

Gold shading denotes parameters that have an LOR higher than the water quality objective / trigger value

Cooroy Creek at Cooroy-Noosa Road

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Physico-Chemical									
Temperature - Field	°C	_	22	14.8	17.43	21.82	25.35	28.17	29.8
pH - Field	pH unit	6.5-8.0	22	6.2	6.59	6.7	6.9	7.02	7.1
pH - Lab	ph Unit	6.5-8.0	0	_	_	_	-	-	-
Conductivity - Field	µS/cm	626	22	80	106.13	127	139.6	147.65	197
Conductivity	µS/cm	626	0	_	_	_	-	-	-
Dissolved Oxygen - Field	mg/L	_	22	3.94	5.66	7.12	8.27	9.06	9.42
Dissolved Oxygen - Field	% Saturation	85-110	22	44.4	64.8	82.49	94.88	96.30	98.2
Turbidity - Field	NTU	50	22	2.1	3.04	3.915	6.63	19.51	114.1
Turbidity - Lab	NTU	50	0	_	-	_	_	-	_
Suspended Solids	mg/L	6	69	0.5	3	4	6	8.6	20
Total Dissolved Solids	mg/L	_	0	_	_	_	-	-	-
Alkalinity - Total	mg/L	_	0	_	-	_	_	-	_
BGA - Field	cells/mL	_	0	_	_	_	-	-	_
BGA - Field	RFU	_	0	_	_	_	-	-	_
Eutrophic Depth	mg/L	_	0	_	_	_	-	-	_
Redox Potential	MV	_	3	68.1	72.8	79.8	89.1	93.8	95.3
SUVA	L/mg-M	_	0	_	_	_	_	_	_

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Nutrients							_		
Total Nitrogen as N	mg/L	0.5	78	0.18	0.414	0.51	0.616	0.84	1.08
Total Dissolved Nitrogen	mg/L	_	64	0.15	0.366	0.44	0.52	0.67	0.91
Kjeldahl Nitrogen as N	mg/L	_	0	-	-	-	-	-	-
Total Oxidised N	mg/L	0.06	70	0.001	0.0058	0.0205	0.0558	0.1422	0.21
Nitrate as N	mg/L	0.7	39	0.001	0.0056	0.035	0.0882	0.1439	0.2
Nitrite as N	mg/L	_	40	0.001	0.001	0.003	0.005	0.0081	0.017
Organic Nitrogen	mg/L	0.42	0	_	_	_	-	_	_
Ammonia - Total as N	mg/L	0.02	67	0.0025	0.0232	0.044	0.083	0.15	0.316
Total Phosphorus	mg/L	0.05	78	0.0025	0.018	0.024	0.0316	0.04175	0.075
Total Dissolved Phosphorus	mg/L	_	64	0.0025	0.008	0.012	0.018	0.024	0.034
Filterable Reactive Phosphorus	mg/L	0.02	66	0.0005	0.0005	0.003	0.006	0.007	0.01
Phosphate	mg/L	_	0	_	_	_	_	_	_
Other Organics									
DOC	mg/L	_	67	3.1	5.4	6	7.58	9.49	11
TOC	mg/L	_	67	3.8	5.7	6.2	7.78	9.56	11.2
Major Cations & Anions									
Fluoride - Total	mg/L	_	0	_	_	_	_	_	_
Chloride	mg/L	_	0	_	_	_	_	_	-
Magnesium - Total	mg/L	_	2	1	1.2	1.5	1.8	1.95	2

	l Inits M		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Silica - Reactive as SiO3	mg/L	-	0	-	-	_	_	_	-
Sulphide	mg/L	_	41	0.013	0.025	0.025	0.025	0.025	0.06
Sulphur-Total	mg/L	_	0	_	_	_	_	_	_
Total Metals									_
Aluminium	µg/L	55	2	503	516.8	537.5	558.2	568.55	572
Antimony	µg/L	-	2	0.5	0.5	0.5	0.5	0.5	0.5
Arsenic	µg/L	13	2	0.6	0.64	0.7	0.76	0.79	0.8
Barium	µg/L	_	2	9	9.8	11	12.2	12.8	13
Beryllium	µg/L	-	2	0.5	0.5	0.5	0.5	0.5	0.5
Boron	µg/L	370	2	15	15	15	15	15	15
Cadmium	µg/L	0.2	2	0.05	0.05	0.05	0.05	0.05	0.05
Chromium	µg/L	1	2	0.5	0.6	0.75	0.9	0.975	1
Cobalt	µg/L	-	2	0.5	0.5	0.5	0.5	0.5	0.5
Copper	µg/L	1.4	2	0.5	0.6	0.75	0.9	0.975	1
Iron	µg/L	-	66	370	734	1050	1690	2330	2800
Lead	µg/L	3.4	2	0.5	0.5	0.5	0.5	0.5	0.5
Lithium	µg/L	-	2	0.5	0.8	1.25	1.7	1.925	2
Manganese	µg/L	1900	66	41	92	154.5	211	362.5	699
Mercury	µg/L	0.06	2	0.05	0.05	0.05	0.05	0.05	0.05
Molybdenum	µg/L	11	2	0.5	0.5	0.5	0.5	0.5	0.5

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Nickel	µg/L	-	2	0.5	0.5	0.5	0.5	0.5	0.5
Selenium	µg/L	5	2	1	1	1	1	1	1
Silver	µg/L	0.05	2	0.5	0.5	0.5	0.5	0.5	0.5
Strontium	µg/L	_	2	16	18.4	22	25.6	27.4	28
Thorium	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Tin	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Uranium	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Vanadium	µg/L	_	2	5	5	5	5	5	5
Zinc	µg/L	8	2	2.5	6	11.25	16.5	19.125	20
Soluble Metals									_
Aluminium	µg/L	55	2	87	88.4	90.5	92.6	93.65	94
Arsenic	µg/L	13	0	_	_	-	_	-	_
Barium	µg/L	_	0	_	_	-	_	-	-
Beryllium	µg/L	_	0	-	-	-	-	-	_
Boron	µg/L	370	0	-	-	-	-	-	_
Cadmium	µg/L	0.2	0	-	-	-	-	-	_
Chromium	µg/L	1	0	-	-	-	-	-	_
Cobalt	µg/L	_	0	_	_	-	-	-	-
Copper	µg/L	1.4	0	_	_	-	-	-	_
Iron	µg/L	_	67	70	277.2	434	834.4	1162	1560

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Lead	µg/L	3.4	0	_	-	-	-	-	-
Lithium	µg/L	-	0	-	-	_	_	_	_
Manganese	µg/L	1900	67	11.3	38.64	100	178.4	305	591
Molybdenum	µg/L	11	0	-	-	-	-	-	-
Nickel	µg/L	_	0	-	-	-	-	-	-
Silver	µg/L	0.05	0	-	-	-	-	-	-
Strontium	µg/L	_	0	_	-	-	-	-	-
Thorium	µg/L	_	0	_	_	_	_	_	_
Tin	µg/L	_	0	-	-	-	-	-	-
Uranium	µg/L	_	0	-	-	-	-	-	-
Vanadium	µg/L	_	0	-	-	-	-	-	-
Zinc	µg/L	8	0	_	_	_	_	_	_
Algae									
Chlorophyll a - Field	µg/L	5	0	-	-	-	-	-	-
Chlorophyll a	µg/L	5	0	_	_	_	_	_	_
Microbiological									
Cryptosporidium - Corrected	MPN/100mL	_	0	_	_	_	_	_	_
Cryptosporidium - Detected	MPN/100mL	_	0	_	_	_	_	_	_
E.coli - Colilert	MPN/100mL	_	67	0.5	4.2	12	56.6	233	2400
E.coli - MF	MPN/100mL	_	0	-	_	_	_	_	-

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Enterococci - MF	MPN/100mL	-	67	0.5	5.2	17	49.6	297	2100
Giardia - Corrected	MPN/100mL	_	0	_	-	-	-	_	_
Giardia - Detected	MPN/100mL	_	0	_	_	_	_	_	_
Total Coliforms - Colilert	MPN/100mL	_	67	680	2400	2400	2400	9070	14000
Pesticide									
2.4-D	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
2.4-DB	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
2.4-DP	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
2.4.5-T	µg/L	36	1	0.005	0.005	0.005	0.005	0.005	0.005
2.4.6-T	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
2.6-D	µg/L	280	1	0.05	0.05	0.05	0.05	0.05	0.05
3-Hydroxy Carbofuran	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
4-Chlorophenoxy acetic acid	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
4.4-DDD (pp-DDD)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
4.4-DDE (pp-DDE)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
4.4-DDT (pp-DDT)	µg/L	0.006	1	0.001	0.001	0.001	0.001	0.001	0.001
Acrolein	µg/L	_	0	_	_	_	_	_	_
Aldicarb	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Aldrin (Eldrin)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
alpha-BHC (lindane)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001

	l Inite V		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Ametryn	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Amitrole	µg/L	_	0	_	_	_	_	_	_
AMPA	µg/L	_	1	0.5	0.5	0.5	0.5	0.5	0.5
Asulam	µg/L	_	1	1	1	1	1	1	1
Atrazine	µg/L	13	1	0.005	0.005	0.005	0.005	0.005	0.005
Azinphos methyl	µg/L	0.01	1	0.01	0.01	0.01	0.01	0.01	0.01
Azinphos-ethyl	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Bendiocarb	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Benomyl	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
beta-BHC (lindane)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Bioresmethrin	µg/L	_	0	_	_	_	_	_	_
Brodifacoum	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Bromacil	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Bromophos-ethyl	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Bromoxynil	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Carbaryl	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Carbofuran	µg/L	0.06	1	0.005	0.005	0.005	0.005	0.005	0.005
Carbophenothion	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Chlordane - Total	µg/L	0.03	1	0.001	0.001	0.001	0.001	0.001	0.001
Chlorfenvinphos - Total	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Chlorothalonil	µg/L	-	1	1	1	1	1	1	1
Chlorpyrifos	µg/L	0.01	1	0.01	0.01	0.01	0.01	0.01	0.01
Chlorpyrifos-methyl	µg/L	_	1	0.1	0.1	0.1	0.1	0.1	0.1
Chlorsulfuron	µg/L	-	1	0.1	0.1	0.1	0.1	0.1	0.1
cis-Chlordane	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Clopyralid	µg/L	-	1	0.025	0.025	0.025	0.025	0.025	0.025
Coumaphos	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Cyanazine	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Cyproconazole	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Cyprodinil	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Cyromazine	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
delta-BHC	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Demeton-O & Demeton-S	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Demeton-S methyl	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Diazinon	µg/L	0.01	1	0.005	0.005	0.005	0.005	0.005	0.005
Dicamba	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Dichlorvos (DDVP)	µg/L	_	1	0.1	0.1	0.1	0.1	0.1	0.1
Diclofop-methyl	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Dieldrin	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Difenoconazole	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Diflufenican	µg/L	-	1	0.01	0.01	0.01	0.01	0.01	0.01
Dimethoate	µg/L	0.15	1	0.01	0.01	0.01	0.01	0.01	0.01
Diquat	µg/L	1.4	0	-	_	_	-	-	-
Disulfoton	µg/L	-	1	0.025	0.025	0.025	0.025	0.025	0.025
Diuron	µg/L	-	1	0.01	0.01	0.01	0.01	0.01	0.01
Endosulfan - Total	µg/L	0.03	1	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan I (alpha isomer)	µg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan II (beta isomer)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan sulfate	µg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001
Endrin	µg/L	0.01	1	0.001	0.001	0.001	0.001	0.001	0.001
Endrin aldehyde	µg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001
Endrin ketone	µg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001
EPN	µg/L	-	1	0.025	0.025	0.025	0.025	0.025	0.025
Ethion	µg/L	-	1	0.01	0.01	0.01	0.01	0.01	0.01
Ethoprophos (Prophos)	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Fenamiphos	µg/L	-	1	0.005	0.005	0.005	0.005	0.005	0.005
Fenarimol	µg/L	-	1	0.01	0.01	0.01	0.01	0.01	0.01
Fenchlorphos (Ronnel)	µg/L	_	1	5	5	5	5	5	5
Fenitrothion	µg/L	0.2	1	1	1	1	1	1	1
Fensulphothion	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005

	Units		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Fenthion	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Fenvalerate & Esfenvalerate	µg/L	_	0	-	_	_	_	_	_
Fipronil	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Fluometuron	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Fluroxypyr	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Flusilazole	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Glyphosate	µg/L	370	1	0.5	0.5	0.5	0.5	0.5	0.5
Heptachlor	µg/L	0.01	1	0.0005	0.0005	0.0005	0.0005	0.0005	0.0005
Heptachlor epoxide	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Hexachlorobenzene (HCB)	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Hexazinone	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Iprodione	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Irgarol	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Lindane (gamma-BHC)	µg/L	0.2	1	0.001	0.001	0.001	0.001	0.001	0.001
Malathion (Maldison)	µg/L	0.05	1	0.01	0.01	0.01	0.01	0.01	0.01
MCPA	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
МСРВ	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Месоргор	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Methiocarb	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Methomyl	µg/L	3.5	1	0.005	0.005	0.005	0.005	0.005	0.005

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Methoxychlor	µg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001
Metolachlor	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Metribuzin	µg/L	-	1	0.01	0.01	0.01	0.01	0.01	0.01
Metsulfuron-methyl	µg/L	-	0	_	_	_	_	_	_
Mevinphos	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Molinate	µg/L	3.4	1	0.05	0.05	0.05	0.05	0.05	0.05
Monocrotophos	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Omethoate	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Oryzalin	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Oxamyl	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Oxychlordane	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Oxyfluorfen	µg/L	_	1	0.5	0.5	0.5	0.5	0.5	0.5
Paclobutrazol	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Paraquat	µg/L	-	0	_	_	_	-	_	-
Parathion	µg/L	0.004	1	0.1	0.1	0.1	0.1	0.1	0.1
Parathion-methyl	µg/L	_	1	1	1	1	1	1	1
Penconazole	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Pendimethalin	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Permethrin	µg/L	_	0	_	_	_	_	_	_
Phorate	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05

	Units		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Picloram	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Piperonyl Butoxide	µg/L	-	0	_	_	_	_	-	_
Pirimiphos-ethyl	µg/L	-	1	0.005	0.005	0.005	0.005	0.005	0.005
Pirimiphos-methyl	µg/L	-	1	0.005	0.005	0.005	0.005	0.005	0.005
Profenofos	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Prometryn	µg/L	-	1	0.005	0.005	0.005	0.005	0.005	0.005
Propazine	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Propiconazole	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Prothiophos (Tokuthion)	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Pyrimethanil	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Silvex (2,4,5-TP/Fenoprop)	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Simazine	µg/L	3.2	1	0.01	0.01	0.01	0.01	0.01	0.01
Sulprofos	µg/L	_	1	0.025	0.025	0.025	0.025	0.025	0.025
Sum of DDD + DDE + DDT	µg/L	_	1	0.001	0.001	0.001	0.001	0.001	0.001
Tebuconazole	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Tebuthiuron	µg/L	2.2	1	0.01	0.01	0.01	0.01	0.01	0.01
Temephos	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Terbufos	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Terbuthylazine	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Terbutryn	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Tetrachlorvinphos	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Tetraethyl dithiopyrophos	µg/L	_	1	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Thiamethoxam	µg/L	-	1	0.01	0.01	0.01	0.01	0.01	0.01
Thiobencarb	µg/L	2.8	1	0.005	0.005	0.005	0.005	0.005	0.005
Thiodicarb	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
trans-Chlordane	µg/L	-	1	0.001	0.001	0.001	0.001	0.001	0.001
Triazophos	µg/L	_	1	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Trichlorfon	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01
Trichloronate (Trichloronat)	µg/L	-	1	0.25	0.25	0.25	0.25	0.25	0.25
Triclopyr	µg/L	_	1	0.005	0.005	0.005	0.005	0.005	0.005
Trifluralin	µg/L	2.6	1	5	5	5	5	5	5
Polyaromatic Hydrocarbon									
2-Chloronaphthalene	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
2-Methylnapthalene	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Acenaphthene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Acenapthylene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Anthracene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[a]anthracene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[a]pyrene	µg/L	_	2	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Benzo[b+j]fluoranthene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01

	llnite W		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Benzo[e]pyrene	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Benzo[ghi]perylene	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[k]fluoranthene	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Chrysene	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Dibenzo[ah]anthracene	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Fluoranthene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Fluorene	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Indeno(1.2.3-cd)pyrene	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Napthalene	µg/L	16	2	0.01	0.01	0.01	0.01	0.01	0.01
Perylene	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Phenanthrene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Pyrene	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Total PAH	µg/L	_	2	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Taste and Colour									
2-Methylisoborneol	ng/L	_	0	-	-	-	-	-	-
2-methylisoborneol + Geosmin	ng/L	_	0	-	-	-	-	-	-
Geosmin	ng/L	_	0	-	-	-	-	-	-
Volatile Organic Compound									
Benzene	µg/L	950	2	0.25	0.25	0.25	0.25	0.25	0.25
Toluene	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5

	Unite	WQO	Surface								
Parameter	Units		Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum		
Ethylbenzene	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5		
meta & para-Xylene	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5		
ortho-Xylene	µg/L	350	2	0.5	0.5	0.5	0.5	0.5	0.5		
Total Xylenes	µg/L	-	2	0.5	0.5	0.5	0.5	0.5	0.5		
Hexaconazole	µg/L	_	1	0.01	0.01	0.01	0.01	0.01	0.01		

Grey shading denotes parameters that exceed the water quality object / trigger value

Gold shading denotes parameters that have an LOR higher than the water quality objective / trigger value

Lake Macdonald Tailwater Below Dam Wall

D			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Physico-Chemical									
Temperature - Field	°C	_	23	11.69	13.744	16.2	20.64	24.91	25.9
pH - Field	pH unit	6.5-8.0	25	5.1	6	6.4	6.64	7.02	7.5
pH - Lab	ph Unit	6.5-8.0	114	5.7	6.3	6.5	6.7	7	7.8
Conductivity - Field	µS/cm	626	21	61	79	90	96	99	100.4
Conductivity	µS/cm	626	118	51	112.4	131.5	147.2	174.2	220
Dissolved Oxygen - Field	mg/L	_	21	0.24	0.69	1.16	2.4	3.6	3.74
Dissolved Oxygen - Field	% Saturation	85-110	20	2.5	6.9	11.78	21.74	34.48	36.03
Turbidity - Field	NTU	50	19	7.07	11.20	17.9	30.31	60.50	70.43
Turbidity - Lab	NTU	50	7	7.4	9.52	12	32.28	96.08	122
Suspended Solids	mg/L	6	185	2	4	7	13	23	65
Total Dissolved Solids	mg/L	_	11	44	56	73	85	102.5	120
Alkalinity - Total	mg/L	_	0	-	_	_	_	_	_
BGA - Field	cells/mL	_	0	-	_	_	_	_	_
BGA - Field	RFU	_	0	-	_	_	_	_	_
Eutrophic Depth	mg/L	_	0	-	_	_	_	_	_
Redox Potential	MV	_	6	62.1	63.7	76	141	141.05625	141.075
SUVA	L/mg-M	_	2	3.9	4.04	4.25	4.46	4.565	4.6

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Nutrients							_		
Total Nitrogen as N	mg/L	0.5	189	0.09	0.366	0.55	0.974	1.46	4.5
Total Dissolved Nitrogen	mg/L	_	57	0.04	0.342	0.45	0.646	0.812	0.95
Kjeldahl Nitrogen as N	mg/L	_	69	0	0.396	0.6	1.04	1.72	4.4
Total Oxidised N	mg/L	0.06	130	0	0.001	0.01	0.0892	0.1865	0.464
Nitrate as N	mg/L	0.7	36	0.001	0.001	0.001	0.005	0.00625	0.077
Nitrite as N	mg/L	_	37	0.001	0.001	0.003	0.0078	0.009	0.02
Organic Nitrogen	mg/L	0.42	12	0.47	0.58	0.705	1.008	1.5495	2.16
Ammonia - Total as N	mg/L	0.02	77	0.0025	0.016	0.035	0.0934	0.1746	0.487
Total Phosphorus	mg/L	0.05	192	0	0.0222	0.0405	0.07	0.1245	0.89
Total Dissolved Phosphorus	mg/L	_	58	0.0025	0.016	0.022	0.029	0.037	0.043
Filterable Reactive Phosphorus	mg/L	0.02	76	0	0.0005	0.002	0.008	0.0155	0.03
Phosphate	mg/L	_	1	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Other Organics									
DOC	mg/L	_	65	4.7	6.96	10.8	14	21	24
TOC	mg/L	_	65	4.7	7.38	11.6	15.32	23.4	25
Major Cations & Anions									
Fluoride - Total	mg/L	_	0	_	_	_	-	_	_
Chloride	mg/L	_	0	_	_	_	_	_	-
Magnesium - Total	mg/L	_	3	0.5	0.7	1	1.6	1.9	2

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Silica - Reactive as SiO3	mg/L	_	0	_	_	_	_	_	-
Sulphide	mg/L	_	0	_	_	-	_	_	_
Sulphur-Total	mg/L	_	0	_	_	-	_	_	_
Total Metals									_
Aluminium	µg/L	55	3	84	105.2	137	1128.8	1624.7	1790
Antimony	µg/L	-	3	0.5	0.5	0.5	0.5	0.5	0.5
Arsenic	µg/L	13	3	0.8	0.8	0.8	0.92	0.98	1
Barium	µg/L	_	3	7	7.8	9	9.6	9.9	10
Beryllium	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Boron	µg/L	370	3	15	15	15	15	15	15
Cadmium	µg/L	0.2	3	0.05	0.05	0.05	0.05	0.05	0.05
Chromium	µg/L	1	3	0.5	0.5	0.5	1.4	1.85	2
Cobalt	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Copper	µg/L	1.4	3	0.5	1.1	2	2	2	2
Iron	µg/L	-	62	1490	2292	3150	4200	5591.5	7640
Lead	µg/L	3.4	4	0.5	0.5	0.5	0.7	0.925	1
Lithium	µg/L	-	3	0.5	0.5	0.5	0.5	0.5	0.5
Manganese	µg/L	1900	62	49.8	132.2	229.5	330.8	469.85	1300
Mercury	µg/L	0.06	3	0.05	0.05	0.05	0.32	0.455	0.5
Molybdenum	µg/L	11	3	0.5	0.5	0.5	0.5	0.5	0.5

			Surface							
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum	
Nickel	µg/L	-	3	0.5	0.5	0.5	0.8	0.95	1	
Selenium	µg/L	5	3	1	1	1	1	1	1	
Silver	µg/L	0.05	3	0.5	0.5	0.5	0.5	0.5	0.5	
Strontium	µg/L	_	3	10	12.8	17	20.6	22.4	23	
Thorium	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5	
Tin	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5	
Uranium	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5	
Vanadium	µg/L	_	3	5	5	5	5	5	5	
Zinc	µg/L	8	3	6	6	6	9.6	11.4	12	
Soluble Metals									_	
Aluminium	µg/L	55	3	47	48.6	51	177	240	261	
Arsenic	µg/L	13	0	_	-	_	_	_	_	
Barium	µg/L	_	0	_	-	_	_	_	_	
Beryllium	µg/L	_	0	_	-	_	_	_	_	
Boron	µg/L	370	0	_	-	_	_	_	_	
Cadmium	µg/L	0.2	0	_	-	_	_	_	_	
Chromium	µg/L	1	0	-	-	_	-	-	-	
Cobalt	µg/L	_	0	-	-	_	-	-	-	
Copper	µg/L	1.4	0	-	-	_	-	-	-	
Iron	µg/L	_	64	650	1120	1605	2116	2579	3440	

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Lead	µg/L	3.4	0	-	_	-	_	-	-
Lithium	µg/L	_	0	_	-	_	-	_	_
Manganese	µg/L	1900	64	14.5	92.88	162	277.6	320	366
Molybdenum	µg/L	11	0	-	-	-	-	-	-
Nickel	µg/L	_	0	-	_	-	_	-	_
Silver	µg/L	0.05	0	-	-	-	-	-	-
Strontium	µg/L	_	0	-	-	-	-	-	-
Thorium	µg/L	_	0	-	-	-	-	-	-
Tin	µg/L	_	0	-	-	-	-	-	-
Uranium	µg/L	_	0	-	-	-	-	-	-
Vanadium	µg/L	_	0	-	-	-	-	-	-
Zinc	µg/L	8	0	-	-	-	-	-	-
Algae									
Chlorophyll a - Field	µg/L	5	0	-	-	-	-	-	-
Chlorophyll a	µg/L	5	0	_	_	_	_	_	_
Microbiological									
Cryptosporidium - Corrected	MPN/100mL	_	0	-	_	-	_	-	_
Cryptosporidium - Detected	MPN/100mL	_	1	2.5	2.5	2.5	2.5	2.5	2.5
E.coli - Colilert	MPN/100mL	_	66	4	24	77	240	595	1700
E.coli - MF	MPN/100mL	_	0	_	_	_	_	_	_

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Enterococci - MF	MPN/100mL	-	66	2	13	37	100	322.5	880
Giardia - Corrected	MPN/100mL	_	0	-	-	_	-	_	_
Giardia - Detected	MPN/100mL	-	2	0.7	1.06	1.6	2.14	2.41	2.5
Total Coliforms - Colilert	MPN/100mL	-	66	180	1400	2400	2400	5150	7400
Pesticide									
2.4-D	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4-DB	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4-DP	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4.5-T	µg/L	36	2	0.005	0.005	0.005	0.005	0.005	0.005
2.4.6-T	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
2.6-D	µg/L	280	2	0.05	0.05	0.05	0.05	0.05	0.05
3-Hydroxy Carbofuran	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
4-Chlorophenoxy acetic acid	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
4.4-DDD (pp-DDD)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
4.4-DDE (pp-DDE)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
4.4-DDT (pp-DDT)	µg/L	0.006	2	0.001	0.001	0.001	0.001	0.001	0.001
Acrolein	µg/L	_	1	1	1	1	1	1	1
Aldicarb	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Aldrin (Eldrin)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
alpha-BHC (lindane)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Ametryn	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Amitrole	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
AMPA	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Asulam	µg/L	_	2	1	1	1	1	1	1
Atrazine	µg/L	13	2	0.005	0.005	0.005	0.005	0.005	0.005
Azinphos methyl	µg/L	0.01	2	0.01	0.01	0.01	0.01	0.01	0.01
Azinphos-ethyl	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Bendiocarb	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Benomyl	µg/L	_	2	0.005	0.006	0.0075	0.009	0.00975	0.01
beta-BHC (lindane)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Bioresmethrin	µg/L	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Brodifacoum	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Bromacil	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Bromophos-ethyl	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Bromoxynil	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Carbaryl	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Carbofuran	µg/L	0.06	2	0.005	0.005	0.005	0.005	0.005	0.005
Carbophenothion	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Chlordane - Total	µg/L	0.03	2	0.001	0.001	0.001	0.001	0.001	0.001
Chlorfenvinphos - Total	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01

	Lipito W(Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Chlorothalonil	µg/L	_	2	1	1	1	1	1	1
Chlorpyrifos	µg/L	0.01	2	0.01	0.01	0.01	0.01	0.01	0.01
Chlorpyrifos-methyl	μg/L	_	2	0.1	0.1	0.1	0.1	0.1	0.1
Chlorsulfuron	µg/L	_	2	0.1	0.1	0.1	0.1	0.1	0.1
cis-Chlordane	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Clopyralid	μg/L	-	2	0.025	0.025	0.025	0.025	0.025	0.025
Coumaphos	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Cyanazine	μg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Cyproconazole	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Cyprodinil	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Cyromazine	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
delta-BHC	μg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Demeton-O & Demeton-S	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Demeton-S methyl	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Diazinon	μg/L	0.01	2	0.005	0.005	0.005	0.005	0.005	0.005
Dicamba	μg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Dichlorvos (DDVP)	μg/L	_	2	0.1	0.1	0.1	0.1	0.1	0.1
Diclofop-methyl	μg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Dieldrin	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Difenoconazole	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01

	Units		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Diflufenican	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Dimethoate	µg/L	0.15	2	0.01	0.01	0.01	0.01	0.01	0.01
Diquat	µg/L	1.4	1	0.025	0.025	0.025	0.025	0.025	0.025
Disulfoton	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Diuron	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Endosulfan - Total	µg/L	0.03	2	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan I (alpha isomer)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan II (beta isomer)	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endosulfan sulfate	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endrin	µg/L	0.01	2	0.001	0.001	0.001	0.001	0.001	0.001
Endrin aldehyde	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Endrin ketone	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
EPN	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Ethion	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Ethoprophos (Prophos)	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Fenamiphos	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Fenarimol	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Fenchlorphos (Ronnel)	µg/L	_	2	5	5	5	5	5	5
Fenitrothion	µg/L	0.2	2	1	1	1	1	1	1
Fensulphothion	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005

	Units		Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Fenthion	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Fenvalerate & Esfenvalerate	µg/L	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Fipronil	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Fluometuron	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Fluroxypyr	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Flusilazole	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Glyphosate	µg/L	370	2	0.5	0.5	0.5	0.5	0.5	0.5
Heptachlor	µg/L	0.01	2	0.0005	0.0005	0.0005	0.0005	0.0005	0.0005
Heptachlor epoxide	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Hexachlorobenzene (HCB)	µg/L	-	2	0.001	0.001	0.001	0.001	0.001	0.001
Hexazinone	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Iprodione	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Irgarol	µg/L	-	2	0.001	0.001	0.001	0.001	0.001	0.001
Lindane (gamma-BHC)	µg/L	0.2	2	0.001	0.001	0.001	0.001	0.001	0.001
Malathion (Maldison)	µg/L	0.05	2	0.01	0.01	0.01	0.01	0.01	0.01
MCPA	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
МСРВ	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Mecoprop	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Methiocarb	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Methomyl	µg/L	3.5	2	0.005	0.005	0.005	0.005	0.005	0.005

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Methoxychlor	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Metolachlor	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Metribuzin	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Metsulfuron-methyl	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Mevinphos	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Molinate	µg/L	3.4	2	0.05	0.05	0.05	0.05	0.05	0.05
Monocrotophos	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Omethoate	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Oryzalin	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Oxamyl	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Oxychlordane	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Oxyfluorfen	µg/L	_	2	0.5	0.5	0.5	0.5	0.5	0.5
Paclobutrazol	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Paraquat	µg/L	_	1	0.05	0.05	0.05	0.05	0.05	0.05
Parathion	µg/L	0.004	2	0.1	0.1	0.1	0.1	0.1	0.1
Parathion-methyl	µg/L	_	2	1	1	1	1	1	1
Penconazole	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Pendimethalin	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Permethrin	µg/L	_	1	0.25	0.25	0.25	0.25	0.25	0.25
Phorate	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
			Surface						
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Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Picloram	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Piperonyl Butoxide	µg/L	-	1	0.25	0.25	0.25	0.25	0.25	0.25
Pirimiphos-ethyl	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Pirimiphos-methyl	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Profenofos	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Prometryn	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Propazine	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Propiconazole	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Prothiophos (Tokuthion)	µg/L	_	2	0.05	0.05	0.05	0.05	0.05	0.05
Pyrimethanil	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Silvex (2,4,5-TP/Fenoprop)	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Simazine	µg/L	3.2	2	0.01	0.01	0.01	0.01	0.01	0.01
Sulprofos	µg/L	_	2	0.025	0.025	0.025	0.025	0.025	0.025
Sum of DDD + DDE + DDT	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Tebuconazole	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Tebuthiuron	µg/L	2.2	2	0.01	0.01	0.01	0.01	0.01	0.01
Temephos	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01
Terbufos	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Terbuthylazine	µg/L	_	2	0.005	0.008	0.0125	0.017	0.01925	0.02
Terbutryn	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005

			Surface						
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Tetrachlorvinphos	µg/L	_	2	0.005	0.005	0.005	0.005	0.005	0.005
Tetraethyl dithiopyrophos	µg/L	-	2	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Thiamethoxam	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Thiobencarb	µg/L	2.8	2	0.005	0.005	0.005	0.005	0.005	0.005
Thiodicarb	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
trans-Chlordane	µg/L	_	2	0.001	0.001	0.001	0.001	0.001	0.001
Triazophos	µg/L	-	2	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Trichlorfon	µg/L	-	2	0.01	0.01	0.01	0.01	0.01	0.01
Trichloronate (Trichloronat)	µg/L	-	2	0.25	0.25	0.25	0.25	0.25	0.25
Triclopyr	µg/L	-	2	0.005	0.005	0.005	0.005	0.005	0.005
Trifluralin	µg/L	2.6	2	5	5	5	5	5	5
Polyaromatic Hydrocarbon									
2-Chloronaphthalene	µg/L	-	3	0.05	0.05	0.05	0.05	0.05	0.05
2-Methylnapthalene	µg/L	_	3	0.05	0.05	0.05	0.05	0.05	0.05
Acenaphthene	µg/L	-	3	0.01	0.01	0.01	0.01	0.01	0.01
Acenapthylene	µg/L	-	3	0.01	0.01	0.01	0.01	0.01	0.01
Anthracene	µg/L	-	3	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[a]anthracene	µg/L	-	3	0.01	0.01	0.01	0.01	0.01	0.01
Benzo[a]pyrene	µg/L	-	3	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025
Benzo[b+j]fluoranthene	µg/L	-	3	0.01	0.01	0.01	0.01	0.01	0.01

			Surface	Surface							
Parameter	Units	WQO	Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum		
Benzo[e]pyrene	µg/L	_	3	0.05	0.05	0.05	0.05	0.05	0.05		
Benzo[ghi]perylene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Benzo[k]fluoranthene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Chrysene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Dibenzo[ah]anthracene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Fluoranthene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Fluorene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Indeno(1.2.3-cd)pyrene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Napthalene	µg/L	16	3	0.01	0.01	0.01	0.01	0.01	0.01		
Perylene	µg/L	_	3	0.05	0.05	0.05	0.05	0.05	0.05		
Phenanthrene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Pyrene	µg/L	_	3	0.01	0.01	0.01	0.01	0.01	0.01		
Total PAH	µg/L	_	3	0.0025	0.0025	0.0025	0.0025	0.0025	0.0025		
Taste and Colour											
2-Methylisoborneol	ng/L	_	0	_	_	_	-	_	_		
2-methylisoborneol + Geosmin	ng/L	_	0	_	_	_	-	_	_		
Geosmin	ng/L	_	0	_	-	-	-	_	_		
Volatile Organic Compound											
Benzene	µg/L	950	3	0.25	0.25	0.25	0.25	0.25	0.25		
Toluene	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5		

	Units	WQO	Surface						
Parameter			Count	minimum	20th percentile	median	80th percentile	95th percentile	maximum
Ethylbenzene	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5
meta & para-Xylene	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5
ortho-Xylene	µg/L	350	3	0.5	0.5	0.5	0.5	0.5	0.5
Total Xylenes	µg/L	_	3	0.5	0.5	0.5	0.5	0.5	0.5
Hexaconazole	µg/L	_	2	0.01	0.01	0.01	0.01	0.01	0.01

Grey shading denotes parameters that exceed the water quality object / trigger value

Gold shading denotes parameters that have an LOR higher than the water quality objective / trigger value

Appendix E Summary of frc environmental Water Quality Monitoring Data for Lake Macdonald and Six Mile Creek

Six Mile Creek – Downstream

SMC4

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	5	5	5	5	5	5
minimum	19.3	135	6.24	2.8	33.8	4.8
20 th percentile	20.5	146	6.75	4.1	46.0	6.3
median	20.9	159	7.08	4.5	49.1	11.4
80 th percentile	23.4	175	7.21	5.7	64.7	12.4
maximum	25.5	197	7.30	5.9	65.8	13.9

SMC5

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	5	5	5	5	5	5
minimum	17.7	160	5.82	2.0	24.0	5.4
20 th percentile	19.3	173	6.36	2.6	29.9	7.9
median	21.0	189	7.14	4.1	41.7	9.3
80 th percentile	23.4	239	7.34	5.5	63.4	11.2
maximum	24.6	244	7.43	6.1	68.8	15.6

SMCDS05

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	1	1	1	1	1	1
minimum	18.3	185	7.16	7.6	80.4	10.8
20 th percentile	18.3	185	7.16	7.6	80.4	10.8
median	18.3	185	7.16	7.6	80.4	10.8
80 th percentile	18.3	185	7.16	7.6	80.4	10.8
maximum	18.3	185	7.16	7.6	80.4	10.8

SMCDS04

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	3	3	3	3	3	3
minimum	18.8	110	6.16	2.2	26.7	7.8
20 th percentile	18.9	133	6.32	2.5	26.8	8.6
median	19.1	167	6.56	3.0	26.9	9.8
80 th percentile	22.4	173	6.73	4.4	30.6	12.0
maximum	24.5	177	6.85	5.3	33.0	13.5

SMCDS03

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	3	3	3	3	3	3
minimum	18.3	117	5.55	3.3	39.7	7.3
20 th percentile	18.5	135	6.01	3.5	40.3	7.6
median	18.9	162	6.70	3.8	41.2	7.9
80 th percentile	22.2	168	6.81	4.8	51.6	9.3
maximum	24.4	172	6.89	5.5	58.6	10.3

SMCDS02

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	3	3	3	3	3	3
minimum	20.3	89	5.72	3.0	37.8	2.5
20 th percentile	20.4	113	6.07	3.9	46.4	2.6
median	20.5	148	6.60	5.3	59.2	2.9
80 th percentile	23.8	159	6.95	5.4	60.3	2.9
maximum	26.0	166	7.18	5.4	61.0	3.0

SMCDS01

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	4	4	4	4	4	4
minimum	20.0	76	5.94	3.2	39.9	5.1
20 th percentile	22.4	85	6.26	3.9	48.2	5.2
median	24.8	98	6.49	5.8	67.3	5.4
80 th percentile	26.0	112	6.82	7.6	85.8	5.9
maximum	26.6	122	7.31	7.9	93.3	6.4

Six Mile Creek – In Lake Macdonald

DS Lake

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	4	4	4	4	4	4
minimum	23.2	42	6.69	4.2	54.8	3.5
20th percentile	24.1	69	6.78	6.3	82.2	3.5
median	26.4	89	6.99	8.8	108.8	4.6
80th percentile	28.3	96	7.26	9.9	118.2	5.7
maximum	28.8	103	7.46	10.0	119.8	5.8

US Lake

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	2	2	2	2	2	2
minimum	22.9	53	6.77	8.2	99.2	3.1
20 th percentile	23.2	60	6.88	8.5	101.5	3.6
median	23.6	70	7.04	8.8	105.0	4.3
80 th percentile	24.0	80	7.20	9.2	108.4	4.9
maximum	24.3	87	7.31	9.4	110.7	5.4

Six Mile Creek – Upstream of Lake Macdonald

CU02

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	2	2	2	2	2	2
minimum	21.6	59	6.31	6.4	72.9	5.7
20 th percentile	22.4	66	6.45	6.7	77.5	6.1
median	23.6	77	6.67	7.1	84.5	6.6
80 th percentile	24.9	88	6.88	7.5	91.5	7.0
maximum	25.7	95	7.02	7.8	96.1	7.4

CU01

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	2	2	2	2	2	2
minimum	17.4	82	6.16	4.9	51.1	4.5
20 th percentile	17.8	93	6.38	5.7	62.9	6.0
median	18.6	109	6.72	6.9	80.6	8.3
80 th percentile	19.3	126	7.06	8.1	98.3	10.7
maximum	19.8	137	7.28	8.9	110.1	12.2

CU03

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	2	2	2	2	2	2
minimum	21.2	92	6.27	4.1	46.2	12.2
20 th percentile	54.6	103	6.44	4.8	56.0	12.8
median	104.6	119	6.71	5.8	70.7	13.8
80 th percentile	154.6	135	6.97	6.8	85.4	14.8
maximum	188.0	146	7.14	7.4	95.2	15.4

SMCUS02

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	2	2	2	2	2	2
minimum	19.3	92	6.53	3.4	35.6	4.7
20 th percentile	20.1	111	6.56	4.2	45.7	6.6
median	21.3	141	6.61	5.3	61.0	9.6
80 th percentile	22.4	170	6.65	6.4	76.2	12.5
maximum	23.2	189	6.68	7.2	86.3	14.5

SMCUS01

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	3	3	3	3	3	3
minimum	18.7	107	6.24	2.1	24.7	5.9
20 th percentile	18.9	121	6.41	2.2	25.9	7.8
median	19.1	141	6.67	2.3	27.7	10.8
80 th percentile	22.4	142	7.23	3.7	41.1	13.0
maximum	24.5	142	7.61	4.6	50.1	14.5

SMCUS03

Summary statistic	Temperature (°C)	Electrical Conductivity (µS/cm)	pH (unit)	Dissolved Oxygen (mg/L)	Dissolved Oxygen (% saturation)	Turbidity (NTU)
count	2	2	2	2	2	2
minimum	18.7	87	5.80	2.8	33.1	6.2
20 th percentile	19.4	97	6.04	3.4	38.4	6.5
median	20.6	112	6.41	4.2	46.3	6.8
80 th percentile	21.7	127	6.77	5.0	54.2	7.1
maximum	22.5	137	7.01	5.5	59.5	7.3

Appendix F Detailed Fish and Turtle Survey Methods

The survey methods were consistent with methods for the survey of large freshwater perches and lungfish presented in the *Survey Guidelines for Australia's Threatened Fish* (SEWPAC 2011a) and the *Survey Guidelines for Australia's Threatened Reptiles* (SEWPAC 2011b), and included use of:

- Active fishing methods:
 - Underwater camera with live viewing. Where turbidity and light conditions allowed, an underwater camera with live viewing was used to actively search for the MNES species at each site. For watercourse sites, the full length of the site was walked on the bank (where possible) so as not to disturb fish and turtles, with the camera mounted on a pole to provide live footage of underwater life in deep pools, under logs, in undercut banks and amongst other habitat features. For reservoir sites, the camera was mounted on a pole and operated from the boat prior to electrofishing;
 - Electrofishing. Experienced and certified operators electrofished in accordance with the Australian Code of Electrofishing Practice; boat units were used at the two sites in the reservoir zone; backpack units were used in the watercourse sites in the upstream and downstream zones⁹;
 - Angling. Two set lines using baited barbless hooks were set at each site for one hour at dusk on each survey;
- · Passive fishing methods:
 - Fyke nets. Two fyke nets were set overnight with appropriate floating devices to ensure that air breathing species had access to the surface at all times. For watercourse sites in the upstream and downstream zones deep pools were targeted, with one fyke was oriented in the upstream direction and the other in the downstream direction. At the reservoir sites, fyke nets were set at sites where water depth allowed (i.e. if water was too deep, i.e. > 3 m, then fyke nets could not be set);

⁹ Electrofishing was not used in the downstream zone during the August survey, so as not to interfere with a research project that is assessing Mary River cod recruitment and larval ecology. However, electrofishing was used as site SMCDS05 during this survey to provide additional fish data, noting that fish caught at this site could be vagrants from the main stem of the Mary River.

- Baited box traps / bait fish traps. Five box traps were set overnight to sample small / juvenile fish. These traps were placed in appropriate habitat, such as shallow water amongst aquatic plants or other habitat features;
- Baited cathedral traps. Three cathedral traps, baited with ox heart meat, were set overnight to capture turtles at sites that had suitable characteristics for setting cathedral traps.

The sampling effort used at each site on each survey is presented in Tables C1 and C2.

All fish and turtle species were identified to species in the field. Identification of some turtle species was confirmed in the office by review of photographs taken during the field surveys.

The total length of all fish was measured in the field, and the life history stage (juvenile, intermediate, adult) of turtles was recorded.

Fish and turtle surveys were conducted under ethics (CA 2015/08/893) and fisheries (181742) permits held by frc environmental, and all specimens of native fish and turtle were released unharmed at the location of capture. Pest species of fish were euthanised in accordance with methods approved under the ethics permit.

The Department of Natural Resources and Mines provided fish data for a survey (November 2015) of large bodied fish (boat electrofishing) at the Lake Macdonald tail water, which included records for Mary River cod and Australian lungfish at this site.

Site	Method	Habitat	Date	Time In	Time Out	Settings	Effort ^a
Downstream	Zone						
SMCDS05	backpack electrofishing	pool	2015-08-28	1400	1510	225 V	1208 s
						30 Hz	
SMCDS04	small bait traps (5)	pool	2015-08-28	1200	0915		106.25 h
	fyke nets (2)						42.5 h
	cathedral trap (3)						63.75
	line fishing (2)			1640	1740		2 h
SMCDS03	small bait traps (5)	pool	2015-08-28	1115	0845		107.5 h
	fyke nets (2)						43 h
	cathedral trap (3)						64.5 h

Table F.1Fishing effort for the August 2015 survey.

F3

Site	Method	Habitat	Date	Time In	Time Out	Settings	Effort ^a
	line fishing (2)			1630	1730		2 h
SMCDS02	small bait traps (5)	pool	2015-08-27	1430	0930		95 h
	fyke nets (2)						38 h
	cathedral trap (3)						57 h
	line fishing (2)			1000	1045		1.5 h
SMCDS01	small bait traps (5)	pool	2015-08-27	1330	0900		97.5 h
	fyke nets (2)						39 h
	cathedral trap (3)						58.5 h
	line fishing (2)			1715	1830		2.5 h
Reservoir Zo	ne						
DS Lake	small bait traps (5)	pool	2015-08-26	1630	1000		87.5 h
	fyke nets (2)						35 h
	cathedral trap (3)						52.5 h
	line fishing (2)		2015-08-26	1645	1730		1.5 h
	boat electrofishing			1645	1730	50–1000 V	2000 s
						4 amps	
						100%	
US Lake	small bait traps (5)	pool	2015-08-26	1600	1015		91.25 h
	fyke nets (2)						36.5 h
	cathedral trap (3)						54.75 h
	line fishing (2)		2015-08-25	1800	1845		1.5 h
	boat electrofishing			1445	1555	50–1000 V	2000 s
						4 amps	
						100%	
Upstream Zo	one						
CU02	small bait traps (5)	pool	2015-08-25	1130	900		107.5 h
	fyke nets (2)						43 h
	cathedral traps (3)						64.5 h
	boat electrofishing			1245	1345	50–1000 V	2000 s
						4 amps	
						100%	
	line fishing (2)		2015-08-25	1630	1730		2 h

Site	Method	Habitat	Date	Time In	Time Out	Settings	Effort ^a
CU01	fyke nets (2)	pool	2015-08-24	1445	1200		42.5 h
	small bait traps (5)						106.25 h
	cathedral traps (3)						63.75 h
	boat electrofishing			1245	1430	50–1000 V	2000 s
						4 amps	
						80%	
CU03	fyke nets (2)	pool	2015-08-25	1245	1000		42.5 h
	small bait traps (5)						106.25 h
	cathedral traps (3)						63.75 h
	line fishing (2)		2015-08-26	1745	1845		2 h
	boat electrofishing			1145	1230	50–1000 V	2000 s
						4 amps	
						80%	
SMCUS02	small bait traps (5)	pool	2015-08-29	1200	0800		100 h
	fyke nets (2)						40 h
	cathedral trap (3)						60 h
	line fishing (2)			1645	1745		2 h
SMCUS01	small bait traps (5)	pool	2015-08-24	1700	0900		80 h
	fyke nets (2)			1700	0915		32.5 h
	cathedral trap (3)						48.75 h
	line fishing (2)			1730	1830		2 h
SMCUS03	small bait traps (5)	pool	2015-08-29	1045	730		103.75 h
	fyke nets (2)						41.5 h
	cathedral trap (3)						62.25
	line fishing (2)			1530	1630		2 h
a s – seco	onds: h – hours						

s – seconds; h – hours

Site	Method	Habitat	Date	Time In	Time Out	Settings	Effort ^a
Downstream	n Zone						
SMCDS04	small bait traps (5)	pool	2015-10-20	1745	0730	_	68.75 h
	fyke nets (2)						27.5 h
	cathedral trap (3)						41.25 h
	line fishing (2)			1745	1815		1 h
	backpack electrofishing			1630	1730	270 V	2001 s
						40 Hz	
SMCDS03	small bait traps (5)	pool	2015-10-20	1530	900	-	87.5 h
	fyke nets (2)						35 h
	cathedral trap (3)						52.5 h
	backpack electrofishing			1245	1500	265 V	2012 s
						40 Hz	
	line fishing (2)		2015-10-21	720	840		2.67 h
SMCDS02	small bait traps (5)	pool	2015-10-18	1735	715	-	68.33 h
	fyke nets (2)						27.33 h
	cathedral trap (3)						41 h
	line fishing (2)			1755	1850		1.83 h
	backpack electrofishing			1515	1715	275 V	2007 s
						40 Hz	
SMCDS01	small bait traps (5)	pool	2015-10-18	1400	900		95 h
	fyke nets (2)						38 h
	cathedral trap (3)						57 h
	line fishing (2)			1820	1900		1.33 h
	backpack electrofishing			1045	1300	240 V	2004 s
						40 Hz	
Reservoir Z	one						
DS Lake	small bait traps (5)	pool	2015-10-19	1630	1015		88.75 h
	fyke nets (2)						35.5 h

Table F.2	Fishing e	ffort for the	October	2015	survey.

Site	Method	Habitat	Date	Time In	Time Out	Settings	Effort ^a
	cathedral trap (3)						53.25 h
	line fishing (2)			1800	1830		1 h
	boat electrofishing			1400	1500	50–1000 V	2004 s
						4–7 amps	
						100%	
US Lake	small bait traps (5)	pool	2015-10-19	1700	930		82.5 h
	fyke nets (2)						33 h
	cathedral trap (3)						49.5 h
	line fishing (2)			1730	1750		0.67 h
	boat electrofishing			1115	1245	50–1000 V	2000 s
						4–7 amps	
						100%	
Upstream Zo	ne						
CU02	small bait traps (4)	pool	2015-10-22	1445	1030		79 h
	fyke nets (2)						39.5 h
	cathedral trap (2)						39.5 h
	line fishing (2)		2015-10-23	1830	1900		1 h
	boat electrofishing			1515	1630	50–1000 V	2002 s
						4–7 amps	
						100%	
CU01	small bait traps (4)	pool	2015-10-22	1530	0915		71 h
	fyke nets (2)						35.5 h
	cathedral trap (3)						53.25 h
	boat electrofishing		2015-10-23	1330	1500	50–1000 V	2004 s
						4–7 amps	
						100%	
CU03	small bait traps (4)	pool	2015-10-22	1245	0830		79 h
	fyke nets (2)						39.5 h
	cathedral trap (3)						59.25 h
	boat electrofishing		2015-10-23	1500	1645	50–1000 V	2018 s
						4–7 amps	
						100%	

Site	Method	Habitat	Date	Time In	Time Out	Settings	Effort ^a
SMCUS02	small bait traps (5)	pool	2015-10-21	1700	1000		85 h
	fyke nets (2)						34 h
	cathedral trap (3)						51 h
	line fishing (2)			1745	1820		1.17 h
	boat electrofishing			1500	1645	50–1000 V	2000 s
						4–7 amps	
						100%	
SMCUS01	small bait traps (5)	pool	2015-10-21	1215	0820		100.42 h
	fyke nets (2)						40.17 h
	cathedral trap (3)						60.3 h
	line fishing (2)			1830	1850		0.67 h
	backpack electrofishing			1230	1400	240 V	2018 s
						40 Hz	
SMCUS03	backpack electrofishing	pool	2015-10-22	1615	1745	275 V	2008 s
						40 Hz	
	line fishing (2)			1745	1815		1 h
	small bait traps (5)		2015-10-23	1730	0720		69.17 h
	fyke nets (2)						27.67 h
	cathedral trap (3)						41.5 h

^a s – seconds; h – hours

G1

Appendix G Detailed Fish Survey Results

Table G.1Summary of fish survey counts.

Species	Common Name	Caught August 2015 Survey	Caught October 2015 Survey
Native Species			
Ambassidae			
Ambassis agassizii	Agassiz's glassfish	112	188
Ambassis marianus	estuary glassfish	0	0
Anguillidae			
Anguilla australis	southern shortfin eel	2	0
Anguilla reinhardtii	longfin eel	11	38
Apogonidae			
Glossamia aprion	mouth almighty	0	1
Atherinidae			
Craterocephalus marjoriae	silverstreak hardyhead	0	0
Craterocephalus stercusmuscarum	flyspecked hardyhead	60	347
Eleotridae			
Gobiomorphus australis	striped gudgeon	0	0
<i>Hypseleotris</i> spp.	common carp gudgeons	1106	3099
Hypseleotris compressa	empire gudgeon	0	0
Mogurnda adspersa	purple spotted gudgeon	0	13
Philypnodon macrostomus	dwarf flathead gudgeon	5	26
Philypnodon grandiceps	flathead gudgeon	34	20
Melanotaeniidae			
Melanotaenia duboulayi	crimson-spotted rainbowfish	83	581
Percichthyidae			
Maccullochella mariensis	Mary River cod	0	0 (3)
Percalates novemaculeata	Australian bass	2	15 (75)
Macquaria ambigua	yellowbelly	0	0 (2)

Species	Common Name	Caught August 2015 Survey	Caught October 2015 Survey
Plotosidae			
Tandanus tandanus	eel-tailed catfish	14	23 (5)
Neosilurus hyrtlii	Hyrtl's tandan	0	0
Pseudomugilidae			
Pseudomugil signifer	Pacific blue-eye	1	27
Retropinnidae			
Retropinna semoni	Australian smelt	25	104
Clupeidae			
Nematolosa erebi	bony bream	31	112 (16)
Ceratodontidae			
Neoceratodus forsteri	Australian lungfish	0	0 (1)
Mugilidae			
Trachystoma petardi	pinkeye mullet	0	0
Mugil cephalus	sea mullet	0	0
Tetrarogidae			
Notesthes robusta	bullrout	0	0
Terapontidae			
Leiopotherapon unicolor	spangled perch	0	0 (20)
Osteoglossidae			
Scleropages leichardti	southern saratoga	0	0 (3)
Pest Species			
Gambusia holbrooki	eastern Gambusia	55	62
Xiphophorus maculatus	platy	0	0
Xiphophorus hellerii	swordtail	0	2
Poecilia reticulata	guppy	0	0

Bracketed numbers were provided by DNRM during the expert consultation process and were the result of a survey completed by DNRM in November 2015 at the Six Mile Creek Dam tailwater.

Site	Common Name	Scientific Name	Method	Catch	CPUE (catch per hour)
Downstream	zone				
SMCDS05	carp gudgeon	Hypseleotris spp.	BPEF	1	0.05
	flathead gudgeon	Philypnodon grandiceps	BPEF	1	0.05
	freshwater catfish	Tandanus tandanus	BPEF	1	0.05
	longfin eel	Anguilla reinhardtii	BPEF	6	0.30
SMCDS05 To	tal			9	0.45
SMCDS04	Australian smelt	Retropinna semoni	Fyke	1	0.01
	carp gudgeon	Hypseleotris spp.	Fyke	8	0.05
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	9	0.06
	Pacific blue eye	Pseudomugil signifer	Fyke	1	0.01
SMCDS04 To	tal			19	0.13
SMCDS03	Australian smelt	Retropinna semoni	Fyke	18	0.12
	carp gudgeon	Hypseleotris spp.	Fyke	4	0.03
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	6	0.04
	longfin eel	Anguilla reinhardtii	Fyke	1	0.02
SMCDS03 To	tal			29	0.19
SMCDS02	Agassiz's glassfish	Ambassis agassizii	Fyke	18	0.14
	carp gudgeon	<i>Hypseleotris</i> spp.	Fyke	50	0.38

 Table G.2
 Catch Per Unit Effort (CPUE) of native fish species caught at each site during the August 2015 survey.

Site	Common Name	Scientific Name	Method	Catch	CPUE (catch per hour)
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	9	0.07
	flathead gudgeon	Philypnodon grandiceps	Fyke	2	0.02
	flyspecked hardyhead	Craterocephalus stercusmuscarum	Fyke	1	0.01
	freshwater catfish	Tandanus tandanus	Fyke	2	0.05
SMCDS02 To	tal			82	0.61
SMCDS01	Agassiz's glassfish	Ambassis agassizii	Fyke	16	0.12
	carp gudgeon	Hypseleotris spp.	Bait, Fyke	45	0.33
	freshwater catfish	Tandanus tandanus	Fyke	2	0.05
SMCDS01 To	tal			63	0.45
Reservoir zoi	ne				
DS Lake	Agassiz's glassfish	Ambassis agassizii	BPE, Fyke	35	0.22
	bony bream	Nematalosa erebi	BPE	9	0.13
	carp gudgeon	Hypseleotris spp.	BPE, Fyke	270	1.73
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	2	0.01
	flathead gudgeon	Philypnodon grandiceps	Fyke	3	0.02
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BPE, Fyke	35	0.22
	freshwater catfish	Tandanus tandanus	Fyke	3	0.04
DS Lake Tota	1			357	2.27
US Lake	Agassiz's glassfish	Ambassis agassizii	BPE, Fyke	9	0.06
	Australian bass	Percalates novemaculeata	BPE	1	0.01

Site	Common Name	Scientific Name	Method	Catch	CPUE (catch per hour)
	Australian smelt	Retropinna semoni	BPE	1	0.01
	bony bream	Nematalosa erebi	BPE	2	0.03
	carp gudgeon	Hypseleotris spp.	BPE, Fyke	195	1.21
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	4	0.02
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BPE, Fyke	18	0.11
US Lake Tota	l			230	1.41
Upstream zo	ne				
CU02	Agassiz's glassfish	Ambassis agassizii	Fyke	15	0.08
	bony bream	Nematalosa erebi	BEF, Fyke	13	0.17
	carp gudgeon	Hypseleotris spp.	BEF, Bait, Fyke	60	0.33
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BEF, Fyke	5	0.03
	freshwater catfish	Tandanus tandanus	Fyke	1	0.01
	longfin eel	Anguilla reinhardtii	Fyke	2	0.03
CU02 Total				96	0.52
CU01	Agassiz's glassfish	Ambassis agassizii	Fyke	3	0.02
	Australian bass	Percalates novemaculeata	BEF	1	0.01
	carp gudgeon	Hypseleotris spp.	BEF, Bait, Fyke	128	0.70
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BEF, Fyke	13	0.07
	flathead gudgeon	Philypnodon grandiceps	Fyke	4	0.02
	flyspecked hardyhead	Craterocephalus stercusmuscarum	Fyke	1	0.01

Site	Common Name	Scientific Name	Method	Catch	CPUE (catch per hour)
	southern shortfin eel	Anguilla australis	Line	2	0.03
CU01 Total				152	0.83
CU03	Agassiz's glassfish	Ambassis agassizii	Fyke	5	0.03
	bony bream	Nematalosa erebi	BEF, Fyke	3	0.04
	carp gudgeon	Hypseleotris spp.	BEF, Bait, Fyke	155	0.85
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BEF	4	0.02
	flathead gudgeon	Philypnodon grandiceps	BEF, Fyke	15	0.08
	freshwater catfish	Tandanus tandanus	Fyke	2	0.03
	longfin eel	Anguilla reinhardtii	Line	1	0.01
CU03 Total				185	1.00
SMCUS02	Agassiz's glassfish	Ambassis agassizii	Fyke	10	0.07
	carp gudgeon	Hypseleotris spp.	Fyke	50	0.36
	dwarf flathead gudgeon	Philypnodon macrostomus	Fyke	5	0.04
	flathead gudgeon	Philypnodon grandiceps	Fyke	5	0.04
SMCUS02 To	tal			70	0.49
SMCUS01	Agassiz's glassfish	Ambassis agassizii	Fyke	1	0.01
	Australian smelt	Retropinna semoni	Fyke	5	0.04
	bony bream	Nematalosa erebi	Fyke	4	0.12
	carp gudgeon	Hypseleotris spp.	Bait	80	0.71
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	1	0.01

Site	Common Name	Scientific Name	Method	Catch	CPUE (catch per hour)
	freshwater catfish	Tandanus tandanus	Fyke	2	0.06
SMCUS01 To	tal		93	0.81	
SMCUS03	carp gudgeon	Hypseleotris spp.	Line	60	0.41
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Line	35	0.24
	flathead gudgeon	Philypnodon grandiceps	Line	4	0.03
	freshwater catfish	Tandanus tandanus	Line	1	0.02
	longfin eel	Anguilla reinhardtii	Line	1	0.02
SMCUS03 To	tal			101	0.69

Site	Common Name	Scientific Name	Method	Catch	CPUE (hr)
Downstream	zone				
SMCDS04	Agassiz's glassfish	Ambassis agassizii	Fyke	1	0.01
	Australian smelt	Retropinna semoni	Fyke	2	0.03
	carp gudgeon	Hypseleotris spp.	BPEF, Box, Fyke	78	0.60
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BPEF, Box, Fyke	60	0.46
	dwarf flathead gudgeon	Philypnodon macrostomus	BPEF	2	0.02
	flathead gudgeon	Philypnodon grandiceps	BPEF	3	0.02
	freshwater catfish	Tandanus tandanus	Fyke	1	0.02
	Pacific blue eye	Pseudomugil signifer	Fyke	1	0.01
	purple-spotted gudgeon	Mogurnda adspersa	BPEF	5	0.04
SMCDS04 To	tal			153	1.17
SMCDS03	Agassiz's glassfish	Ambassis agassizii	Fyke	4	0.03
	Australian smelt	Retropinna semoni	Fyke	4	0.06
	carp gudgeon	Hypseleotris spp.	BPEF, Fyke	147	0.94
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BPEF, Box, Fyke	102	0.65
	dwarf flathead gudgeon	Philypnodon macrostomus	BPEF, Fyke	2	0.01
	flathead gudgeon	Philypnodon grandiceps	BPEF	2	0.01
	freshwater catfish	Tandanus tandanus	BPEF	1	0.01
	longfin eel	Anguilla reinhardtii	BPEF	4	0.06
	Pacific blue eye	Pseudomugil signifer	BPEF, Fyke	20	0.13

 Table G.3
 Catch Per Unit Effort (CPUE) for native fish species caught at each site during the October 2015 survey.

Site	Common Name	Scientific Name	Method	Catch	CPUE (hr)
	purple-spotted gudgeon	Mogurnda adspersa	BPEF	2	0.01
SMCDS03 Total				288	1.81
SMCDS02	Agassiz's glassfish	Ambassis agassizii	BPEF, Fyke	18	0.14
	carp gudgeon	Hypseleotris spp.	BPEF, Fyke	61	0.47
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BPEF, Box, Fyke	124	0.96
	flathead gudgeon	Philypnodon grandiceps	BPEF	1	0.01
	freshwater catfish	Tandanus tandanus	BPEF, Line	5	0.08
	longfin eel	Anguilla reinhardtii	BPEF, Fyke	11	0.18
	Pacific blue eye	Pseudomugil signifer	Fyke	6	0.05
SMCDS02 To	tal			226	1.73
SMCDS01	Agassiz's glassfish	Ambassis agassizii	BPEF, Fyke	15	0.09
	Australian bass	Percalates novemaculeata	BPEF, Fyke	2	0.03
	carp gudgeon	Hypseleotris spp.	BPEF, Fyke	278	1.67
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BPEF, Fyke	170	1.02
	dwarf flathead gudgeon	Philypnodon macrostomus	BPEF	2	0.01
	flathead gudgeon	Philypnodon grandiceps	BPEF	2	0.01
	flyspecked hardyhead	Craterocephalus stercusmuscarum	Fyke	3	0.02
	freshwater catfish	Tandanus tandanus	BPEF, Line, Fyke	4	0.05
	longfin eel	Anguilla reinhardtii	BPEF	6	0.08
	mouth almighty	Glossamia aprion	Fyke	1	0.01
SMCDS01 To	tal			483	2.88

Site	Common Name	Scientific Name	Method	Catch	CPUE (hr)
Reservoir zo	ne				
DS Lake	Agassiz's glassfish	Ambassis agassizii	BEF, Fyke	50	0.32
	Australian bass	Percalates novemaculeata	BEF	1	0.01
	Australian smelt	Retropinna semoni	BEF	4	0.06
	bony bream	Nematalosa erebi	BEF	5	0.07
	carp gudgeon	Hypseleotris spp.	BEF, Fyke	515	3.27
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BEF, Fyke	32	0.20
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BEF, Fyke	194	1.23
	freshwater catfish	Tandanus tandanus	BEF	5	0.07
DS Lake Tota	I			806	5.08
US Lake	Agassiz's glassfish	Ambassis agassizii	Fyke	55	0.37
	bony bream	Nematalosa erebi	BEF	23	0.34
	carp gudgeon	Hypseleotris spp.	BEF, Box, Fyke	543	3.65
	dwarf flathead gudgeon	Philypnodon macrostomus	BEF	1	0.01
	flathead gudgeon	Philypnodon grandiceps	Fyke	1	0.01
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BEF, Fyke	66	0.44
	longfin eel	Anguilla reinhardtii	Fyke	1	0.01
US Lake Tota	I			690	4.62
Upstream zor	e				
CU02	Agassiz's glassfish	Ambassis agassizii	Fyke	2	0.01
	Australian smelt	Retropinna semoni	BEF	2	0.03

Site	Common Name	Scientific Name	Method	Catch	CPUE (hr)
	bony bream	Nematalosa erebi	BEF	1	0.01
	carp gudgeon Hypseleotris spp.		BEF, Box, Fyke	659	4.34
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BEF, Fyke	12	0.08
	dwarf flathead gudgeon	Philypnodon macrostomus	BEF, Fyke	6	0.04
	flathead gudgeon	Philypnodon grandiceps	Fyke	1	0.01
	freshwater catfish	Tandanus tandanus	BEF	1	0.01
	longfin eel	Anguilla reinhardtii	BEF, Fyke	11	0.15
	purple-spotted gudgeon	Mogurnda adspersa	BEF, Fyke	2	0.01
CU02 Total	Total			697	4.56
CU01	Agassiz's glassfish Ambassis agassizii		Fyke	11	0.08
	Australian bass	Pervalates novemaculeata	BEF	3	0.04
	bony bream	Nematalosa erebi	BEF	14	0.20
	carp gudgeon	Hypseleotris spp.	BEF, Box, Fyke	227	1.62
	crimson-spotted rainbowfish Melanotaenia duboulayi		BEF, Fyke	15	0.11
	flyspecked hardyhead	Craterocephalus stercusmuscarum	Fyke	8	0.06
	longfin eel	Anguilla reinhardtii	BEF	1	0.01
CU01 Total				279	1.99
CU03	Agassiz's glassfish Ambassis agassizii		Fyke	7	0.05
	Australian bass	Percalates novemaculeata	BEF	6	0.08
	bony bream Nematalosa erebi		BEF	38	0.52
	carp gudgeon	Hypseleotris spp.	BEF, Box, Fyke	104	0.68

Site	Common Name	Scientific Name	Method	Catch	CPUE (hr)
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BEF	16	0.11
	flathead gudgeon	Philypnodon grandiceps	Fyke	2	0.01
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BEF, Fyke	44	0.29
	freshwater catfish	Tandanus tandanus	Fyke	3	0.04
	longfin eel	Anguilla reinhardtii	BEF, Fyke	2	0.03
	purple-spotted gudgeon	Mogurnda adspersa	BEF	1	0.01
CU03 Total				223	1.47
SMCUS02	Agassiz's glassfish	Ambassis agassizii	BEF, Fyke	23	0.15
	Australian bass	Percalates novemaculeata	BEF	1	0.01
	Australian smelt	Retropinna semoni	BEF	66	0.96
	bony bream	Nematalosa erebi	BEF	31	0.45
	carp gudgeon	Hypseleotris spp.	BEF, Box, Fyke	97	0.64
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BEF	4	0.03
	dwarf flathead gudgeon	Philypnodon macrostomus	Fyke	9	0.06
	flyspecked hardyhead	Craterocephalus stercusmuscarum	BEF	32	0.21
	freshwater catfish	Tandanus tandanus	BEF	1	0.01
	longfin eel	Anguilla reinhardtii	BEF	1	0.01
	purple-spotted gudgeon	Mogurnda adspersa	Fyke	1	0.01
SMCUS02 Total				266	1.73
SMCUS01	Agassiz's glassfish	Ambassis agassizii	Fyke	1	0.01
	Australian smelt	Retropinna semoni	Fyke	7	0.09

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Site Common Name		Scientific Name	Method	Catch	CPUE (hr)
	carp gudgeon	Hypseleotris spp.	BPEF, Box, Fyke	310	1.78
	crimson-spotted rainbowfish	Melanotaenia duboulayi	Fyke	7	0.04
	dwarf flathead gudgeon	Philypnodon macrostomus	BPEF, Fyke	3	0.02
	flathead gudgeon	Philypnodon grandiceps	BPEF, Fyke	7	0.04
	freshwater catfish	Tandanus tandanus	Fyke	2	0.03
	longfin eel	Anguilla reinhardtii	Fyke	1	0.01
	purple-spotted gudgeon Mogurnda adspersa		BPEF, Fyke	2	0.01
SMCUS01 Total				340	1.94
SMCUS03	Agassiz's glassfish	Ambassis agassizii	Fyke	1	0.01
	Australian bass	Percalates novemaculeata	BPEF	2	0.03
	Australian smelt	Retropinna semoni	Fyke	19	0.31
	carp gudgeon	Hypseleotris spp.	BPEF, Fyke	80	0.61
	crimson-spotted rainbowfish	Melanotaenia duboulayi	BPEF, Fyke	39	0.30
	dwarf flathead gudgeon	Philypnodon macrostomus	Fyke	1	0.01
	flathead gudgeon	Philypnodon grandiceps	Fyke	1	0.01
SMCUS03 Total				143	1.09





Figure G.1 Total length of each fish caught from each assessment zone for each survey, by species.



Appendix H Detailed Turtle Survey Results

0	0:4-	August 2015		October 2015	
Species	Site	Juvenile	Adult	Juvenile	Adult
Wollumbinia latisternum	CU01	_	1	_	_
	CU02	_	1	_	4
	SMCUS01	-	2	-	3
	SMCUS02	_	1	_	_
	SMCUS03	_	3	_	1
	SMCDS02	_	_	_	1
	SMCDS04	_	3	2	_
Emydura macquarii krefftii	CU02	_	2	_	_
	SMCUS02	-	4	-	3
	SMCUS03	_	_	_	1
	US Lake	_	1	_	_
	DS Lake	_	2	_	2
Chelodina longicolis	CU03	_	1	_	1
	DS Lake	_	1	_	_
Chelodina expansa	CU01	-	-	-	1

Table H.1Raw turtle survey data.

Appendix I Summary of critical flows for MNES species in Six Mile Creek at Cooran

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Critical flow	Description	Discharge required to achieve critical flow (ML/day)	When required for MNES species	Percentage of time exceeded			
				Aug – Nov	Dec – Feb	Mar – Apr	May – Jun
10 cm above cease to flow	Stable base flow; maintain shallow riffles; maintain water levels in deep pools; maintain dissolved oxygen	0.02 – 1.0	constant	>90	_	_	_
30 cm above cease to flow	Maintain riffle-run habitats, connect habitats	0.9 – 20.0	Constant, but especially in spring and early summer for all species	42 – 64	-	_	>90
2 m above cease to flow	Stimulate dispersal in Mary cod and probably other MNES species; maintain breeding habits for MNES species	676	Twice per year, preferably not between December and January in areas where Mary River turtles nest, or between August – February for lungfish; Needed May to June, and September to February for Mary River cod	4.5 (spring) – 8.5 (late winter)	13.5	20.5	13
5 year ARI event	Geomorphic maintenance of habitats via scouring	12,729	Once every 5 years	0	0	4	3

Table I.1Summary of critical flows for MNES species in Six Mile Creek at Cooran (gauging station 138107B).

Adapted from Hydrobiology (2007)